

Oncogene

Oncogenic Effects of Evolutionarily Conserved Non-coding RNA *ECONEXIN* on Gliomagenesis

Running title: Evolutionarily conserved lncRNA *ECONEXIN*

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Abstract

Accumulating studies have demonstrated the importance of long non-coding RNAs (lncRNAs) during oncogenic transformation. However, because most lncRNAs are currently uncharacterized, the identification of novel oncogenic lncRNAs is difficult. Given that intergenic lncRNA have substantially less sequence conservation patterns than protein-coding genes across species, evolutionary conserved intergenic lncRNAs are likely to be functional. The current study identified a novel intergenic lncRNA, *LINC00461* (*ECONEXIN*) using a combined approach consisting of searching lncRNAs by evolutionary conservation and validating their expression in a glioma mouse model. *ECONEXIN* was the most highly conserved intergenic lncRNA containing 83.0% homology with the mouse orthologue (*C130071C03Rik*) for a region over 2,500 bp in length within its exon 3. Expressions of *ECONEXIN* and *C130071C03Rik* were significantly upregulated in both human and mouse glioma tissues. Moreover, the expression of *C130071C03Rik* was upregulated even in pre-cancerous conditions and markedly increased during glioma progression. Functional analysis of *ECONEXIN* in glioma cell lines, U87 and U251, showed it was dominantly located in the cytoplasm and interacted with miR-411-5p via two binding sites within *ECONEXIN*. Inhibition of *ECONEXIN* upregulated miR-411-5p together with the downregulation of its target, Topoisomerase 2 alpha (TOP2A), in glioma cell lines, resulting in decreased cell proliferation. Our data demonstrated that *ECONEXIN* is a potential oncogene that regulates TOP2A by sponging miR-411-5p in glioma. In addition, our investigative approaches to identify conserved lncRNA and their molecular characterization by validation in mouse tumor models may be useful to functionally annotate novel lncRNAs, especially cancer-associated lncRNAs.

Keywords: long non-coding RNA, conservation, glioma

Introduction

Glioma is the most common and lethal brain tumor. Although recent comprehensive molecular profiling in glioma has uncovered several mutations and the aberrant expressions of protein-coding genes, these findings are still insufficient to explain the molecular basis of gliomagenesis^{1, 2}. Because the coding genome accounts for less than 2% of all sequences, it is likely that dysregulation of non-coding RNAs might also affect tumor phenotype^{3, 4}.

Among non-coding RNAs, long non-coding RNAs (lncRNAs) are defined as transcripts >200 nucleotides in length. Recent studies have demonstrated that lncRNAs are involved in multiple biological processes including oncogenic transformation by regulating gene expression. They affect the chromatin structures and RNA interactions, such as those with the microRNA (miRNA) sponge, which upregulates protein expression by inhibiting miRNA binding to their targets^{4, 5}. It was reported that lncRNA *HOTAIR* in glioma has an oncogenic role in tumor formation by upregulating FGF1 (Fibroblast growth factor1) by sponging miR-326, which results in activation of the PI3K/AKT and MEK1/2 pathways⁶. In addition, we recently found that Notch1 activation in glioma cells specifically induced the expression of *TUG1*, which coordinately promotes self-renewal by sponging miR-145 in the cytoplasm and recruiting polycomb to repress differentiation genes by the locus-specific methylation of histone H3K27 in the nucleus⁷.

Although some lncRNAs share sequences with different protein-coding genes, many lncRNAs are located and transcribed within the intergenic regions that do not overlap exons of protein-coding genes, and are therefore termed intergenic lncRNAs⁸⁻¹⁰. Because evolutionary pressure may differently affect the degree of sequence variation between presumed non-functional regions (e.g. intergenic regions) and functional regions (e.g. protein

coding regions), most intergenic lncRNAs evolve rapidly and show poor sequence conservation^{9, 10}. However, recent studies showed that a set of intergenic lncRNAs have a high degree of sequence homology and may have important biological functions across species⁹⁻¹⁵. Indeed, a study of two highly conserved lncRNAs across vertebrates revealed that these lncRNAs have important roles in brain morphogenesis and eye development, which are retained through fishes to mammals¹³.

The current study used a pragmatic combination approach of *in silico* analysis focusing on intergenic lncRNA sequence conservation between humans and mice, and the *in vivo* analysis of a glioma mouse model (*Tp53*^{-/-}, *Nf1*^{-/-})¹⁶. We identified a novel lncRNA, *LINC00461*, termed evolutionary conserved and expressed in neural tissues (*ECONEXIN*), which has an important role in glioma formation. Our data not only demonstrate a novel axis between lncRNA and miRNA, but also provides information on a novel lncRNA during tumorigenesis.

Results

Identification of a highly conserved lncRNA, *ECONEXIN* (*LINC00461*)

First, we searched for highly conserved intergenic lncRNAs between humans and mice using a public database. The GENCODE catalog (Ver. 21) is one of the most comprehensive databases for non-coding genes with annotations (e.g. gene name, structure, and predicted biological function) in various species. Among the 7,666 human intergenic lncRNAs deposited in GENCODE, only 67 intergenic lncRNAs showed more than 80% sequence homology with mice intergenic lncRNAs in sequences longer than 200 bp (Fig. 1a, Supplementary Table 1, see Materials and Methods)¹⁷.

Among these 67 intergenic lncRNAs, *ECONEXIN* (*LINC00461*, 3,558 bp), was the most highly conserved intergenic lncRNA containing 83.0% homology with the mouse orthologue *C130071C03Rik* in a 2,521bp length within its exon 3 (Fig. 1b, c). *ECONEXIN* was also conserved among other species, including other mammals and birds, suggesting the functional importance of this lncRNA through evolution (Supplementary Fig. 1a). Intriguingly, phylogenetic tree analysis based on the sequences of *ECONEXIN* and its orthologues in each species revealed that evolution of this lncRNA reflects the proposed “timeline of evolutionary history of life” (Supplementary Fig. 1b)¹¹. In addition, *ECONEXIN* and *C130071C03Rik* expression was specific to the central nervous system in both humans and mice (Fig. 1d)¹⁸.

Increased expression of *ECONEXIN* in human gliomas

Next, we analyzed the expression of *ECONEXIN* in gliomas, the most malignant central nervous system tumor, by qPCR. *ECONEXIN* expression is upregulated in glioma tissues (n=40) compared to normal brain tissues (n=3) ($P<0.001$, Fig. 2a, Supplementary Table 2). Two glioma cell lines (U87 and U251) showed a higher expression of *ECONEXIN* compared with a normal neural stem cell line (F3) ($P<0.001$, Fig. 2b). *ECONEXIN* expression was further examined using a public database TCGA (glioma, n=530; normal, n=4), which showed a significantly higher level of *ECONEXIN* expression in glioma tissues compared to normal brain tissues ($P<0.001$, Fig. 2c). Intriguingly, lower grade glioma (grade II and III) also showed a significantly high expression level of *ECONEXIN*, suggesting that the dysregulation of *ECONEXIN* is an early event of glioma tumorigenesis ($P<0.001$, Fig. 2c).

Analysis of *C130071C03Rik* in a glioma mouse model

The use of a tumor-bearing animal model can help understand changes in lncRNA expression during tumor formation. *ECONEXIN* and *C130071C03Rik* are the most highly conserved intergenic lncRNA between humans and mice. Therefore, we examined the contribution of *C130071C03Rik* to glioma tumorigenesis using a spontaneous glioma mouse model, Mosaic Analysis with Double Markers (MADM), which precisely reflects the mosaicism of normal and cancer cells in the brain¹⁶. In the MADM model, only *Tp53* and *Nf1* homozygous deleted cells express green fluorescent protein (GFP) and these were subsequently transformed into gliomas at post-natal 90-120 days (P90-P120). We examined *C130071C03Rik* expression change in mutant cells (GFP positive cells) from pre-cancerous cells to tumor cells (P20, 60, 90, 120), which were collected by FACS sorting. *C130071C03Rik* expression was significantly increased even in pre-cancerous cells and was significantly increased in P90 and P120 cells ($P<0.01$), which correspond to tumor cells (Fig. 2d). Although *Tp53* and *Nf1* was homozygously deleted in the GFP positive cells at P6, *C130071C03Rik* expression in the GFP positive cells was comparable with the normal brain tissue cells at the same age (Fig. 2e), indicating that the *C130071C03Rik* expression was not directly regulated by *Tp53* and *Nf1*.

Inhibition of *ECONEXIN* decreases glioma cell proliferation

To examine the effects of *ECONEXIN* in glioma cell proliferation, *ECONEXIN* was depleted in U87 and U251 using five different siRNAs (siRNA#1, #2, #3, #4 and #5), which showed < 50% depletion of *ECONEXIN* expression. Depletion of *ECONEXIN* significantly suppressed glioma cell proliferation (#1–5 in U87, $P<0.05$; #1–5 in U251, $P<0.01$; Fig. 1c, 2f, 2g, Supplementary Fig. 2a, 2b). Notably, the miR-9-2 transcript overlapped with the third exon of *ECONEXIN* (Fig. 1c). However, the expression of miR-9-2 was substantially lower compared

to *ECONEXIN* expression and was not affected by siRNA against *ECONEXIN* (Fig. 1c, Fig. 2h, 2i, Supplementary Fig. 2c). These data indicated that the regulation of miR-9-2 expression might be independent of the regulation of *ECONEXIN* expression and that the suppression of proliferation in glioma cell lines by *ECONEXIN* inhibition was independent of the effect of miR-9 activities in this situation.

***ECONEXIN* is dominantly located in cytoplasm and interacts with miR-411-5p**

Next, we examined the subcellular localization of *ECONEXIN* by RNA-FISH analysis. *ECONEXIN* appeared to be dominantly located in the cytoplasm of both U87 and U251 cell lines (Fig. 3a). Almost half of the cells (49.8%) showed the cytoplasmic localization of *ECONEXIN*, while nuclear localization was observed in 23.8% of these cells ($P<0.001$, Fig. 3b).

Cytoplasmic lncRNA sometimes functions as a “sponge”, terminology used to describe the binding of lncRNA to miRNA and repress its inhibitory effect on the target mRNA¹⁹. To identify potential target miRNAs of the *ECONEXIN* sponge effect, we searched a public database, starBase v2.0. This database is uniquely designed to decode the interactions between different types of RNAs, such as mRNAs, miRNAs, and lncRNAs using large-scale cross-linking immunoprecipitation (CLIP) coupled with sequencing analysis data to avoid potential false positive predictions²⁰. Using this database, we found that miR-411-5p was the only miRNA that predictively binds to *ECONEXIN*, which contains two miR-411-5p binding sites (Fig. 1c).

The TCGA database showed that miR-411-5p was significantly downregulated in gliomas (n=526) compared to normal brain tissues (n=5) ($P<0.01$, Fig. 3c). In addition, miR-411-5p expression was significantly inversely correlated with *ECONEXIN* expression ($R=-0.261$,

$P < 0.001$) (Fig. 3d).

Consistent with this expression analysis, the depletion of *ECONEXIN* increased miR-411-5p expression in both U87 and U251 cell lines (Fig. 3e, Supplementary Fig. 3). The relationship between miR-411-5p and *ECONEXIN* expression levels were further examined by RNA-FISH analysis. Although a small number of miR-411-5p signal spots were detected in both U87 and U251 cells, they were significantly increased after *ECONEXIN* inhibition, especially in the cytoplasm (Fig. 3f). Further, we examined miR-411 expression changes in the mutant cells of MADM mouse. In contrast to *C130071C03Rik*, miR-411 expression was significantly downregulated, especially in P90 and P120 cells ($P < 0.01$, Fig. 2d, Fig. 3g).

Involvement of *ECONEXIN* and miR-411-5p in RNA-induced silencing complex

The Argonaute protein 2 (AGO2) has a central role in RNA-mediated silencing processes as a catalytic component of the RNA-induced silencing complex (RISC)²¹. To examine whether *ECONEXIN* and miR-411-5p are involved in RNA-mediated silencing, we performed an RNA immunoprecipitation (RIP) assay using antibodies against AGO2 and found that both *ECONEXIN* and miR-411-5p bound to AGO2 in U87 and U251 cells (Fig. 4a). Furthermore, U87 was transfected by expression vectors encoding Flag-AGO2 (WT), Flag-AGO2 mutated PAZ domain (PAZ-mut), or Flag-AGO2 deleted PAZ domain (PAZ-del). FLAG-AGO2 protein expression levels were analyzed by western blotting with anti-FLAG antibody (Fig. 4b). Bindings of *ECONEXIN* and miR-411-5p to AGO2 protein were impaired in mutant forms of AGO2 that had mutated or were devoid of the PAZ domain (Y311A/F312A or complete deletion), which serves as a module for miRNA transfer in RNA silencing and as an anchoring site for the 3' end of guide RNA within silencing effector complexes (Fig. 4c)^{7, 22}.

***ECONEXIN* regulates *TOP2A* by sponging miR-411-5p in gliomagenesis**

We analyzed the miR-411-5p targets using starBase V2.0 and identified 700 predicted human genes. These target genes were expected to be upregulated in human gliomas and MADM tumors during tumorigenesis. In fact, microarray analysis showed that among 700 predicted target genes, 305 genes (44%) were upregulated (>1.5) in MADM tumors compared to normal mouse brain tissues. Further, TCGA data showed that among these 305 genes, 120 genes were also significantly upregulated (>1.5) in human glioma compared to normal brain ($P<0.05$). Among these predicted target genes, topoisomerase 2a (*TOP2A*) showed the highest expression ratio not only in MADM tumors but also in human glioma compared to normal brain. (Fig. 5a, Supplementary Fig 4a, Supplementary Table 3). In addition, *Top2a* was upregulated at early tumorigenesis (P20) ($P<0.05$, Fig. 5b).

In clinical samples, *TOP2A* showed significantly higher expression in human gliomas compared to normal brains in the TCGA database (glioma $n=530$, normal $n=5$, $P<0.001$, Fig. 5c). *TOP2A* expression was positively correlated with *ECONEXIN* ($R=0.204$, $P<0.001$, Fig. 5d). Consistently, the depletion of *ECONEXIN* by siRNA repressed *TOP2A* expression (Fig. 5e, 5f, Supplementary Figure. 4b, 4c). Further, to investigate the potential relationship between *TOP2A*-3' UTR and miR-411-5p, the *TOP2A*-3' UTR, which contained a predictive miR-411-5p target site, was cloned just downstream of the firefly luciferase coding sequence (pmir-*TOP2A*-3' UTR) and transfected into the U87 and U251 cell lines. Significant repression of luciferase activities were observed in both cell lines when precursor molecules mimicking miR-411-5p was transfected ($P<0.05$, Fig. 5g, Supplementary Fig. 4a).

TOP2A appeared to have an important role in gliomagenesis. Inhibition of *TOP2A* by siRNA in U87 and U251 cell lines repressed cell growth (Fig. 5h, 5i, 5j). Interestingly, *TOP2A* overexpression did not significantly affect cell proliferation, indicating that the high expression

of TOP2A is a prerequisite for sustaining continuous tumor cell proliferation but does not promote glioma cell growth (Fig. 5l). The depletion of *ECONEXIN* significantly suppressed cell proliferation, which was rescued by ectopic TOP2A expression (Fig. 2f, Fig. 5k, 5l). These data indicate that TOP2A is an important effector to maintain glioma cell proliferation, which is regulated by *ECONEXIN*-miR-411-5p interactions via a sponge function during gliomagenesis (Fig. 5k, 5l).

Discussion

Although dysregulation of the non-coding genome derives important cancer phenotypes, most of the lncRNAs involved are currently uncharacterized. Deciphering the physiological roles of cancer-associated lncRNAs needs a transition in investigative approaches from lncRNA annotation and molecular characterization to tumorigenic mouse models^{4, 23}. In the current study, we used a public database to identify 67 intergenic lncRNAs that are highly conserved with more than 80% sequence homology between humans and mice in sequences longer than 200 bp. This criterion was also used to identify the conserved lncRNA, Tcl1 upstream neuron-associated (*TUNA*), which shows remarkable sequence conservation beyond species and which has a vital role in the pluripotency and neural differentiation of embryonic stem cells¹⁷. Intriguingly, the 67 identified lncRNAs contained *MIAT*, *KANTR*, *TUNA*, *MEG3*, and *MALAT1*, whose functions were reported to also be associated with neuronal development and function^{17, 23-26}, suggesting that this strategy for the identification of functional lncRNAs is effective and that evolutionally conserved lncRNAs sometimes have important roles in neural differentiation as was reported previously (Supplementary Table 1)¹².

We found that *ECONEXIN* was the most highly conserved among the 67 identified lncRNAs. Expression analysis of the *ECONEXIN* orthologue, *C130071C03Rik*, in a glioma

mouse model, showed that the dysregulation of *C130071C03Rik* occurred in early glioma tumorigenesis. Orthologous lncRNAs generally show similar cell type specific expression and stage specific regulation during embryonic development. Indeed, a previous study showed that human islet lncRNAs and mouse orthologous transcripts were regulated in a similar manner and that dysregulation of these lncRNAs was closely associated with type 2 diabetes²⁷. These data suggest that the integrated analysis of conservation-based transcriptomes coupled with an experimental mouse model may be an effective strategy to identify the potential roles of lncRNAs.

ECONEXIN/C130071C03Rik expression appears not to be directly regulated by *Tp53* and *Nf1*. Intriguingly, *ECONEXIN* and *C130071C03Rik* showed more than 80% sequence homology in their promoter region (~500 bp upstream of transcriptional start site). Transcriptional factor binding motif analysis using the JASPAR database revealed that 140 transcription factors may bind to the *ECONEXIN* and *C130071C03Rik* promoter regions²⁸. Among the 140 predicted transcription factors, we identified 13 transcription factors, of which expression levels were higher in both human gliomas and mouse gliomas than in normal brain tissues. These transcription factors included SOX3 and TCF12, which have been known to be involved in neural development and glioma formation as well^{29 30} (Supplementary Table 4). Therefore, it may be plausible that these transcription factors are involved in the regulation of *ECONEXIN* and *C130071C03Rik* expression.

It was considered that at least more than twice the number of lncRNAs are located in the cytoplasm compared to the nucleus³¹. Some lncRNAs in the cytoplasm act as miRNA sponges, by binding to miRNAs to quench their activity, resulting in the regulation of protein expression^{4, 7}.

It might be unlikely that *ECONEXIN* has only one target miRNA (i.e. miR-411-5p). Indeed,

using public database, miRWALK 2.0³², we found 51 miRNAs, which have possible miRNA binding sites within both *ECONEXIN* and *C130071C03Rik* sequences. Interestingly, among these 51 miRNA, expression of 42 miRNAs (82.4%) including miR-411-5p was significantly inversely correlated with *ECONEXIN* expression in gliomas (n=526) in TCGA database, suggesting that *ECONEXIN* may function as multiple miRNA sponge during glioma formation. However, in the current study, we identified miR-411-5p as an *ECONEXIN* target using more empirical database, Starbase v2.0, which is based on large-scale CLIP coupled with sequencing analysis data to avoid potential false positive predictions²⁰ and focused on the functional analysis of this miRNA during glioma formation.

ECONEXIN is dominantly located in the cytoplasm, and contains two miR-411-5p binding sites. We discovered that both *ECONEXIN* and miR-411-5p bound to AGO2 protein, a component of RISC, indicating that *ECONEXIN* is involved in the regulation of miRNA via RISC machinery. It was also proposed that the amount of cytoplasmic lncRNA should be at least more than the amount of miRNA to demonstrate miRNA sponge function³³. We found that cytoplasmic *ECONEXIN* expression was more abundant compared to miR-411-5p in glioma cells, while *ECONEXIN* depleted cells showed significantly increased levels of miR-411-5p. Thus, *ECONEXIN* modulates miR-411-5p levels through its function as a miRNA sponge to trap miR-411-5p, which results in TOP2A upregulation during glioma formation.

ECONEXIN-miR-411-5p-TOP2A axis appears to play an important role in early glioma tumorigenesis. *TOP2A* encodes DNA topoisomerase II alpha (TOP2A), an enzyme that controls DNA topologic states during DNA replication³⁴. Our data indicated that the overexpression of TOP2A is required for the maintenance of aggressive tumor cell proliferation, but its expression is not sufficient to promote tumor cell growth. Consistent with our findings, the overexpression of TOP2A was associated with an advanced stage,

aggressive tumor phenotype, and poor prognosis in various types of cancers³⁵. Based on this evidence, a DNA topoisomerase 2 inhibitor, etoposide, is widely used for the treatment of many types of cancers including glioma^{35, 36}. However, etoposide treatment sometimes causes severe adverse effects such as myelosuppression and secondary malignancies, which may be partly caused by the inhibition of DNA replication by etoposide in hematopoietic cells with proliferative activity^{37, 38}. In the current study, we showed that the inhibition of *ECONEXIN* effectively decreased TOP2A expression. Because *ECONEXIN* is specifically expressed in neural tissues, but not in blood cells, TOP2A suppression by the inhibition of *ECONEXIN* may reduce the myelosuppressive side effects observed for systemic etoposide treatments. Taken together, the current study demonstrated a novel mechanism, in which the *ECONEXIN*-miR-411-5p-TOP2A axis is an essential contributor to the acquired malignant phenotype in glioma, which may be a potential therapeutic target for glioma treatment.

Recent accumulating studies of glioma have reported a wide variety of mechanisms involved in gliomagenesis. Here, we identified a novel lncRNA associated pathway in the formation of glioma. Targeting lncRNA or its function in cancer cells provides a novel strategy for cancer therapy, because certain lncRNAs are gene specific and/or tissue epigenetic regulators^{4, 39}. Developing RNA-targeting therapeutics may be a promising option to modulate lncRNAs for anti-cancer treatments. Regarding targeting lncRNAs methods, antisense oligonucleotides (ASO) might also be a potent treatment option. ASO are chimeric RNA/DNA oligonucleotides that direct RNase H to cleave target lncRNA and deplete the transcripts regardless of their cellular localization^{4, 7, 40}. Using this type of treatment might provide more effective therapeutic options for glioma with fewer side-effects. In conclusion, our results highlight the importance of the lncRNA derived miR-411-5p-TOP2A axis in glioma formation and provide a strong rationale for targeting *ECONEXIN* as a specific and potent

therapeutic approach to treat gliomas.

Materials and Methods

Glioma tissues and cell line samples

Forty glioma tissue samples were obtained from patients undergoing surgical treatment at Nagoya University Hospital, Japan. Written informed consent was received from all patients. A diagnosis of glioma was determined histologically by experienced pathologists. The backgrounds of the patients are shown in Supplementary Table 2. Human U87 and U251 glioma cell lines were obtained from American Type Culture Collection (ATCC, Manassas, VA, USA). Although these cell lines were not authenticated, relatively low passage number cells were obtained. F3, a human neural stem cell (NSC) line, was established as described previously⁴¹. All cell lines were maintained in DMEM medium (Thermo Fisher Scientific, Waltham, MA, USA) containing 10% fetal bovine serum (Thermo Fisher Scientific) and 1% penicillin-streptomycin (Thermo Fisher Scientific) at 37°C in a humidified incubator with 5% CO₂.

Glioma mouse model and cell sorting of GFP positive cells

The MADM glioma mouse model, which is used to analyze aberrations in individual cell lineages prior to final transformation, was established and examined according to a previous study^{16, 42}. In this model, mutant cells with the homozygous deletion of *Tp53* and *Nf1* genes via Cre/loxP-mediated mitotic inter-chromosomal recombination show a green color by GFP expression, while their sibling wild type cells show a red color by red fluorescent protein (RFP). Only green colored cells spontaneously developed glioma at post-natal 90-120 days.

GFP-positive mutant cells were collected at P20, P60, P90, and P120 by sorting cells using a FACS ARIA II (BD Bioscience, San Jose, CA, USA). All experiments were conducted under protocols approved by the Institutional Animal Care and Use Committee of Nagoya City University Graduate School of Medical Sciences.

RNA extraction and quantitative reverse transcription-polymerase chain reaction (qRT-PCR)

Total RNA was extracted from tissues and cells with TRIzol reagent (Thermo Fisher Scientific). Total RNA from normal brain tissues (n=3) was purchased from BioChain (Cat # R1234043-10, Lot # B501146; Cat # R1234042-10, Lot # B810027; Cat # R1234051-50, Lot # B604039, Hayward, CA, USA). RNA was reverse-transcribed with Prime Script RT Master Mix (Takara, Kusatsu, Japan) or a TaqMan MicroRNA RT Kit (Thermo Fisher Scientific) according to the manufacturer's instructions. TaqMan PCR and SYBR Green quantitative PCR (qPCR) were carried out in triplicate for the target genes. Expression levels of target genes were determined using the delta-delta Ct (cycle threshold) method, and normalized to *GAPDH*. Expression levels of target microRNAs were normalized to RNU6B (human) or snoRNA 202 (mouse). Oligonucleotide primers for TaqMan PCR assays (Thermo Fisher Scientific) and primer sets for SYBR Green assays are shown in Supplementary Table 5.

Western blot analysis

Cell lysates were extracted from U87 and U251 cells. Fifty µg of each protein was run on 10% SDS/PAGE gels, transferred to nitrocellulose membranes, and incubated with the following antibodies as primary antibodies: rabbit polyclonal anti-TOP2A (ab12318; Abcam, Cambridge, UK), mouse monoclonal anti-FLAG (F1804; Sigma-Aldrich, St. Louis, MO, USA),

and mouse monoclonal anti- β -actin (#3700; Cell Signaling Technology, Danvers, MA). HRP-linked anti-mouse IgG (#7076, Cell Signaling Technology) and HRP-linked anti-rabbit IgG (#7074, Cell Signaling Technology) antibodies were used as secondary antibodies.

Microarray analysis

RNA from mouse tissues was amplified into cRNA and labeled according to the Agilent One-Color Microarray-Based Gene Expression Analysis protocol (Agilent Technologies, Santa Clara, CA). Labeled samples were purified with the RNeasy Kit (Qiagen, Valencia, CA, USA) and hybridized to SurePrint G3 Mouse GE 8 x 60K array slides (G4858A, Agilent Technologies) at 65°C with rotation at 10 rpm for 17 hours. The arrays were scanned using an Agilent Microarray Scanner (G2565BA, Agilent Technologies). The scanned images were analyzed using the Feature Extraction software, version 10.7.3.1 (Agilent Technologies) with background correction. Data analysis was performed with GeneSpring GX, version 12.6.0 (Silicon Genetics, Redwood City, CA, USA). Microarray analysis was performed in duplicate for wild type mice and in triplicate for the MADM model. The data from our microarrays are available in ArrayExpress (<http://www.ebi.ac.uk/arrayexpress>; Accession code: GSE91050).

RNA interference

To inhibit gene expression, U87 and U251 were transfected with 50 nM specific gene siRNA or control non-targeting siRNA (Silencer Select Negative Control #1 siRNA, 4390844, Thermo Fisher Scientific) using Lipofectamine 3000 (Thermo Fisher Scientific). Forty-eight hours after the transfection of siRNA, total RNA was extracted from U87 or U251 cell lines, and the effects were validated by qRT-PCR. siRNA sequences are shown in Supplementary Table 6.

Cell proliferation assay

U87 and U251 cells at a density of 2.5×10^3 cells per well were cultured in 96-well plates for 24 hours in normal conditions before treatments. Cell proliferation was assessed every 24 hours using a Cell Counting Kit 8 (Dojindo, Kumamoto, Japan) according to the manufacturer's instructions.

RNA Fluorescence in situ hybridization

RNA Fluorescence in situ hybridization (RNA-FISH) was performed using the QuantiGene ViewRNA ISH Cell Assay (Affymetrix, Frederick, MD, USA). Briefly, tissue sections were rehydrated and incubated with proteinase K. Subsequently, they were incubated with ViewRNA probe sets designed against human *LINC00461 (ECONEXIN)* (VA1-18109-06, Affymetrix) and miR-411-5p (VM1-99999-01, Affymetrix).

RNA binding protein immunoprecipitation assay

RIP assay was performed using RiboCluster Profiler RIP-Assay Kit (RN1005, MBL International, Watertown, MA) ⁷. RNA-protein complexes were immunoprecipitated with anti-AGO2 antibody (RN005M, MBL International), anti-Flag antibody (F1804, Sigma-Aldrich), and anti-IgG antibody as a negative control (PM035, MBL International). Immunoprecipitated RNA was analyzed by qRT-PCR. To examine the interaction between RNA and the AGO2 PAZ domain, U87 cells were transfected with expression vectors encoding either Flag-AGO2, Flag-AGO2 with mutated PAZ domain (Y311A/F312A) or Flag-AGO2 with complete deletion of PAZ domain. The details of the AGO2 vectors mutated PAZ domain were described previously⁷. Primer sequences for construction of FLAG-AGO2 variant vector were shown in

Supplementary Table 7.

Dual-Luciferase Reporter Assay

The fragments of the *TOP2A*-3' UTR region were amplified by PCR with the primers which were shown in Supplementary Table 8. The fragments of *TOP2A*-3' UTR were then ligated into the XbaI and XhoI site of the pmirGLO luciferase reporter vector (Promega, Madison, WI). For the reporter assay, luciferase constructs (100 ng) were co-transfected into the cells using Lipofectamine 3000 (Thermo Fisher Scientific) with 50nM precursor molecules mimicking miR-411-5p (ID: MC11409) (Thermo Fisher Scientific) or Pre-miR miRNA precursor negative control #1 (Thermo Fisher Scientific). Luciferase activity was measured 48 hours after transfection using a Dual-Luciferase reporter assay system (Promega). To obtain the relative luciferase activity, firefly luminescence was normalized by the Renilla luminescence.

Construction of the TOP2A expression vector

The *TOP2A* gene was amplified by PCR using KOD-plus-neo (TOYOBO, Osaka, Japan). The primer sequences are shown in Supplementary Table 8. The amplified DNA fragment was cloned into pENTR/D-TOPO vector (Thermo Fisher Scientific), followed by transferring into a Gateway pcDNA-DEST47 Vector using Gateway cloning technology (Thermo Fisher Scientific). The vector sequences were validated by conventional sequencing analysis. Transfection of DNA vectors was performed using Lipofectamine 3000 according to the manufacturer's instructions (Thermo Fisher Scientific).

Identification of highly conserved intergenic lncRNAs

Using GENCODE gene annotation, lncRNAs were classified by their location with respect to

the protein coding gene. A subtype “intergenic lncRNA, lincRNA” was defined as transcripts that are intergenic noncoding RNA loci with a length > 200 bp. Specifically, this type of lncRNA does not intersect with any protein coding loci. The boundaries of the protein coding loci were defined as 5 kb upstream of the start codon and 30 kb downstream from the stop codon⁴³. We downloaded the human and murine intergenic lncRNA sequences from GENCODE 21 and GENCODE M4 (<http://www.gencodegenes.org/>), respectively. Then, we analyzed the sequence homologies of intergenic lncRNAs between humans and mice by using the Basic Local Alignment Search Tool (BLAST) website at the National Center for Biotechnology (NCBI) (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). We sorted human intergenic lncRNAs with highly conserved regions (more than 80% sequence homology with mouse sequences longer than 200 bp). We also sorted murine intergenic lncRNAs with highly conserved regions. Finally, by overlapping each result, we identified highly conserved intergenic lncRNAs between humans and mice.

Database and phylogenetic analysis

To analyze gene expression (*ECONEXIN*, hsa-miR-411-5p and *TOP2A*) in clinical glioma samples, we used the level-3 preprocessed expression data of RNA-seq (IlluminaHiseq) from The Cancer Genome Atlas (TCGA) (<https://tcga-data.nci.nih.gov/>). Phylogenetic trees were constructed using MEGA 6.0 software⁴⁴ with the neighbor joining method. The RNA sequences in each species, which are orthologues of *LINC00461*, were obtained from the BLAST database.

Statistics

The statistical significance of differences between two groups was analyzed by the paired Student's *t*-test. All reported *P* values were two-sided, and $P < 0.05$ was considered statistically significant.

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Figure legends

Figure 1. Identification of a highly conserved intergenic lncRNA, *LINC00461*, *ECONEXIN*. (a) Pipeline for the identification of highly conserved intergenic lncRNAs between humans and mice. Annotations of intergenic lncRNAs were obtained from the GENCODE database. (b) Length of the conserved region (homology >80% between humans and mice, X-axis) of 67 highly conserved intergenic lncRNAs. (c) Upper panel shows a schematic diagram of the *LINC00461* (*ECONEXIN*) gene locus as annotated on the UCSC Genome Browser. Pre-miR-9-2 (horizontal stripes) is overlapped with the third exon of *LINC00461*. Lower panel shows the transcribed RNA of *LINC00461*. Black arrow heads show siRNA target sites against *LINC00461* (#1-5) used in this study. Gray arrows show predicted binding sites of miR-411-5p in *LINC00461*. (d) Expression level of *ECONEXIN* in 53 different human tissues. Analyzed data were obtained from GTEx (Genotype Tissue Expression) (version V6p, <http://www.gtexportal.org/home/>). X-axis indicates tissue names. The examined numbers of samples are shown after tissue names in parentheses. Y-axis indicates the median RPKM (Reads per kilobase of transcript per million mapped reads) value.

Figure 2. Inhibition of *ECONEXIN* (si-*ECONEXIN* #1-3) decreases glioma cell proliferation. (a) Expression levels of *ECONEXIN* in clinical normal brain (cerebral cortex, n=3) and glioma (n=40) samples were examined by qRT-PCR. Bars indicate median. ***, $P<0.001$. (b) Expression levels of *ECONEXIN* in normal neural stem cell (F3) and glioma cell lines (U87 and U251). Y-axis indicates expression level relative to F3 (\log_2). Experiments were performed in triplicate. Error bars indicate S.D (standard deviation). ***, $P<0.001$.

(c) Expression levels of *ECONEXIN* in clinical normal brain (n=4), lower grade glioma (LGG,

n=530), and glioblastoma (GBM, n=151) samples are presented as a box-whisker plot. Data were obtained from the TCGA database. Middle horizontal line inside box indicates the median. Bottom and top of the box indicates the 25th and 75th percentiles, respectively. The ends of the whiskers indicate the minimum and maximum of all of the data, respectively. Y-axis indicates expression levels of *ECONEXIN* by RPKM value +1 (\log_2). ***, $P<0.001$ (d) Expression levels of *C130071C03Rik* in normal brain (gray bars) and glioma cells (black bars) in MADM mice. X-axis indicates post-natal days. Y-axis indicates expression level normalized to *Gapdh*. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$, ***, $P<0.001$. (e) Expression levels of *C130071C03Rik* in normal brain tissues (gray bar) and pre-cancerous GFP positive cells (*Tp53*^{-/-}, *Nf1*^{-/-}; black bar) in MADM mice at postnatal 6 days. Y-axis indicates expression level normalized to *Gapdh*. Experiments were performed in triplicate. N.S, not significant. (f) Expression levels of *ECONEXIN* in U87 (left) and U251 (right) cells treated with si-negative control (si-NC), si-*ECONEXIN* #1, #2 or #3. Y-axis indicates expression level relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$. (g) Effects of si-*ECONEXIN* #1, #2 and #3 on proliferation of U87 (left) and U251 cells (right). X-axis indicates time after each siRNA treatment. Y-axis indicates cell number relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$. (h) Expression levels of *ECONEXIN* and miR-9-2 in U87 (left) and U251 cells (right). Y-axis indicates expression level relative to *ECONEXIN*. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$. (i) Expression levels of miR-9-2 in U87 (left) and U251 cells (right) treated with si-NC, si- *ECONEXIN* #1, #2 or #3. Y-axis indicates expression level relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D (n=3). N.S, not significant

Figure 3. Predominant location of *ECONEXIN* in cytoplasm. (a) Left panel shows the RNA-FISH analysis of *ECONEXIN* (red) in U87 (left) and U251 cells (right). Nuclei are stained with DAPI. Bars: 10 μ m. Right graph shows the spot numbers related to *ECONEXIN* detection per cell in U87 and U251 cells. Error bars indicate S.D (n=10). (b) Three patterns of subcellular localization of *ECONEXIN* in U251 cells (left). Type 1 and 3 show dominant *ECONEXIN* localization in either the nucleus or cytoplasm, respectively. Type 2 shows comparable localization of *ECONEXIN* in the nucleus and cytoplasm. Numbers of the three types of cells were calculated per 100 cells (right). Cell counting was performed four times. Bar indicates the median. **, $P<0.01$. (c) Expression levels of miR-411-5p in clinical normal brain and glioma samples are presented as a box-whisker plot. Data were obtained from TCGA data sets. Middle horizontal line inside box indicates the median. Bottom and top of the box indicates the 25th and 75th percentiles, respectively. The ends of the whiskers indicate the minimum and maximum of all of the data, respectively. Y-axis indicates expression levels of miR-411-5p by RPM (reads per million) miRNA mapped values (\log_2) in either normal brain (n=5) or glioma tissues (n=526). **, $P<0.01$. (d) Relationship of expression levels between *ECONEXIN* and miR-411-5p in glioma samples (n=526) from the TCGA database. X-axis indicates expression levels of *ECONEXIN*. Y-axis indicates expression levels of miR-411-5p. P values were obtained by Pearson's correlation coefficient analysis. The red line in the graph indicates linear approximation ($R=-0.261$). (e) Expression levels of miR-411-5p in U87 and U251 cells treated with si-NC, si-*ECONEXIN* #1, #2 or #3. Y-axis indicates expression level relative to siRNA-negative control (si-NC). Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$. (f) Left panel shows RNA-FISH analysis of miR-411-5p (red) in U87 (upper) and U251 cells (lower) treated with si-NC (left)

or si-*ECONEXIN* (right). Nuclei are stained with DAPI. Bars: 10 μ m. Right graph shows spot numbers related to miR-411-5p detection per cell in U87 (left) and U251 (right) cells treated with either si-NC or si-*ECONEXIN*. Error bars indicate S.D (n=3). ***, $P<0.001$. (g) Expression levels of mmu-miR-411 in normal brain (gray bars) and glioma cells (black bars) in MADM mice. X-axis indicates post-natal days. Y-axis indicates expression level normalized to *snoRNA202*. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$, ***, $P<0.001$.

Figure 4. Involvement of *ECONEXIN* and miR-411-5p in RISC. (a) RIP analysis of *ECONEXIN* and miR-411-5p in U87 (left) and U251 cells (right) was performed using an antibody against AGO2 or IgG as a negative control. Enrichment of *ECONEXIN* and miR-411-5p are expressed as a percentage of input RNA. Y-axis indicates normalized fold enrichment to IgG. Experiments were performed in triplicate. Error bars indicate S.D. ***, $P<0.001$. (b) Schematic diagram shows construction of the FLAG-AGO2 vector. The PAZ domain was mutated as Y311A/F312A (PAZ-mut) or completely deleted (229-347, PAZ-del) (upper). FLAG-AGO2 protein expression levels were analyzed by western blotting with anti-FLAG antibody in U87 cells. WT, wild type FLAG-AGO2. β -actin protein expression was used as an internal control. (Lower) (c) RIP analysis of *ECONEXIN* and miR-411-5p was performed using an antibody against Flag (black bar) or IgG (gray bar) as a negative control. Enrichment of *ECONEXIN* and miR-411-5p is expressed as a percentage of input RNA. Y-axis indicates normalized fold enrichment to IgG. Experiments were performed in triplicate. Error bars indicate S.D. **, $P<0.01$, ***, $P<0.001$.

Figure 5. *ECONEXIN* regulates DNA replication by sponging miR-411-5p in

gliomagenesis. (a) Schematic diagram to identify the target of miR-411-5p. Human miR-411-5p target genes were predicted by starBase V2.0. Among the 700 genes, *Top2a* showed the highest expression ratio between both human glioma tissues and normal brain tissues, and MADM glioma cells and normal mouse brain cells. (b) Expression levels of *Top2a* in normal mouse brain cells (gray bars) and tumor cells in MADM (black bars). X-axis indicates post-natal days. Y-axis indicates expression level relative to *Gapdh*. Experiments were performed in triplicate. Error bars indicate S.D (n=3). **, $P<0.01$, ***, $P<0.001$. (c) Expression levels of *TOP2A* in clinical normal brain and glioma samples are presented as a box-whisker plot. Data were obtained from TCGA data sets. Middle horizontal line inside box indicates the median. Bottom and top of the box indicates the 25th and 75th percentiles, respectively. The ends of the whiskers indicate the minimum and maximum of all of the data, respectively. Y-axis indicates expression levels of *TOP2A* by RSEM (RNA-Seq by Expectation-Maximization) value plus 1 (\log_2) in normal brain (n=5) and glioma (n=530) tissues. ***, $P<0.001$. (d) Relationship between the expression levels of *ECONEXIN* and *TOP2A* in glioma samples (n=530) from the TCGA database. X-axis and Y-axis indicate expression levels of *ECONEXIN* and *TOP2A*, respectively. Data were analyzed by Pearson's correlation coefficient analysis. The red line in the graph indicates linear approximation ($R=0.204$). (e) Expression level of *TOP2A* in U87 (left) and U251 cells (right) treated with si-NC, si-*ECONEXIN* #1, #2 or #3. Y-axis indicates expression level relative to siRNA-negative control (si-NC). Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$. (f) Protein expression levels of *TOP2A* in U87 (left) and U251 cells (right) treated with si-NC, si-*ECONEXIN* #1, #2 or #3. (g) Dual-Luciferase assay with the pmir-*TOP2A*-3' UTR reporter vector in U87 and U251 cells. Cells were co-transfected with 50nM precursor molecules pre-miR-411-5p (Pre-411-5p) or negative control miRNA

precursor (Pre-NC). Y-axis indicates relative luciferase activity to pmir-emp control (i.e. reporter vector without *TOP2A*-3' UTR sequence). The assays were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$. (h) Expression levels of *TOP2A* in U87 and U251 cells treated with si-NC or si-*TOP2A*. Y-axis indicates expression level relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D. **, $P<0.01$, $P<0.001$. (i) Protein expression levels of *TOP2A* in U87 and U251 cells treated with si-NC or si-*TOP2A*. (j) Effects of si-*TOP2A* on the proliferation of U87 and U251 cells. X-axis indicates time after treatment of si-RNA against each target. Y-axis indicates cell number relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D (n=3). *, $P<0.05$, **, $P<0.01$, ***, $P<0.001$. (k) U87 and U251 cells were transfected with si-NC or si-*ECONEXIN* and with empty vector or expression vectors encoding *TOP2A*. Protein expression levels of *TOP2A* in U87 (left) and U251 cells (right) after 72 hours of transfection were analyzed by western blotting with anti-*TOP2A* antibody. β -actin was used as an internal control. (l) Effects of ectopic *TOP2A* expression on cell proliferation in *ECONEXIN* depleted U87 (left) and U251 (right) cells. X-axis indicates time after each treatment. Y-axis indicates cell number relative to si-NC plus empty vector. Experiments were performed in triplicate. Error bars indicate S.D. ***, $P<0.001$.

Figure 1

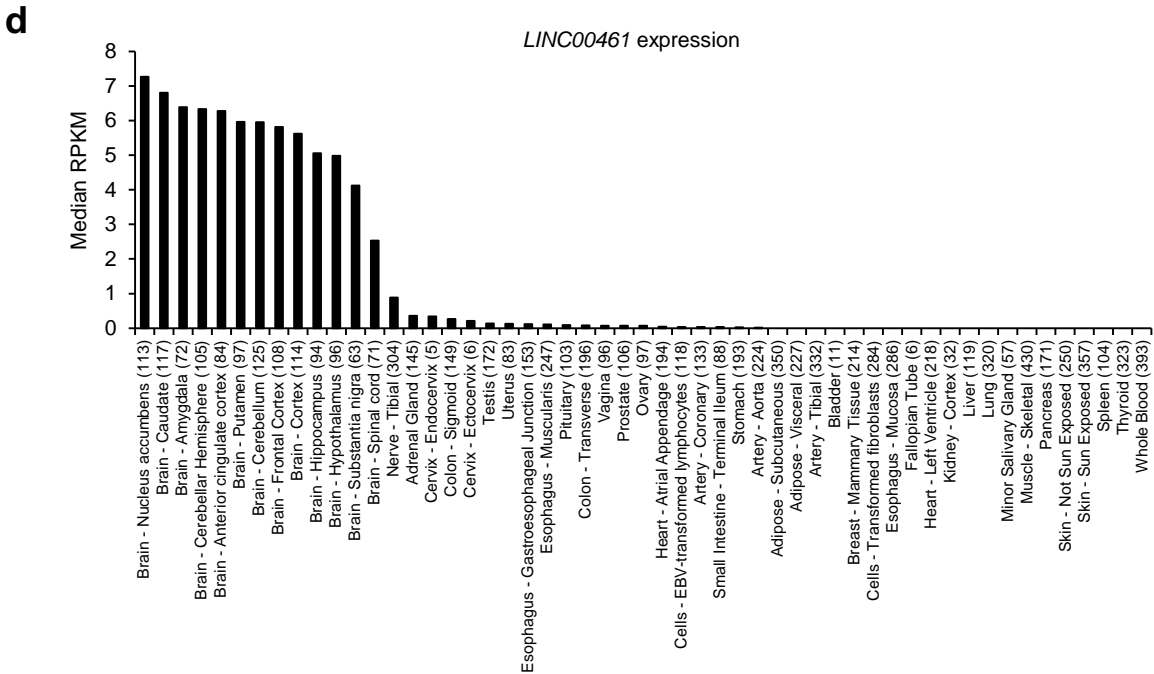
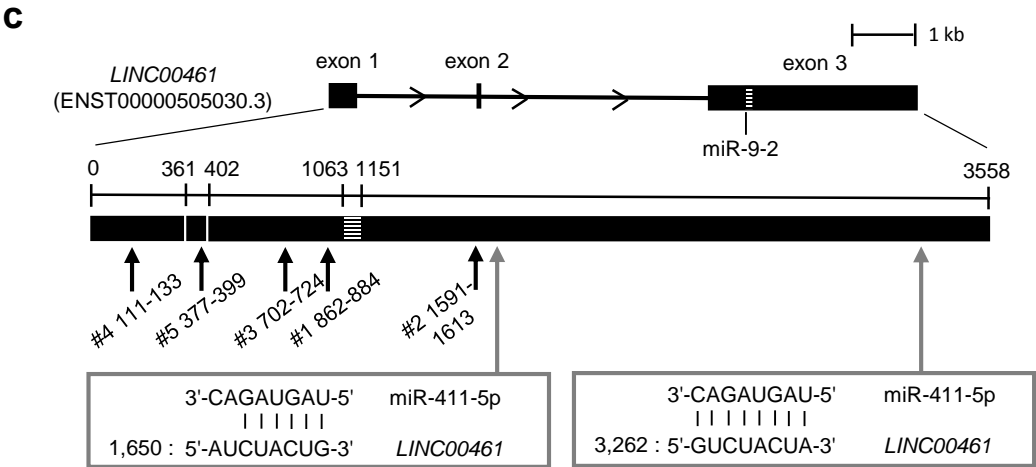
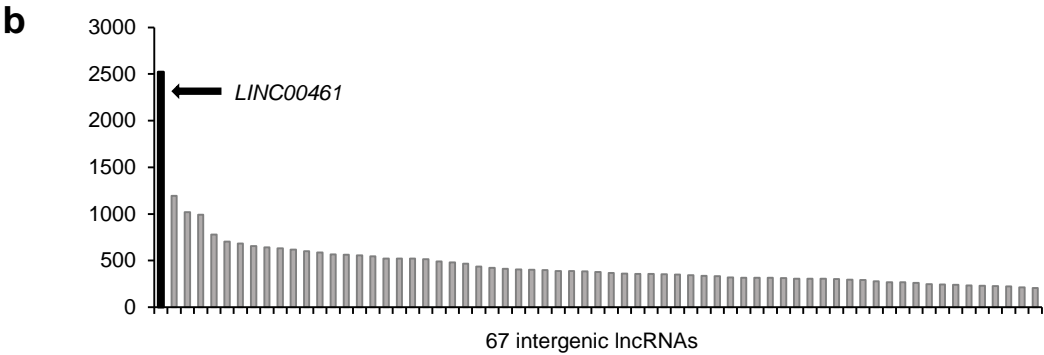
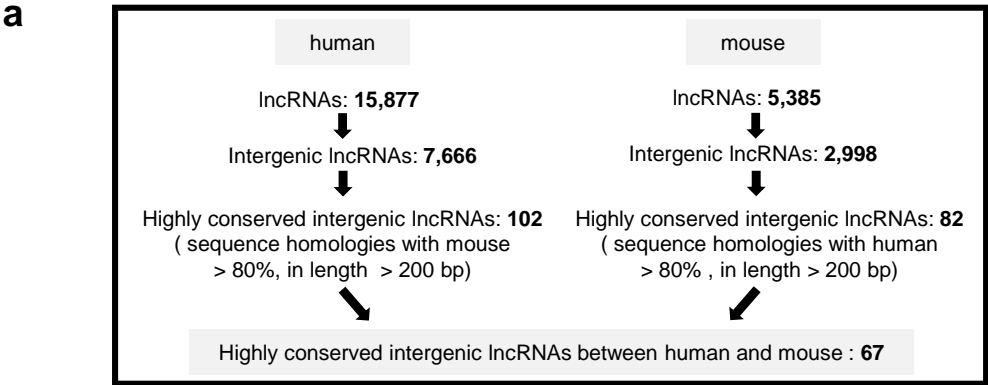


Figure 2

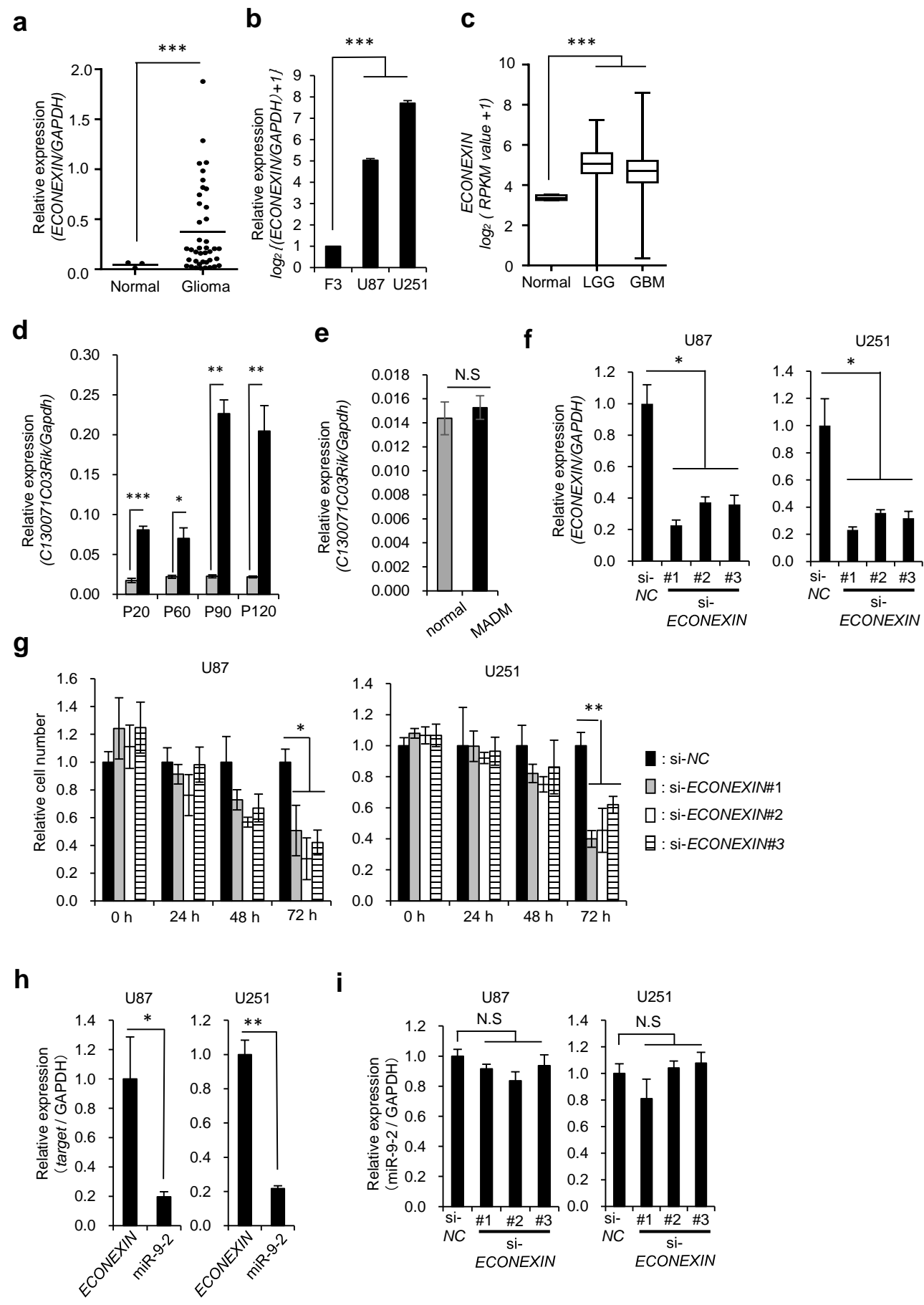


Figure 3

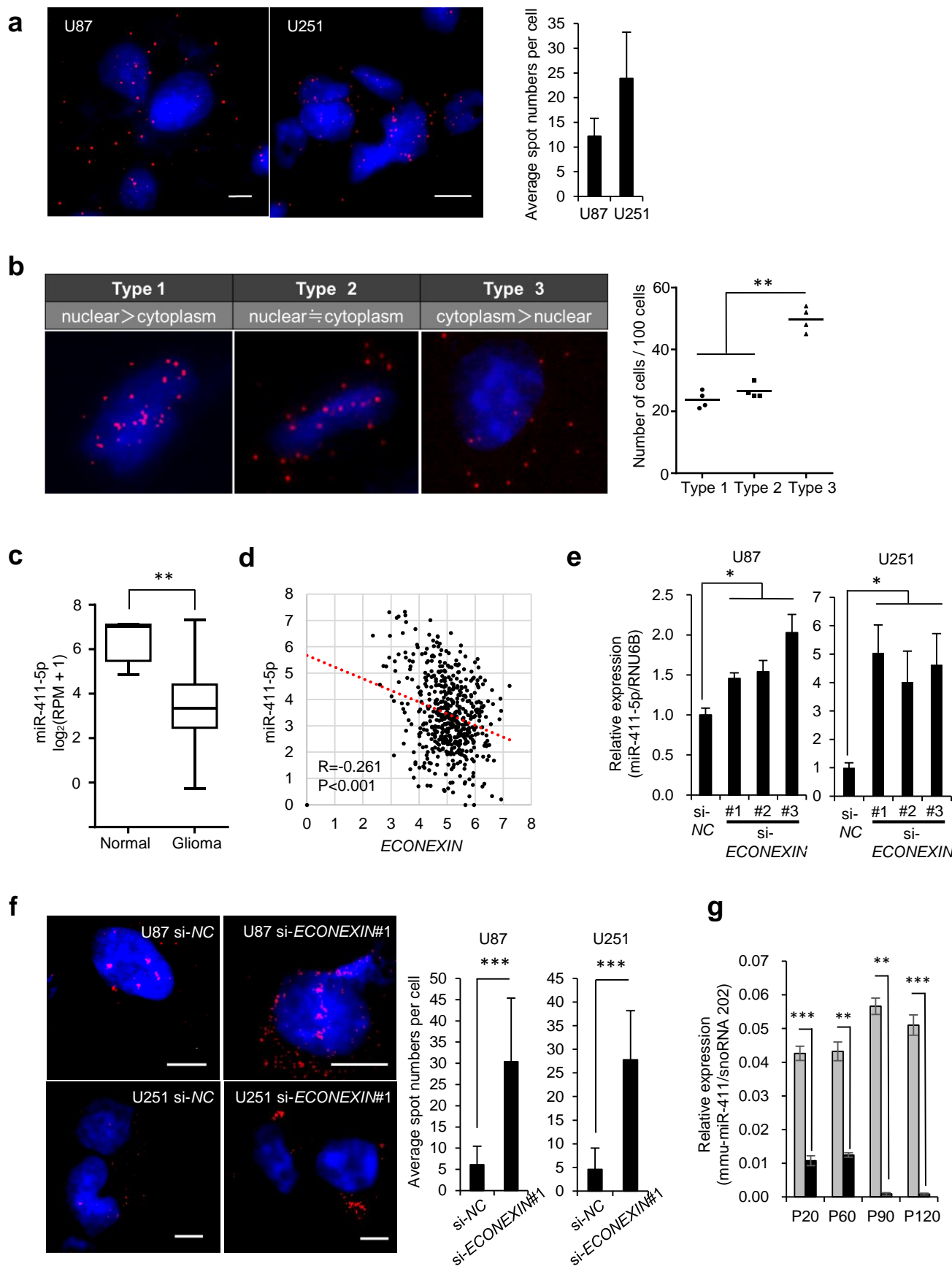
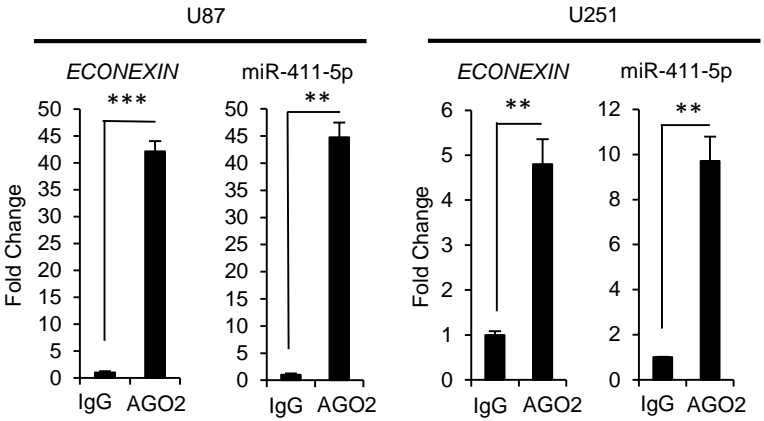
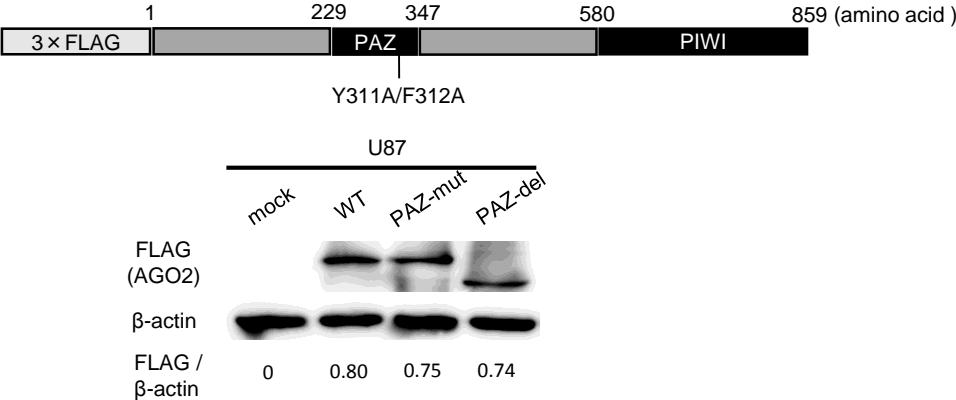


Figure 4

a



b



c

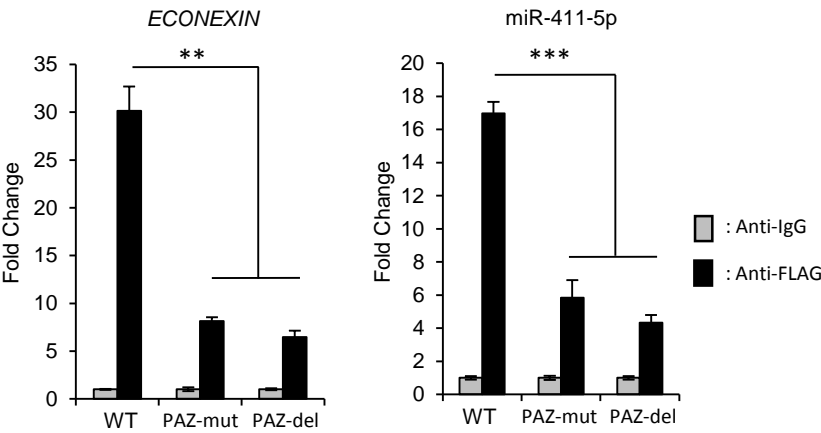
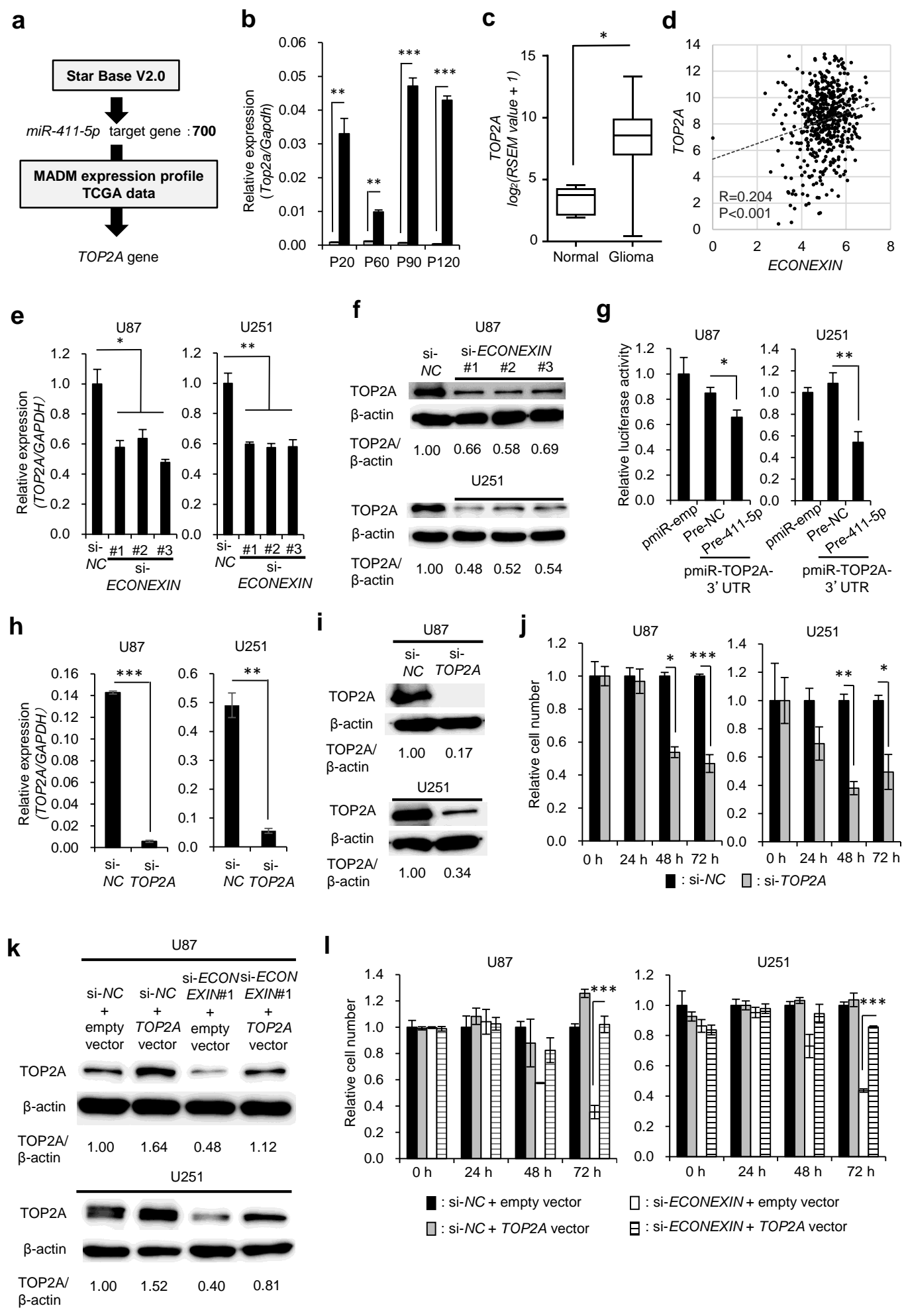


Figure 5

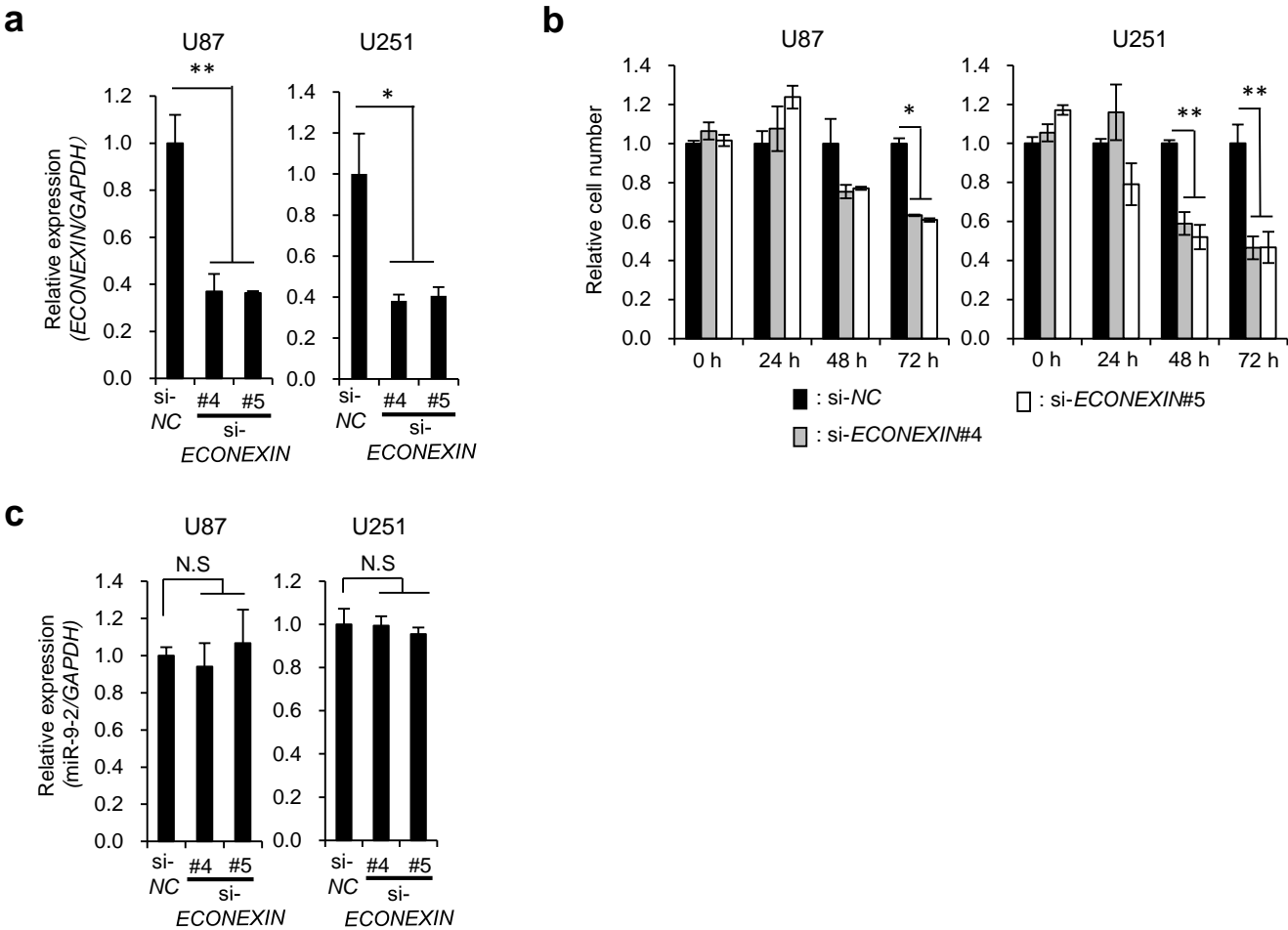


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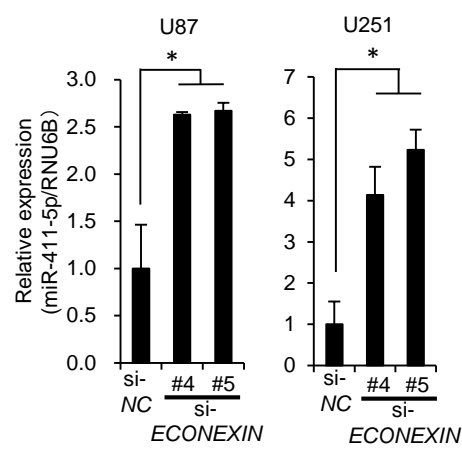
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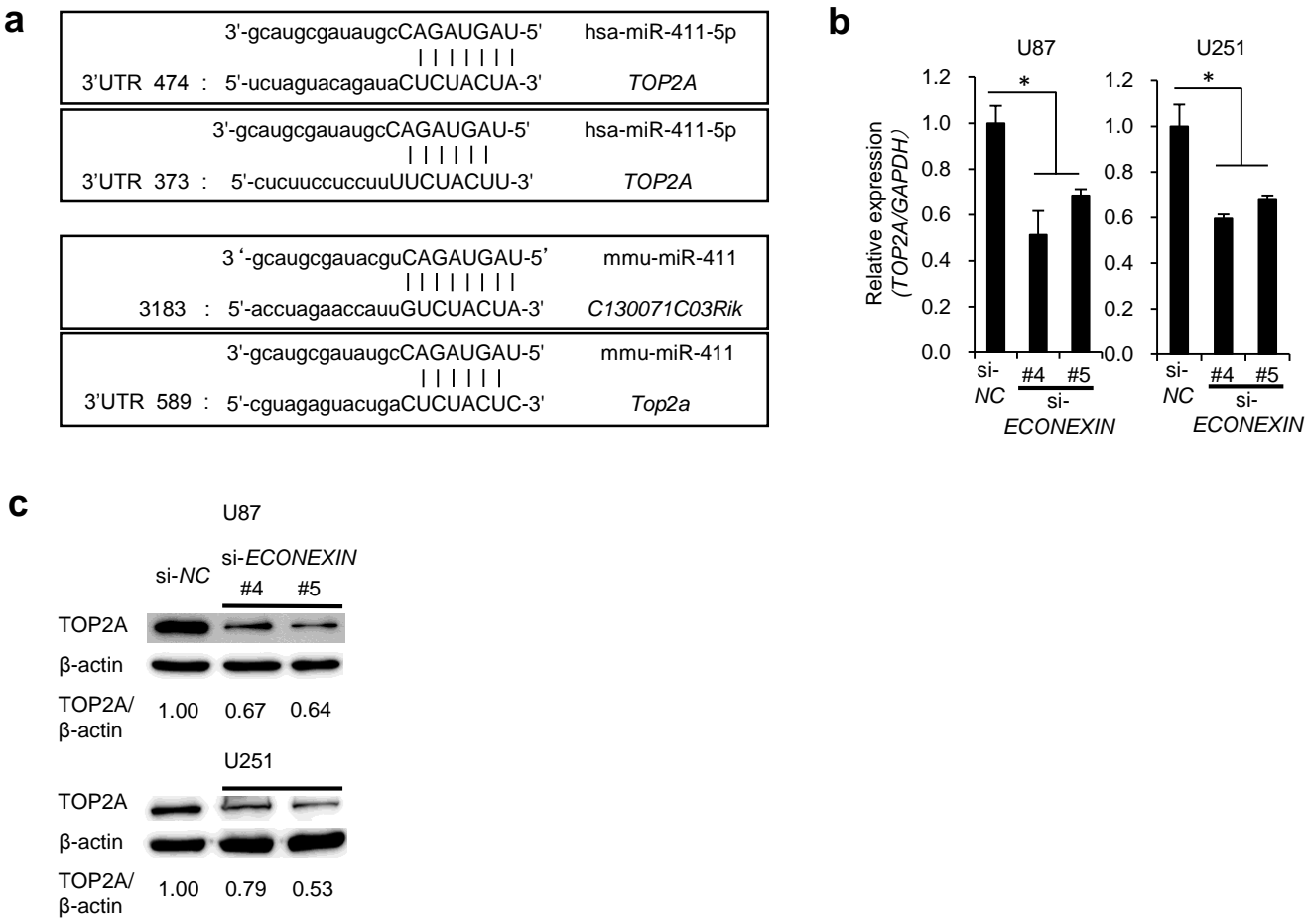
Supplementary Figure 2



Supplementary Figure 3



Supplementary Figure 4



Supplementary Figure legends

Supplementary Figure 1. Characteristics of *ECONEXIN*. (a) Sequence conservation of *LINC00461* between humans and 9 other species. White circle indicates sequence homology >80% in a length >200 bp within the *ECONEXIN* homologues. (b) Phylogenetic tree of *ECONEXIN* homologues from each species. Scale bar represents evolutionary distance (0.05 indicates 5 substitutions per 100 bps).

Supplementary Figure 2. Inhibition of *ECONEXIN* (si-*ECONEXIN* #4.5) decreases glioma cell proliferation. (a) Expression levels of *ECONEXIN* in U87 (left) and U251 cells (right) treated with si-negative control (si-NC), si-*ECONEXIN* #4, or #5. Y-axis indicates expression level relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$. (b) Effects of si-*ECONEXIN* #4 and #5 on cell proliferation of U87 (left) and U251 cells (right). X-axis indicates time after transfection of si-RNA. Y-axis indicates cell number relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D. *, $P<0.05$, **, $P<0.01$. (c) Expression level of *miR-9-2* in U87 (left) and U251 cells (right) treated with si-NC, si-*ECONEXIN* #4 or #5. Y-axis indicates expression level relative to si-NC. Experiments were performed in triplicate. Error bars indicate S.D. N.S, not significant

Supplementary Figure 3. Inhibition of *ECONEXIN* (si-*ECONEXIN* #4.5) increases miR-411-5p expression. Expression levels of miR-411-5p in U87 (left) and U251 cells (right) treated with si-NC, si-*ECONEXIN* #4 or #5. Y-axis indicates expression level relative to siRNA-negative control (si-NC). Experiments were performed in triplicate. Error bars indicate

S.D. *, $P < 0.05$.

Supplementary Figure 4. Inhibition of *ECONEXIN* (si-*ECONEXIN* #4.5) decreases *TOP2A* expression. (a) Upper panels indicates predicted binding sites of miR-411-5p in *Top2a*. Lower panels indicates predicted binding sites of mmu-miR-411 in *C130071C03Rik* and *Top2a*. (b) Expression levels of *TOP2A* in U87 and U251 cells treated with si-NC, si-*ECONEXIN* #4 or #5. Y-axis indicates expression level relative to siRNA-negative control (si-NC). Experiments were performed in triplicate. Error bars indicate S.D. *, $P < 0.05$. (c) Protein expression levels of *TOP2A* in U87 and U251 cells treated with si-NC, si-*ECONEXIN* #4 or #5.

Supplementary Table 1. Highly conserved intergenic lncRNAs between humans and mice

Gene Name	Transcript ID	Length of Conserved region (bp)	Sequence homology (%)
LINC00461	ENST00000505030.3	2521	82.98
LINC01142	ENST00000440404.5	1193	82.48
RP11-458K10.2	ENST00000470532.1	1019	91.17
MALAT1	ENST00000534336.1	993	81.97
AC004158.2	ENST00000561611.2	779	93.32
MIR137HG	ENST00000424528.2	704	87.5
CTC-529L17.2	ENST00000503489.1	683	84.33
LINC00925	ENST00000536780.4	655	85.5
RP11-681H18.2	ENST00000612252.1	642	83.64
MIR503HG	ENST00000440570.3	633	80.25
LINC00643	ENST00000334389.5	617	89.79
CTD-2249K22.1	ENST00000512859.1	601	98.5
MEG3	ENST00000455531.1	588	83.16
LINC00371	ENST00000563710.3	567	81.83
AC007383.3	ENST00000420509.1	562	87.19
AC007392.3	ENST00000435389.3	556	87.41
RP11-345F18.1	ENST00000504250.1	545	82.94
RP3-418C23.2	ENST00000574739.1	523	88.15
RP11-347I19.7	ENST00000613093.1	522	91.38
AC096649.3	ENST00000438963.1	520	91.73
RP11-340E6.1	ENST00000474979.3	516	83.72
RP11-545H22.1	ENST00000512628.1	489	80.37
RP11-107I14.5	ENST00000450669.2	482	85.27
RP11-204E9.1	ENST00000447728.1	466	85.41
AC093901.1	ENST00000449075.1	437	96.8
RP1-30E17.2	ENST00000449485.1	423	83.22
RP4-665J23.2	ENST00000425185.1	411	90.02
LINC00890	ENST00000569275.1	405	88.89
RP11-308N19.3	ENST00000446206.1	400	80.5
LINC00867	ENST00000445161.3	399	81.45
RP11-684B21.1	ENST00000560056.1	389	85.09
RP11-397G17.1	ENST00000415106.1	388	84.54

<i>RP11-76I14.1</i>	ENST00000545549.1	385	80.26
<i>LINC01109</i>	ENST00000603837.1	379	92.88
<i>RP4-665J23.1</i>	ENST00000606660.1	368	83.97
<i>LINC01412</i>	ENST00000413525.1	360	81.67
<i>AC103564.7</i>	ENST00000437330.1	358	83.8
<i>MEG9</i>	ENST00000429368.1	356	87.08
<i>LINC01505</i>	ENST00000444985.4	353	96.6
<i>RP11-2L8.1</i>	ENST00000450500.1	351	83.48
<i>FLJ37035</i>	ENST00000527483.3	342	83.92
<i>RP1-104O17.1</i>	ENST00000433637.1	337	88.13
<i>RP11-507B12.1</i>	ENST00000559590.1	333	87.69
<i>RP11-159D12.2</i>	ENST00000577267.1	320	80.94
<i>KANTR</i>	ENST00000605526.3	316	84.49
<i>LINC01355</i>	ENST00000566551.1	316	80.06
<i>LINC00599</i>	ENST00000521863.1	314	80.25
<i>RP11-472K22.2</i>	ENST00000512268.1	311	88.42
<i>AC073094.4</i>	ENST00000453798.1	305	95.41
<i>RP11-343J18.2</i>	ENST00000441473.1	305	87.21
<i>RP11-60A8.2</i>	ENST00000511174.1	305	81.64
<i>RP11-271F18.1</i>	ENST00000455871.1	301	87.04
<i>CTD-2012K14.6</i>	ENST00000605277.1	294	82.99
<i>RP11-1008C21.2</i>	ENST00000559232.1	293	89.08
<i>RP11-802H3.2</i>	ENST00000515263.1	279	89.61
<i>RP4-535B20.1</i>	ENST00000448344.1	268	83.96
<i>RP11-107I14.2</i>	ENST00000442188.1	267	80.52
<i>RP4-738P15.1</i>	ENST00000438287.3	261	80.08
<i>LINC01315</i>	ENST00000412060.1	246	85.37
<i>OTX2-AS1</i>	ENST00000534909.2	242	88.43
<i>RP11-290M5.4</i>	ENST00000609367.1	241	95.44
<i>LINC00354</i>	ENST00000412119.1	233	84.12
<i>TUNAR</i>	ENST00000554321.1	230	84.78
<i>MIAT</i>	ENST00000620145.2	227	87.22
<i>RP11-266O8.1</i>	ENST00000556519.2	222	84.68
<i>C1orf132</i>	ENST00000608023.3	212	90.57
<i>RP11-4B14.3</i>	ENST00000482559.1	205	93.17

Supplementary Table 2. Backgrounds of glioma patients

	Grade II	Grade III	Grade IV
Sample number	12	4	24
Mean age (years old)	45.5	44.3	53.9
Male (%)	75	50	79.2
Female (%)	25	50	20.8

Age (years old)	Gender	WHO grade
82	M	IV
82	M	IV
78	F	IV
78	M	IV
77	M	IV
76	M	IV
67	M	IV
67	F	IV
66	F	IV
65	M	IV
59	M	IV
58	M	IV
57	M	IV
56	M	IV
47	M	IV
42	M	IV
40	M	IV
36	M	IV
36	M	IV
35	M	IV
29	M	IV
28	F	IV
23	F	IV
10	M	IV
59	M	III
55	F	III
33	M	III

30	F	III
73	M	II
58	M	II
52	M	II
50	M	II
50	M	II
48	F	II
44	M	II
39	F	II
37	F	II
35	M	II
31	M	II
29	M	II

Supplementary Table 3. Expressions of 700 predicted miR-411-5p target genes in MADM tumors (n=3) and normal mouse brains (n=2)

Gene name	Average of gene expression in MADM (log ₂)	Average of gene expression in normal (log ₂)	Expression ratio (MADM/normal) (log ₂)
<i>TOP2A</i>	11.19493767	4.1605284	7.034409267
<i>CKAP2</i>	9.344965	2.8856164	6.4593486
<i>PBK</i>	10.9954075	5.077962	5.9174455
<i>CDCA5</i>	10.35888067	4.471646	5.887234667
<i>CDK1</i>	7.818162667	2.605294	5.212868667
<i>KIF4A</i>	9.299170167	4.447746	4.851424167
<i>OASL</i>	13.14041717	8.305103	4.835314167
<i>TUBB6</i>	12.48201433	8.08913975	4.392874583
<i>CASC5</i>	6.6426148	2.3809836	4.2616312
<i>C6orf145</i>	6.4358621	2.2560105	4.1798516
<i>PXDC1</i>	6.4358621	2.2560105	4.1798516
<i>VCAN</i>	11.70413	7.58733375	4.11679625
<i>TFPI</i>	11.254461	7.33970015	3.91476085
<i>CENPK</i>	10.30970867	6.5436695	3.766039167
<i>MAP3K1</i>	14.08929433	10.3624095	3.726884833
<i>SOX4</i>	13.07506633	9.5672005	3.507865833
<i>MTBP</i>	11.89828167	8.4142215	3.484060167
<i>TFPI2</i>	5.558894067	2.08898845	3.469905617
<i>FAM111A</i>	9.251714333	5.937359	3.314355333
<i>SCML2</i>	9.261733333	6.01137805	3.250355283
<i>SAMD9</i>	11.32274	8.083223	3.239517
<i>PCDHB2</i>	9.536795667	6.4009826	3.135813067
<i>PRKD3</i>	12.46737933	9.3891455	3.078233833
<i>TRAF3IP2</i>	7.713111767	4.68371125	3.029400517
<i>DRAM1</i>	7.7684438	4.74601475	3.02242905
<i>TUBB2B</i>	16.07383433	13.0775175	2.996316833
<i>IER3</i>	11.78588517	8.815164	2.970721167
<i>ISOC1</i>	12.00272967	9.05297	2.949759667
<i>RAD51</i>	9.228243667	6.3357129	2.892530767
<i>CASP8</i>	10.62461833	7.7509513	2.873667033
<i>ICAM1</i>	7.427212233	4.5645805	2.862631733
<i>LGALS3</i>	11.42582633	8.58668625	2.839140083
<i>RCC2</i>	12.795094	9.9560645	2.8390295
<i>RFC3</i>	9.916803667	7.079398	2.837405667
<i>SRPX</i>	7.7978711	4.967217	2.8306541
<i>ZNF184</i>	10.622722	7.87212895	2.75059305
<i>POU4F1</i>	11.172652	8.44396525	2.72868675
<i>SOX11</i>	11.44767867	8.719468	2.728210667
<i>SHMT1</i>	8.1071195	5.44972785	2.65739165
<i>ADAM12</i>	7.595619833	4.94505685	2.650562983
<i>AIF1L</i>	12.90668267	10.2563715	2.650311167
<i>TNPO1</i>	12.04867533	9.5331385	2.515536833

<i>ACTL6A</i>	11.72275033	9.2198525	2.502897833
<i>SKAP2</i>	12.618853	10.1325055	2.4863475
<i>HERC5</i>	7.992234333	5.57534775	2.416886583
<i>PALLD</i>	10.92391	8.5547745	2.3691355
<i>TMTC2</i>	12.34435233	9.975565	2.368787333
<i>LRRC17</i>	4.6272435	2.2704331	2.3568104
<i>LPP</i>	9.409155667	7.11366615	2.295489517
<i>PAWR</i>	8.440575333	6.174366	2.266209333
<i>C13orf27</i>	10.52838633	8.3199965	2.208389833
<i>HMGNA4</i>	13.92341267	11.715461	2.207951667
<i>DDX21</i>	10.7735605	8.5841425	2.189418
<i>MLF1IP</i>	7.723280867	5.58360195	2.139678917
<i>MTHFD2</i>	9.92036	7.8007365	2.1196235
<i>PUS7</i>	12.468984	10.3599125	2.1090715
<i>PPP4R1</i>	14.09485683	11.99153	2.103326833
<i>ZNF146</i>	11.66878233	9.5711815	2.097600833
<i>POLD1</i>	11.732299	9.645311	2.086988
<i>CYBRD1</i>	8.0838369	6.0056215	2.0782154
<i>SS18</i>	14.11558233	12.055675	2.059907333
<i>ZNF655</i>	10.53922733	8.561806	1.977421333
<i>NHSL1</i>	12.12592733	10.1576835	1.968243833
<i>HCLS1</i>	11.635642	9.6890325	1.9466095
<i>CROT</i>	12.26801367	10.3379355	1.930078167
<i>ARHGAP18</i>	9.548047	7.63547885	1.91256815
<i>EYA1</i>	15.0297465	13.170573	1.8591735
<i>LARS</i>	13.30717633	11.4789	1.828276333
<i>GPR56</i>	13.33601267	11.5106575	1.825355167
<i>SP1</i>	11.764234	9.964281	1.799953
<i>ARL6IP6</i>	13.8575035	12.0607835	1.79672
<i>ZNF800</i>	8.382414667	6.5957147	1.786699967
<i>DTL</i>	9.110019667	7.3479525	1.762067167
<i>C1orf144</i>	10.52352733	8.7695215	1.754005833
<i>PDE3B</i>	10.28752833	8.536499	1.751029333
<i>SLC7A6</i>	8.604131333	6.87858535	1.725545983
<i>ROD1</i>	9.059855	7.351861	1.707994
<i>MTHFD1L</i>	11.36663167	9.659465	1.707166667
<i>C16orf80</i>	13.760375	12.0605075	1.6998675
<i>POLR2D</i>	11.38461833	9.687041	1.697577333
<i>NRF1</i>	9.831534667	8.134284	1.697250667
<i>PRMT6</i>	9.042447	7.3694912	1.6729558
<i>CNPY4</i>	9.252001333	7.58660075	1.665400583
<i>KPNA2</i>	13.416049	11.7560565	1.6599925
<i>HMMR</i>	10.95662267	9.303328	1.653294667
<i>TOR1AIP2</i>	10.320505	8.6839075	1.6365975
<i>EPB41L5</i>	7.324287633	5.7122538	1.612033833
<i>BTBD7</i>	6.569314733	4.957443	1.611871733

<i>C14orf147</i>	11.97519133	10.363698	1.611493333
<i>ERH</i>	15.31675367	13.7531035	1.563650167
<i>PPIE</i>	8.922634667	7.3673362	1.555298467
<i>CCNK</i>	13.71288933	12.1877805	1.525108833
<i>TANC1</i>	9.361559	7.83677435	1.52478465
<i>TRAM1</i>	10.178868	8.667819	1.511049
<i>CNIH4</i>	13.61134067	12.10617125	1.505169417
<i>C9orf30</i>	11.69050233	10.18583275	1.504669583
<i>FAT4</i>	8.117183667	6.61786185	1.499321817
<i>STK17A</i>	10.04415033	8.544891	1.499259333
<i>MDC1</i>	10.20663467	8.70921025	1.497424417
<i>SSRP1</i>	13.78975633	12.2978595	1.491896833
<i>MOBK1B</i>	9.813573	8.3239795	1.4895935
<i>RUVBL2</i>	13.402765	11.9161415	1.4866235
<i>MPHOSPH6</i>	11.9304925	10.445779	1.4847135
<i>XRN2</i>	12.78236333	11.3082875	1.474075833
<i>GKAP1</i>	12.33944767	10.868156	1.471291667
<i>FAM129A</i>	6.3482872	4.89919115	1.44909605
<i>BZW1</i>	12.2464565	10.8003185	1.446138
<i>YEATS2</i>	12.24813167	10.80285325	1.445278417
<i>NFIA</i>	8.832175	7.4225929	1.4095821
<i>RPL7</i>	16.59612733	15.190894	1.405233333
<i>MAML3</i>	8.972285667	7.58088885	1.391396817
<i>HSD17B4</i>	14.098134	12.711978	1.386156
<i>C15orf23</i>	11.457782	10.073535	1.384247
<i>PFN1</i>	11.57100767	10.227104	1.343903667
<i>FANCD2</i>	6.727573233	5.3840162	1.343557033
<i>ANKRD57</i>	10.13229683	8.79559475	1.336702083
<i>C7orf27</i>	11.53980117	10.2073715	1.332429667
<i>ITFG3</i>	13.01707233	11.685719	1.331353333
<i>ZNF516</i>	6.010824	4.680294	1.33053
<i>SMAD4</i>	12.62974233	11.306637	1.323105333
<i>MDFIC</i>	10.25332067	8.930839	1.322481667
<i>GATM</i>	12.507838	11.1986085	1.3092295
<i>C11orf54</i>	12.40355433	11.0994685	1.304085833
<i>TOR1AIP1</i>	10.152193	8.8500565	1.3021365
<i>FAM46A</i>	10.29806633	9.006315	1.291751333
<i>TNS3</i>	12.24492367	10.9554515	1.289472167
<i>IL20RB</i>	5.918272467	4.63060985	1.287662617
<i>ARID2</i>	9.260570333	7.97375385	1.286816483
<i>HNRPA1</i>	12.49367133	11.217175	1.276496333
<i>HNRPA1L-2</i>	12.49367133	11.217175	1.276496333
<i>HNRNPA1</i>	12.49367133	11.217175	1.276496333
<i>NAB1</i>	10.47960367	9.2184305	1.261173167
<i>JMJD1C</i>	12.16534533	10.905525	1.259820333
<i>PRKCQ</i>	9.156520667	7.896846	1.259674667

<i>FARSA</i>	11.591929	10.3353415	1.2565875
<i>PATZ1</i>	9.303815	8.0473167	1.2564983
<i>MAGED1</i>	15.83678567	14.5935955	1.243190167
<i>HNRPA3</i>	14.06221033	12.819583	1.242627333
<i>HNRNPA3</i>	14.06221033	12.819583	1.242627333
<i>FAM108B1</i>	12.80785733	11.577971	1.229886333
<i>UTP11L</i>	12.36110333	11.1336195	1.227483833
<i>PCBP2</i>	13.97468233	12.7480655	1.226616833
<i>SLC4A7</i>	10.98394833	9.7583305	1.225617833
<i>PAIP1</i>	9.949900333	8.72644425	1.223456083
<i>HLTF</i>	10.84015133	9.6199445	1.220206833
<i>SET</i>	12.90068233	11.6825095	1.218172833
<i>PSMB2</i>	10.88040367	9.66334	1.217063667
<i>TXNIP</i>	10.465087	9.2499435	1.2151435
<i>PTPN13</i>	10.42627133	9.219359	1.206912333
<i>TSLP</i>	4.487806133	3.2888168	1.198989333
<i>HIF1A</i>	9.789705	8.59326575	1.19643925
<i>MAPRE1</i>	11.82648433	10.6361505	1.190333833
<i>TARBP2</i>	13.3119	12.1260405	1.1858595
<i>PRRC1</i>	8.978159	7.7954525	1.1827065
<i>EML4</i>	11.80067767	10.6278295	1.172848167
<i>HEY2</i>	7.1979391	6.0284076	1.1695315
<i>PTPN12</i>	10.35982767	9.191547	1.168280667
<i>RNF182</i>	11.46697467	10.306935	1.160039667
<i>EEF1A1</i>	16.455541	15.299085	1.156456
<i>RBMX</i>	11.59832933	10.442985	1.155344333
<i>NUP54</i>	10.97761233	9.824371	1.153241333
<i>ETAA1</i>	8.324199	7.176437	1.147762
<i>FERMT2</i>	11.84417417	10.7014495	1.142724667
<i>SLC22A5</i>	8.831096667	7.7022585	1.128838167
<i>DOLPP1</i>	13.31661	12.1878165	1.1287935
<i>KT112</i>	11.06163367	9.9361385	1.125495167
<i>SP2</i>	9.432627667	8.3084485	1.124179167
<i>GOLPH3</i>	12.601464	11.4785995	1.1228645
<i>CYTH4</i>	11.39768133	10.2845995	1.113081833
<i>FKBP7</i>	9.6844	8.5756295	1.1087705
<i>ZNF280C</i>	11.04141967	9.935876	1.105543667
<i>ANUBL1</i>	6.159305167	5.07758015	1.081725017
<i>DNAJB12</i>	10.56126067	9.484518	1.076742667
<i>SF3B3</i>	13.567712	12.49131725	1.07639475
<i>FRZB</i>	9.991494	8.915726	1.075768
<i>RAB10</i>	13.266819	12.1959405	1.0708785
<i>ITSN2</i>	12.05704567	11.00214275	1.054902917
<i>ZNF518A</i>	7.377331533	6.3250036	1.052327933
<i>ZNF518</i>	7.377331533	6.3250036	1.052327933
<i>EDARADD</i>	4.029586433	2.98139425	1.048192183

<i>EIF2A</i>	10.504455	9.467629	1.036826
<i>PPAT</i>	9.009667667	7.974397	1.035270667
<i>API5</i>	11.23180317	10.1974675	1.034335667
<i>PSAT1</i>	13.535467	12.503781	1.031686
<i>SCFD1</i>	10.52893633	9.5022405	1.026695833
<i>C22orf28</i>	13.28180533	12.256732	1.025073333
<i>DBT</i>	7.401514033	6.3791883	1.022325733
<i>SGPP1</i>	14.22805633	13.206455	1.021601333
<i>MTA1</i>	10.64938933	9.628594	1.020795333
<i>C21orf91</i>	10.47373733	9.4544045	1.019332833
<i>FAM96A</i>	12.62596083	11.606862	1.019098833
<i>TMOD3</i>	8.232307467	7.21521425	1.017093217
<i>PSMD11</i>	11.950075	10.9395845	1.0104905
<i>E2F3</i>	10.20489267	9.195873	1.009019667
<i>ZKSCAN3</i>	10.72101167	9.715803	1.005208667
<i>ASCC3</i>	10.66187817	9.657851	1.004027167
<i>RBM26</i>	11.71016867	10.71008925	1.000079417
<i>NIPBL</i>	8.80799	7.817832	0.990158
<i>TOPORS</i>	11.20194867	10.21494075	0.987007917
<i>ZNRF2</i>	8.0146098	7.03955555	0.97505425
<i>PDCL</i>	9.953663667	8.9791635	0.974500167
<i>TAF4</i>	11.65959267	10.69551	0.964082667
<i>HDGF</i>	13.021522	12.06166125	0.95986075
<i>EIF2C1</i>	8.480873333	7.52236635	0.958506983
<i>TRPS1</i>	11.03014067	10.0731595	0.956981167
<i>EXOSC3</i>	8.996855333	8.041357	0.955498333
<i>SFXN2</i>	10.47972333	9.5244285	0.955294833
<i>HNRPUL1</i>	10.33144333	9.376985	0.954458333
<i>HNRNPUL1</i>	10.33144333	9.376985	0.954458333
<i>MAP3K7</i>	10.927119	9.9788975	0.9482215
<i>HADH</i>	11.320406	10.3847225	0.9356835
<i>RPL28</i>	16.40758667	15.473649	0.933937667
<i>RAB9A</i>	13.146196	12.21427	0.931926
<i>DHX15</i>	11.24813967	10.333324	0.914815667
<i>TMCO6</i>	8.537101333	7.635406	0.901695333
<i>SAMD4B</i>	12.511014	11.612382	0.898632
<i>FAM160A1</i>	8.784096	7.8888055	0.8952905
<i>SNRPD1</i>	13.50794467	12.622444	0.885500667
<i>BRD2</i>	13.577374	12.695143	0.882231
<i>ZNF281</i>	13.253466	12.37727	0.876196
<i>LRBA</i>	10.57858	9.7101795	0.8684005
<i>UBE2R2</i>	13.38001767	12.5171495	0.862868167
<i>C2orf74</i>	3.003217	2.14190945	0.86130755
<i>PTPRG</i>	9.115675167	8.2599755	0.855699667
<i>SYF2</i>	11.42494467	10.570776	0.854168667
<i>CTDSPL2</i>	7.9055282	7.05172515	0.85380305

<i>MITD1</i>	9.495519	8.6424985	0.8530205
<i>SEC63</i>	9.605885333	8.7542955	0.851589833
<i>ZNF10</i>	13.12742017	12.27662325	0.850796917
<i>MTM1</i>	7.646483267	6.7971245	0.849358767
<i>MEX3C</i>	12.4137885	11.564801	0.8489875
<i>HMGN3</i>	12.71123983	11.8637745	0.847465333
<i>UBE2E3</i>	13.31413333	12.4708635	0.843269833
<i>PDCD2</i>	11.56265167	10.71992025	0.842731417
<i>SASS6</i>	8.587290333	7.7454782	0.841812133
<i>SLC7A5</i>	11.47384367	10.6320525	0.841791167
<i>TIGD2</i>	10.252726	9.413437	0.839289
<i>COG2</i>	9.213897167	8.3839445	0.829952667
<i>PM20D2</i>	6.2989598	5.4720385	0.8269213
<i>C9orf64</i>	7.863701	7.0368717	0.8268293
<i>ALDH3B1</i>	9.785112667	8.960691	0.824421667
<i>XPO7</i>	9.806207	8.982202	0.824005
<i>RPS27A</i>	16.51564067	15.6918965	0.823744167
<i>ZBTB34</i>	6.866484667	6.0450654	0.821419267
<i>NKAP</i>	9.195523	8.3774275	0.8180955
<i>CLIP2</i>	11.17279	10.358368	0.814422
<i>ARPC1A</i>	14.03846533	13.225184	0.813281333
<i>PRDM4</i>	10.30967417	9.4969665	0.812707667
<i>DNAJC19</i>	11.25638767	10.452262	0.804125667
<i>CCDC47</i>	11.73332633	10.931982	0.801344333
<i>PSMA2</i>	15.80439733	15.0058565	0.798540833
<i>KIAA1191</i>	13.95924067	13.1628085	0.796432167
<i>PSMA3</i>	15.27619883	14.4929255	0.783273333
<i>ARL6IP1</i>	13.96232183	13.180584	0.781737833
<i>PRPF38B</i>	10.00931167	9.231162	0.778149667
<i>EIF1AX</i>	13.02006967	12.242232	0.777837667
<i>ZNF384</i>	6.867789133	6.09267865	0.775110483
<i>FAM165B</i>	13.50808867	12.743062	0.765026667
<i>SCARB2</i>	10.730769	9.9688165	0.7619525
<i>TMEM39A</i>	9.660569	8.9073545	0.7532145
<i>NAA15</i>	10.76999883	10.0172195	0.752779333
<i>CTDP1</i>	9.901161333	9.149449	0.751712333
<i>NT5C3L</i>	10.71542233	9.967143	0.748279333
<i>PPP2R3A</i>	6.577442433	5.8327077	0.744734733
<i>MPP6</i>	13.205576	12.464245	0.741331
<i>FAM122B</i>	8.427989	7.6878641	0.7401249
<i>C1orf131</i>	11.4241785	10.686046	0.7381325
<i>CEP76</i>	8.061292733	7.3278961	0.733396633
<i>STAT6</i>	7.748803433	7.0196965	0.729106933
<i>EGLN3</i>	7.628265967	6.9009892	0.727276767
<i>ZC3H11A</i>	14.04229733	13.320265	0.722032333
<i>ZSCAN12</i>	10.79773167	10.07598	0.721751667

<i>RPL9</i>	17.49996133	16.7800435	0.719917833
<i>PTGES3</i>	13.58813767	12.868388	0.719749667
<i>DDX3X</i>	13.4936	12.7766475	0.7169525
<i>PAK1IP1</i>	13.55931267	12.846343	0.712969667
<i>TNRC6B</i>	9.689875667	8.97833925	0.711536417
<i>LRRC59</i>	11.301553	10.5995015	0.7020515
<i>C14orf119</i>	10.55518967	9.8543315	0.700858167
<i>RIT1</i>	13.76958933	13.0743055	0.695283833
<i>SPOPL</i>	8.043102567	7.355728	0.687374567
<i>BCL7B</i>	11.21988367	10.541217	0.678666667
<i>TMCO1</i>	12.101512	11.429836	0.671676
<i>KIF2A</i>	9.452399667	8.7856165	0.666783167
<i>HNRNPH1</i>	14.99481933	14.333635	0.661184333
<i>HNRPH1</i>	14.99481933	14.333635	0.661184333
<i>CTSA</i>	13.48982233	12.8311805	0.658641833
<i>PDS5A</i>	9.215477	8.5579885	0.6574885
<i>DIAPH2</i>	10.62390567	9.968658	0.655247667
<i>RPL26L1</i>	15.97318867	15.3189	0.654288667
<i>PPP1R12A</i>	12.65975433	12.00622	0.653534333
<i>SFRS11</i>	11.41057	10.7610435	0.6495265
<i>NUTF2</i>	12.705992	12.0666155	0.6393765
<i>SRP19</i>	11.77192333	11.139446	0.632477333
<i>TMEM188</i>	10.92748033	10.3007225	0.626757833
<i>STXBP3</i>	11.095598	10.471581	0.624017
<i>ZEB2</i>	10.45435433	9.8319135	0.622440833
<i>CSNK1E</i>	11.921135	11.3055505	0.6155845
<i>RPL4</i>	15.17975333	14.5648335	0.614919833
<i>PPP6C</i>	12.068744	11.4559705	0.6127735
<i>CNBP</i>	13.740117	13.127783	0.612334
<i>C2orf29</i>	8.991943	8.380669	0.611274
<i>ANKRD17</i>	11.94080067	11.33413025	0.606670417
<i>MED21</i>	11.14159367	10.548392	0.593201667
<i>BRD4</i>	13.26419367	12.6734365	0.590757167
<i>ZNF85</i>	6.616636167	6.03524835	0.581387817
<i>FOXO1</i>	11.52472133	10.94465	0.580071333
<i>FOXO1A</i>	11.52472133	10.94465	0.580071333
<i>ACVR1</i>	11.71697633	11.1440155	0.572960833
<i>ZBTB26</i>	7.686974833	7.11812515	0.568849683
<i>RSBN1</i>	9.851864167	9.286532	0.565332167
<i>BIRC6</i>	10.42141217	9.8563525	0.565059667
<i>KTN1</i>	12.740367	12.17646075	0.56390625
<i>PATL1</i>	9.262394	8.6999805	0.5624135
<i>CLIC4</i>	11.73545867	11.1733885	0.562070167
<i>HOXA10</i>	4.213144967	3.6510829	0.562062067
<i>YOD1</i>	8.493824667	7.93247735	0.561347317
<i>GTF2IP1</i>	14.04781	13.48916	0.55865

<i>KIAA1143</i>	9.462226333	8.9077655	0.554460833
<i>ANKRA2</i>	13.1203675	12.5664645	0.553903
<i>ELL</i>	12.04305167	11.489632	0.553419667
<i>CPT1A</i>	10.34946733	9.799435	0.550032333
<i>FAM192A</i>	11.731589	11.181962	0.549627
<i>ARAP2</i>	10.307713	9.7593975	0.5483155
<i>TIPRL</i>	10.63746367	10.0901355	0.547328167
<i>EPB41L2</i>	8.794291	8.2556725	0.5386185
<i>RNF149</i>	11.36055	10.82867675	0.53187325
<i>POLR2A</i>	9.275875667	8.7444035	0.531472167
<i>MAGEF1</i>	10.689974	10.1612265	0.5287475
<i>UBE2L3</i>	14.027295	13.49873825	0.52855675
<i>ARIH2</i>	12.87555567	12.357152	0.518403667
<i>WBP5</i>	12.37341233	11.862783	0.510629333
<i>MYH9</i>	12.37811167	11.8702955	0.507816167
<i>HEATR2</i>	9.681272	9.1768095	0.5044625
<i>MRPL44</i>	11.581505	11.077092	0.504413
<i>PPP1CC</i>	14.36415633	13.86223975	0.501916583
<i>RAB21</i>	9.58748	9.0875005	0.4999795
<i>FAM114A2</i>	12.04244267	11.54326625	0.499176417
<i>RSRC2</i>	11.07299433	10.576798	0.496196333
<i>HMBX1</i>	8.984691667	8.4887855	0.495906167
<i>EED</i>	12.03852767	11.5468795	0.491648167
<i>SDCCAG1</i>	12.326123	11.8351515	0.4909715
<i>SAPS3</i>	8.968611667	8.4829725	0.485639167
<i>FUT8</i>	10.951779	10.46860025	0.48317875
<i>LRRC57</i>	10.88937867	10.411121	0.478257667
<i>CHUK</i>	12.459115	11.985941	0.473174
<i>PIGO</i>	12.59320267	12.122987	0.470215667
<i>CDH2</i>	13.499508	13.0366405	0.4628675
<i>APPL1</i>	11.06090467	10.59809425	0.462810417
<i>CDKN2AIP</i>	6.941651367	6.4800188	0.461632567
<i>ACTG1</i>	16.52307067	16.063874	0.459196667
<i>GOLT1B</i>	11.15495717	10.696702	0.458255167
<i>MRPL42</i>	11.448289	10.9911155	0.4571735
<i>RAD23B</i>	10.38233433	9.930453	0.451881333
<i>TMEM38B</i>	6.0282082	5.58102485	0.44718335
<i>ZNF711</i>	10.02747117	9.581473	0.445998167
<i>MRPL35</i>	9.478753667	9.054354	0.424399667
<i>UBA6</i>	8.172925	7.7488902	0.4240348
<i>C7orf60</i>	9.043251333	8.621266	0.421985333
<i>CAMLG</i>	12.597606	12.1803	0.417306
<i>PUM1</i>	12.368029	11.9532115	0.4148175
<i>ZFP36</i>	2.719485367	2.3093946	0.410090767
<i>ABCA11P</i>	8.107929667	7.69810035	0.409829317
<i>RPAP3</i>	12.697974	12.2932355	0.4047385

<i>PLA1A</i>	7.7574976	7.3614845	0.3960131
<i>MLL5</i>	11.3105585	10.921768	0.3887905
<i>CYB5A</i>	14.22998833	13.8450235	0.384964833
<i>NCK2</i>	11.05282133	10.670069	0.382752333
<i>UGT8</i>	11.12815767	10.7491605	0.378997167
<i>ERICH1</i>	6.932701367	6.55526255	0.377438817
<i>GRK5</i>	8.684448667	8.30946025	0.374988417
<i>HSPA4</i>	12.30820733	11.93805	0.370157333
<i>NUFIP2</i>	11.48359333	11.117281	0.366312333
<i>MYLIP</i>	9.188353333	8.8442415	0.344111833
<i>DYRK1A</i>	8.960179333	8.623358	0.336821333
<i>DDX50</i>	10.78899783	10.454258	0.334739833
<i>MYST2</i>	13.49850567	13.166882	0.331623667
<i>RANBP2</i>	11.044413	10.7141715	0.3302415
<i>S100PBP</i>	7.2220536	6.89271305	0.32934055
<i>WSB1</i>	9.295123	8.975541	0.319582
<i>USP34</i>	9.844100333	9.525011	0.319089333
<i>TSEN15</i>	12.36043933	12.0424535	0.317985833
<i>SLC35A5</i>	7.221875333	6.9088235	0.313051833
<i>SERINC3</i>	13.884642	13.5741045	0.3105375
<i>C1orf35</i>	11.55478067	11.25494075	0.299839917
<i>PPFIA1</i>	10.574596	10.276806	0.29779
<i>PIAS2</i>	11.22387867	10.935567	0.288311667
<i>FAM18B</i>	11.04391967	10.75617775	0.287741917
<i>CDC42SE2</i>	12.65318267	12.366041	0.287141667
<i>USP48</i>	11.347884	11.0613715	0.2865125
<i>USP25</i>	9.667973667	9.3952525	0.272721167
<i>LEF1</i>	10.43159533	10.1726055	0.258989833
<i>SDHB</i>	15.04432	14.78738	0.25694
<i>SHPRH</i>	7.768260533	7.5126476	0.255612933
<i>WWP1</i>	8.051860133	7.796494	0.255366133
<i>GOLGA2</i>	13.46896367	13.2198	0.249163667
<i>GLUD1</i>	14.17524667	13.928764	0.246482667
<i>CAMSAP1L1</i>	9.191947	8.9496975	0.2422495
<i>EIF4G2</i>	12.090261	11.849496	0.240765
<i>RALGDS</i>	11.71140433	11.4736995	0.237704833
<i>ZNF235</i>	9.6403365	9.40318575	0.23715075
<i>SMARCD2</i>	8.067492833	7.8362415	0.231251333
<i>NUDT4</i>	13.08851367	12.8599435	0.228570167
<i>NUDT4P1</i>	13.08851367	12.8599435	0.228570167
<i>PKD1</i>	12.09181333	11.8639325	0.227880833
<i>PDK1</i>	9.062059667	8.834268	0.227791667
<i>SRGAP2</i>	9.067219167	8.846632	0.220587167
<i>TMED3</i>	13.751069	13.531832	0.219237
<i>UBE2D1</i>	7.349367533	7.1320193	0.217348233
<i>ALDH3A2</i>	9.626151667	9.410587	0.215564667

<i>CEP55</i>	6.4713698	6.25650735	0.21486245
<i>AP3S1</i>	11.67902167	11.4647465	0.214275167
<i>SAE1</i>	11.49945917	11.2869705	0.212488667
<i>MGAT4A</i>	7.842548667	7.63477725	0.207771417
<i>NUMA1</i>	13.326883	13.1275775	0.1993055
<i>MBNL1</i>	11.57254167	11.374435	0.198106667
<i>DUSP11</i>	8.209455	8.01685075	0.19260425
<i>C1orf55</i>	9.749012	9.557061	0.191951
<i>TM4SF1</i>	11.94979133	11.758792	0.190999333
<i>TRIM33</i>	10.240359	10.049575	0.190784
<i>LRRFIP1</i>	11.39851033	11.2087305	0.189779833
<i>ORC5L</i>	8.5886675	8.400017	0.1886505
<i>VCPIP1</i>	10.872761	10.6877045	0.1850565
<i>NISCH</i>	16.26442067	16.080325	0.184095667
<i>FAM133B</i>	9.537814667	9.3541265	0.183688167
<i>MGC40405</i>	9.537814667	9.3541265	0.183688167
<i>SIRT7</i>	11.289323	11.1100215	0.1793015
<i>ZNF148</i>	11.80478933	11.6286175	0.176171833
<i>TMEM200B</i>	3.686916633	3.5187699	0.168146733
<i>NEDD8</i>	14.5481705	14.383052	0.1651185
<i>C12orf66</i>	10.24941367	10.0895995	0.159814167
<i>GFPT1</i>	10.81721533	10.658652	0.158563333
<i>NRBP1</i>	12.05195233	11.898632	0.153320333
<i>CTSB</i>	15.31927267	15.174771	0.144501667
<i>BPNT1</i>	9.245479667	9.1072435	0.138236167
<i>COL4A1</i>	9.9055725	9.77018	0.1353925
<i>YPEL5</i>	10.94790433	10.814909	0.132995333
<i>FAM160B1</i>	12.09945467	11.9688185	0.130636167
<i>ZRANB3</i>	7.650254233	7.524397	0.125857233
<i>ACSS2</i>	10.900509	10.7799135	0.1205955
<i>CCDC71</i>	11.670102	11.5506715	0.1194305
<i>MID1IP1</i>	14.152342	14.0338035	0.1185385
<i>CACHD1</i>	10.2285685	10.110708	0.1178605
<i>ZNF236</i>	8.936669	8.819343	0.117326
<i>PCDHA1</i>	4.344098333	4.2400979	0.104000433
<i>TNRC6A</i>	10.84121867	10.73869175	0.102526917
<i>SLTM</i>	13.019364	12.9188355	0.1005285
<i>KIF3A</i>	13.036739	12.9386785	0.0980605
<i>PDXDC1</i>	9.922680667	9.8283915	0.094289167
<i>HOXA13</i>	3.446191867	3.3553231	0.090868767
<i>FBXL5</i>	9.474731333	9.384101	0.090630333
<i>AK3</i>	12.935957	12.84621	0.089747
<i>ZDHHC21</i>	10.1356645	10.0481725	0.087492
<i>ATP5F1</i>	13.62961267	13.54302	0.086592667
<i>ACBD5</i>	9.385822333	9.3019115	0.083910833
<i>FAS</i>	7.6992079	7.6153353	0.0838726

<i>C1orf109</i>	9.443861667	9.3616705	0.082191167
<i>TMEM30A</i>	11.48839	11.416312	0.072078
<i>FSTL5</i>	10.74537733	10.676923	0.068454333
<i>ZNF646</i>	6.918685867	6.85081475	0.067871117
<i>MIB1</i>	10.370216	10.315217	0.054999
<i>ARRDC3</i>	8.896145667	8.852864	0.043281667
<i>KIAA0494</i>	11.92758333	11.8850995	0.042483833
<i>CUTC</i>	6.576739633	6.53463415	0.042105483
<i>MYO18A</i>	11.50076033	11.458709	0.042051333
<i>OVOL2</i>	11.60201833	11.560376	0.041642333
<i>CLTB</i>	11.47718267	11.4380395	0.039143167
<i>SVIP</i>	12.33004333	12.294543	0.035500333
<i>HNRPH3</i>	8.540294667	8.5050685	0.035226167
<i>HNRNPH3</i>	8.540294667	8.5050685	0.035226167
<i>NPEPPS</i>	13.04731017	13.013276	0.034034167
<i>UBE2E2</i>	13.78405633	13.7506	0.033456333
<i>TMEM134</i>	11.98169233	11.9577115	0.023980833
<i>C5orf41</i>	6.587165333	6.5670827	0.020082633
<i>CAPZA2</i>	12.58081967	12.5681145	0.012705167
<i>SCML1</i>	2.354425867	2.3459189	0.008506967
<i>TOM1L1</i>	11.40290033	11.397205	0.005695333
<i>IGSF3</i>	9.928223333	9.9265355	0.001687833
<i>PDS5B</i>	9.862373667	9.864843	-0.002469333
<i>TARSL2</i>	11.42246933	11.4374335	-0.014964167
<i>FAM18B2</i>	10.175684	10.1932825	-0.0175985
<i>AXIN2</i>	4.749037467	4.77437115	-0.025333683
<i>AKAP9</i>	11.871409	11.898516	-0.027107
<i>YME1L1</i>	7.494506233	7.5275979	-0.033091667
<i>CIRH1A</i>	11.39184	11.437048	-0.045208
<i>CAT</i>	10.450668	10.49774325	-0.04707525
<i>NCKAP1</i>	14.02692567	14.0856715	-0.058745833
<i>KLHDC5</i>	8.916073333	8.980655	-0.064581667
<i>PRKAR1A</i>	12.55782733	12.62793	-0.070102667
<i>ZCCHC10</i>	9.641000667	9.7123385	-0.071337833
<i>MSL1</i>	13.267942	13.3402475	-0.0723055
<i>MSL-1</i>	13.267942	13.3402475	-0.0723055
<i>ZNRF3</i>	6.616273133	6.6895093	-0.073236167
<i>ZDHHC17</i>	10.94823633	11.021534	-0.073297667
<i>HNRNPH2</i>	11.103904	11.178838	-0.074934
<i>UNKL</i>	5.985900233	6.0617349	-0.075834667
<i>NOG</i>	7.309365267	7.388024	-0.078658733
<i>FNIP1</i>	11.435613	11.516924	-0.081311
<i>ARF1</i>	15.256873	15.348452	-0.091579
<i>RPS6KA5</i>	8.632861	8.724488	-0.091627
<i>C12orf75</i>	9.421273833	9.521531	-0.100257167
<i>CCNE1</i>	10.25640667	10.357795	-0.101388333

<i>C12orf4</i>	10.61039167	10.712795	-0.102403333
<i>CGRRF1</i>	10.03357017	10.1464975	-0.112927333
<i>FAM178A</i>	8.284600767	8.400291	-0.115690233
<i>ZFR</i>	7.754366667	7.877913	-0.123546333
<i>RNF19A</i>	11.309954	11.43887575	-0.12892175
<i>SEMA4D</i>	9.897528	10.0269665	-0.1294385
<i>TMEM170A</i>	8.377891667	8.5106885	-0.132796833
<i>ARHGAP28</i>	7.5880931	7.73192	-0.1438269
<i>PEA15</i>	13.82505933	13.9695115	-0.144452167
<i>STIM2</i>	7.7785526	7.92402695	-0.14547435
<i>ZC3H13</i>	10.93117583	11.0820265	-0.150850667
<i>RNMT</i>	9.851374333	10.005055	-0.153680667
<i>ZFYVE21</i>	10.513293	10.6848225	-0.1715295
<i>FAM49B</i>	10.98356433	11.1580445	-0.174480167
<i>POLK</i>	7.618777467	7.79444365	-0.175666183
<i>CREM</i>	8.3861128	8.5652215	-0.1791087
<i>ZBTB38</i>	8.1358152	8.316282	-0.1804668
<i>SUCLG1</i>	13.54598733	13.7277175	-0.181730167
<i>C3orf52</i>	3.5607469	3.74693125	-0.18618435
<i>UBE2V2</i>	12.05168167	12.24245675	-0.190775083
<i>CELF1</i>	11.88589967	12.0830305	-0.197130833
<i>VLDLR</i>	11.457448	11.6680295	-0.2105815
<i>CREBZF</i>	10.96209533	11.18086	-0.218764667
<i>ZF</i>	10.96209533	11.18086	-0.218764667
<i>IMMT</i>	12.23348067	12.4584715	-0.224990833
<i>FOXG1</i>	13.453163	13.683055	-0.229892
<i>RAPGEF2</i>	12.76868067	12.9994665	-0.230785833
<i>SLMAP</i>	11.175495	11.4097655	-0.2342705
<i>TMEM216</i>	9.951063667	10.1890265	-0.237962833
<i>UBE4A</i>	9.909663333	10.152771	-0.243107667
<i>UBE2N</i>	12.56183	12.8097835	-0.2479535
<i>YAF2</i>	12.67235633	12.921328	-0.248971667
<i>STARD4</i>	8.598857	8.8480245	-0.2491675
<i>C17orf71</i>	1.737531767	1.9913844	-0.253852633
<i>LOC440248</i>	12.34574067	12.600986	-0.255245333
<i>ASAP1</i>	10.78869933	11.0607815	-0.272082167
<i>EXD2</i>	8.536598667	8.820965	-0.284366333
<i>OLA1</i>	12.06643133	12.3514965	-0.285065167
<i>MTMR4</i>	9.218658333	9.5138205	-0.295162167
<i>OCRL</i>	11.979835	12.281157	-0.301322
<i>GADD45A</i>	8.634649667	8.94306	-0.308410333
<i>BBS2</i>	9.061666333	9.389736	-0.328069667
<i>RYBP</i>	12.274737	12.616579	-0.341842
<i>GCNT2</i>	8.376630167	8.727686	-0.351055833
<i>PIGK</i>	8.610945533	8.9654205	-0.354474967
<i>STON2</i>	6.614778367	6.98561925	-0.370840883

<i>ARHGAP5</i>	9.582782667	9.9555475	-0.372764833
<i>TMCC1</i>	9.591185	9.969172	-0.377987
<i>CAMSAP1</i>	10.98071617	11.3673645	-0.386648333
<i>MNX1</i>	2.662742867	3.055465	-0.392722133
<i>CCDC91</i>	10.08490567	10.479166	-0.394260333
<i>NDFIP1</i>	15.28807233	15.694662	-0.406589667
<i>PTPLAD1</i>	15.10839433	15.522724	-0.414329667
<i>CBWD1</i>	8.510827667	8.9279335	-0.417105833
<i>CBWD5</i>	8.510827667	8.9279335	-0.417105833
<i>ENC1</i>	14.926894	15.3475415	-0.4206475
<i>C1S</i>	5.712193367	6.134924	-0.422730633
<i>FBXO33</i>	10.74888917	11.1779835	-0.429094333
<i>NDUFA5</i>	15.01706867	15.44720125	-0.430132583
<i>PLCL2</i>	12.16204167	12.6175765	-0.455534833
<i>SLC16A9</i>	7.278990433	7.7348343	-0.455843867
<i>Kua</i>	7.747335067	8.20966475	-0.462329683
<i>TMEM189</i>	7.747335067	8.20966475	-0.462329683
<i>DUSP1</i>	9.820587667	10.31625725	-0.495669583
<i>MICB</i>	4.564319433	5.06018775	-0.495868317
<i>GHITM</i>	13.70630167	14.20532775	-0.499026083
<i>NUB1</i>	12.21879983	12.724183	-0.505383167
<i>IPO7</i>	7.990636	8.4965035	-0.5058675
<i>AKT3</i>	11.17457367	11.6880635	-0.513489833
<i>SH3BP5</i>	9.865592667	10.3881775	-0.522584833
<i>SC5DL</i>	10.872775	11.3958445	-0.5230695
<i>SH3BGRL2</i>	7.440453333	7.96968735	-0.529234017
<i>PHLPP2</i>	10.40592967	10.936602	-0.530672333
<i>TMEM65</i>	10.14398533	10.6769745	-0.532989167
<i>COL4A5</i>	6.5221753	7.0623648	-0.5401895
<i>KCTD12</i>	9.883777833	10.4408545	-0.557076667
<i>TUSC3</i>	12.74822017	13.310978	-0.562757833
<i>UBFD1</i>	10.58395467	11.1498955	-0.565940833
<i>UBE2W</i>	9.811682333	10.38336225	-0.571679917
<i>EGR1</i>	13.497957	14.070696	-0.572739
<i>USP32</i>	12.56942267	13.15562075	-0.586198083
<i>KLF4</i>	9.185409833	9.7868945	-0.601484667
<i>LRP12</i>	10.20010533	10.803612	-0.603506667
<i>WASF1</i>	13.06895567	13.6733725	-0.604416833
<i>CDC42EP3</i>	9.315261	9.9219165	-0.6066555
<i>APLP2</i>	14.6111025	15.2254535	-0.614351
<i>PIK3R6</i>	10.03912967	10.654058	-0.614928333
<i>CYLD</i>	11.94282233	12.5945085	-0.651686167
<i>FAM129B</i>	11.748755	12.40305375	-0.65429875
<i>GPD2</i>	9.901104667	10.560238	-0.659133333
<i>HDGFRP3</i>	11.55979067	12.223469	-0.663678333
<i>CPEB4</i>	7.532716033	8.2130225	-0.680306467

MYL12B	14.413844	15.110799	-0.696955
TMEM9B	13.27402967	13.97350425	-0.699474583
BCAS4	9.277151	9.9787605	-0.7016095
PFN2	13.82191733	14.524282	-0.702364667
RNF6	13.277984	13.9810505	-0.7030665
PAFAH1B1	13.447666	14.1639415	-0.7162755
TECPR2	9.707360667	10.4391045	-0.731743833
HERPUD1	12.022726	12.7569885	-0.7342625
CLIP1	10.073685	10.821334	-0.747649
CLASP2	6.556482867	7.31822485	-0.761741983
ARFGEF2	8.6401645	9.407491	-0.7673265
KIAA1324L	8.995288667	9.7842435	-0.788954833
PPP1R9A	11.841578	12.647974	-0.806396
GADD45B	9.378380167	10.18825225	-0.809872083
ZMAT2	13.228039	14.0590575	-0.8310185
AUTS2	8.566506333	9.410874	-0.844367667
SLC20A1	12.35033967	13.1975235	-0.847183833
C12orf34	8.927219	9.7759385	-0.8487195
GNAQ	11.39532467	12.2590095	-0.863684833
ACSL1	10.40712633	11.2955485	-0.888422167
RUNX1T1	10.554712	11.4438095	-0.8890975
LYSMD2	11.95001867	12.8611645	-0.911145833
ISCA1	11.681055	12.609982	-0.928927
AKTIP	10.349402	11.2851205	-0.9357185
PFKM	14.15487467	15.0920675	-0.937192833
TLE4	6.7755346	7.71809965	-0.94256505
NEGR1	9.510857333	10.460776	-0.949918667
PPARG	7.9792308	8.9397705	-0.9605397
MBNL2	11.56707767	12.5457645	-0.978686833
BTRC	10.69247	11.686335	-0.993865
PHF15	12.37131833	13.379538	-1.008219667
RNF14	13.5302745	14.5434255	-1.013151
HSP90AA1	13.97251267	14.993994	-1.021481333
PPTC7	12.168459	13.202816	-1.034357
AUH	10.786573	11.8320765	-1.0455035
MEGF8	11.84077733	12.8871425	-1.046365167
ABT1	4.840745833	5.89794635	-1.057200517
AP1B1	13.11195533	14.171299	-1.059343667
GSK3A	13.796452	14.858959	-1.062507
SYBU	9.864202667	10.929427	-1.065224333
TOMM70A	10.557896	11.625362	-1.067466
PROK2	4.6598262	5.75037885	-1.09055265
ELMO2	8.135605467	9.2317065	-1.096101033
MAP7D2	10.28183	11.4207245	-1.1388945
LOC150786	12.982867	14.1792055	-1.1963385
PDLIM1	10.39069333	11.6151195	-1.224426167

<i>MAP2K1</i>	12.48229567	13.716871	-1.234575333
<i>SMARCA2</i>	10.88608133	12.1244865	-1.238405167
<i>CAMKK2</i>	12.8089115	14.050014	-1.2411025
<i>OXR1</i>	11.73885567	12.98967075	-1.250815083
<i>CALM1</i>	15.65122	16.9556255	-1.3044055
<i>REEP5</i>	11.55542567	12.932383	-1.376957333
<i>CELSR1</i>	7.728699167	9.12277475	-1.394075583
<i>CHSY3</i>	5.313862833	6.76258135	-1.448718517
<i>C5orf23</i>	5.850678133	7.33065715	-1.479979017
<i>NPR3</i>	5.850678133	7.33065715	-1.479979017
<i>MMD</i>	11.08337633	12.61058575	-1.527209417
<i>C17orf65</i>	6.231619767	7.78656505	-1.554945283
<i>L1CAM</i>	7.510627167	9.087492	-1.576864833
<i>PNOC</i>	9.160971333	10.7939505	-1.632979167
<i>TTLL11</i>	8.445152	10.096302	-1.65115
<i>LPGAT1</i>	10.82093967	12.48566	-1.664720333
<i>NRXN3</i>	9.930639667	11.6281585	-1.697518833
<i>STMN1</i>	14.56718933	16.322691	-1.755501667
<i>ADRBK1</i>	12.07454767	13.85847575	-1.783928083
<i>CAMK1D</i>	10.025003	11.8451955	-1.8201925
<i>KLC1</i>	11.00575033	12.88402525	-1.878274917
<i>MAP1B</i>	10.19971067	12.1019095	-1.902198833
<i>LPPR4</i>	10.20632333	12.1533535	-1.947030167
<i>SEMA3A</i>	6.706816733	8.6601	-1.953283267
<i>FLRT2</i>	6.986485733	9.0488225	-2.062336767
<i>ROBO2</i>	7.8601465	9.944533	-2.0843865
<i>PAK1</i>	11.47429433	13.561988	-2.087693667
<i>PRR16</i>	2.9898541	5.0982571	-2.108403
<i>MLLT11</i>	13.40149233	15.523164	-2.121671667
<i>NOV</i>	6.94895	9.0730645	-2.1241145
<i>BNIP3</i>	11.63875833	13.7746475	-2.135889167
<i>BMP6</i>	6.297023667	8.510147	-2.213123333
<i>PLCXD2</i>	10.28670133	12.5517845	-2.265083167
<i>KIF21A</i>	11.076628	13.3606855	-2.2840575
<i>DUSP5</i>	3.340393867	5.72126865	-2.380874783
<i>ZNF483</i>	3.473055	5.85734585	-2.38429085
<i>PREPL</i>	9.386301667	11.7787345	-2.392432833
<i>CACNB4</i>	9.237837667	11.6662525	-2.428414833
<i>LEPRE1</i>	9.402561667	11.8599265	-2.457364833
<i>TMEM74</i>	5.024459867	7.50901245	-2.484552583
<i>TRIM37</i>	11.900515	14.4084515	-2.5079365
<i>PRDM8</i>	7.6409638	10.2139255	-2.5729617
<i>ATP1B1</i>	13.65787783	16.3473775	-2.689499667
<i>EFHD1</i>	8.263604633	11.0071155	-2.743510867
<i>FAIM3</i>	2.624274167	5.40133975	-2.777065583
<i>CDS1</i>	7.033244633	9.9707135	-2.937468867

<i>LUM</i>	5.150454233	8.181677	-3.031222767
<i>FGF9</i>	7.915199667	11.4172335	-3.502033833
<i>MAL2</i>	9.020901433	13.3227485	-4.301847067
<i>C9orf41</i>	N/A	N/A	N/A
<i>ZNF721</i>	N/A	N/A	N/A
<i>ZNF200</i>	N/A	N/A	N/A
<i>LOC645323</i>	N/A	N/A	N/A
<i>ZNF567</i>	N/A	N/A	N/A
<i>ZNF680</i>	N/A	N/A	N/A
<i>LRRC37B2</i>	N/A	N/A	N/A
<i>ZNF197</i>	N/A	N/A	N/A
<i>LOC100093631</i>	N/A	N/A	N/A
<i>LOC100132987</i>	N/A	N/A	N/A
<i>GIMAP2</i>	N/A	N/A	N/A

N/A : not applicable

Supplementary Table 4. Expressions of 13 transcriptional factors in MADM model mice and Glioma tissues

MADM mouse model

	MADM / Normal (log ₂)	MADM (log ₂)	Normal (log ₂)	P value
SOX3	6.330985	14.33052	7.999534	0.000794
PRRX1	2.641517	9.561797	6.92028	0.000789
TCF3	2.141706	10.86213	8.720428	0.002462
SOX9	1.940535	11.54724	9.606703	0.037565
TCF12	1.89209	10.46643	8.574336	0.004121
SP1	1.799953	11.76423	9.964281	0.003437
NRF1	1.697251	9.831535	8.134284	0.000173
SP3	1.622128	3.558814	1.936686	0.001093
ATF1	1.317528	9.347354	8.029826	5.33E-05
HLTF	1.220207	10.84015	9.619945	0.012216
ELK4	1.047303	8.270951	7.223648	0.010908
HINFP	0.910814	8.081337	7.170523	0.047654
TCF4	0.848959	13.63375	12.78479	0.021845

Glioma tissues

	Glioma / Normal (log ₂)	Glioma (log ₂)	Normal (log ₂)	P value
TCF12	2.876284	12.89876	10.02248	0.001095
SOX3	2.785636	6.879216	4.09358	0.001797
PRRX1	2.749149	10.63573	7.88658	0.000677
TCF3	1.933415	10.82735	8.89394	0.000538
SP1	0.969601	10.38584	9.41624	0.027065
SOX9	0.938038	11.8588	10.92076	2.73E-05
TCF4	0.665662	12.31098	11.64532	5.26E-06
SP3	0.625978	10.47168	9.8457	0.001198
ATF1	0.552589	7.696289	7.1437	0.005152
HLTF	0.392144	10.336	9.94386	0.010001
ELK4	0.353717	6.127337	5.77362	0.034357
NRF1	0.327728	8.775868	8.44814	0.018198
HINFP	0.189566	8.805186	8.61562	0.000622

Supplementary Table 5. Primer sequences in this study

Oligonucleotide primers used for TaqMan PCR assays

Target gene	AB assay ID
<i>GAPDH</i>	Hs00266705_g1
<i>hsa-miR-9-2</i>	Hs03303202_pri
<i>miR-411</i>	001610
<i>RNU6B</i>	001093
<i>snoRNA202</i>	001232

Primer sequences for SYBR green PCR assays

Target gene	Primer sequence (5' to 3')
<i>LINC00461</i>	Forward: GACAATCCAACCAACCAAGA Reverse: TAGGGGGAAGATCCTCGATT
<i>TOP2A</i>	Forward: CAAACAAAGGGACCCAAAAA Reverse: CAACTTTGTGTTCAACAACAGGA
<i>C130071C03Rik</i> (mouse)	Forward: TGACACTTCAAAGAAGCATAAAATG Reverse: TGTGAATGTTTTAAGGGAGATCCT
<i>Top2a</i> (mouse)	Forward: GCCATCTTCTTCTGATAGCTCTG Reverse: TATCGTGTCTTCCAAGTCCACA
<i>Gapdh</i> (mouse)	Forward: GAATTTGCCGTGAGTGGAGT Reverse: CGTCCCGTAGACAAAATGGT

Supplementary Table 6. siRNA targeting sequences for each gene

Target gene	Targeted sequence (5' to 3')
<i>LINC00461</i> #1	AAGAAAAGTTCGGAATAAACTCT
<i>LINC00461</i> #2	GTCAATTTGTACGAAATTAGACT
<i>LINC00461</i> #3	GAGAAAGAAGCAATTACAATGC
<i>LINC00461</i> #4	ACCCATTTAACTATTTTAGACG
<i>LINC00461</i> #5	AGCATAAATGAATAGACAATCC
<i>TOP2A</i> (Santa Cruz Biotechnology)	Catalog No.sc-36695

Supplementary Table 7. Primer sequences for construction of FLAG-AGO2 variant vector

Target gene	Primer sequence (5' to 3')
AGO2 with mutated PAZ domain (Y311A/F312A)	Forward: GCAGCTCAAGGACAGGCACA Reverse: TGGGCCACCGTGCACTC
AGO2 with deleted PAZ domain	Forward: CTGTGCCTTGTAACGCT Reverse: GCAGGACAAAGATGTATTAAAAA

Supplementary Table 8. Primer sequences for manipulation of *TOP2A* analyses

Primer sequences for manipulation of *TOP2A* 3'-UTR pmir-GLO luciferase reporter vector

Gene	Primer sequence (5' to 3')
<i>TOP2A</i> -3'UTR	Forward: GCCTCGAGAATGTGAGGCGATTATTTAAGTAA Reverse: ACTCTAGATTCTATGGGTTGCAATGTTTAGAAC

Primer sequences for manipulation of *TOP2A* vector with gateway cloning technology

Gene	Primer sequence (5' to 3')
<i>TOP2A</i>	Forward: CACCATGGAAGTGTACCATTGCA Reverse: TTAACACAGATCATCTTCATCTGACTC