

# **Prognostic analysis according to the 2017 ELN risk stratification by genetics in adult acute myeloid leukemia patients treated in the Japan Adult Leukemia Study Group (JALSG) AML201 study**

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## Abstract

Many genetic alterations that are associated with the prognosis of acute myeloid leukemia (AML) have been identified, and several risk stratification systems based on the genetic status have been recommended. The European LeukemiaNet (ELN) first proposed the risk stratification system for AML in 2010 (ELN-2010), and recently published the revised system (ELN-2017). We validated the long-term prognosis and clinical characteristics of each ELN-2017 risk category in Japanese adult AML patients who were treated in the Japan Adult Leukemia Study Group (JALSG) AML-201 study. We demonstrated that the 3-risk category system of the ELN-2017 could clearly discriminate the overall survival and complete remission rates in our cohort in comparison with the 4-risk category of the ELN-2010. However, there were still genetic categories in which stratification of patients into favorable or intermediate risk categories was controversial; the low allelic ratio of *FLT3*-ITD was not necessarily associated with a better prognosis in patients with *FLT3*-ITD, and cytogenetic abnormalities may

affect the prognosis in patients with favorable genetic lesions such as *NPM1* and *CEBPA* mutations. As many molecular targeting agents, such as FLT3 inhibitors, have been developed, we must continue to modify the genetic risk stratification system with the progress of therapeutic strategies.

*Keywords:*

Acute myeloid leukemia; prognosis; risk stratification; European LeukemiaNet; genetics

## 1. Introduction

Acute myeloid leukemia (AML) is a clinically and genetically heterogeneous disease [1, 2]. Therefore, the evaluation of the prognostic risk is clinically important for the AML patients to determine the appropriate therapeutic strategy. Medical Research Council (MRC) developed the cytogenetic classification system in 1998, and it was refined by considering clinical characteristic and prognostic relevance of rare cytogenetic abnormalities [3, 4]. The refined MRC system, in which three cytogenetic risk groups are distinguished, is widely used for cytogenetic risk stratification of younger adults with AML. However, as there are limitations for patients in the intermediate-risk group, particularly those with cytogenetically normal (CN)-AML [4], more precise risk stratification systems based on genetic status have been proposed [5-20]. The European LeukemiaNet (ELN) first recommended the risk classification system based on the cytogenetic and genetic status in 2010 (ELN-2010) [2]. In this system, risk categories were divided into four groups; favorable-risk (FR), intermediate-I-risk

(IR-I), intermediate-II-risk (IR-II) and adverse-risk (AR). It was a landmark in the genetic risk stratification of CN-AML that patients could be divided into two groups according to the mutation status of *NPM1*, *FLT3*-ITD and *CEBPA*. Although retrospective analysis demonstrated that the ELN-2010 was useful for further risk stratification of younger adult patients with CN-AML [21, 22], the accumulation of information on the prognostic relevance of recurrent genetic alterations have required the modification by including further genetic status [5, 23].

Recently, the ELN published the revised risk stratification system for AML (ELN-2017), in which AML is divided into three risk categories (Favorable, Intermediate and Adverse) rather than the previous 4-category system [24]. In the ELN-2017 system, several modifications have been made; biallelic mutated *CEBPA* is considered as favorable risk, allelic ratio of *FLT3*-ITD is considered for the risk stratification, cytogenetic abnormality is excluded for stratification into favorable risk in patients with *NPM1* or biallelic *CEBPA* mutations, and *RUNX1*, *ASXL1* and *TP53* mutations, and monosomal karyotype are additionally included in the adverse risk category. In this study, we evaluated the usefulness of the ELN-

2017 risk stratification system in Japanese AML patients, who were registered in the Japan Adult Leukemia Study Group (JALSG) AML201 study in comparison with the ELN-2010 and refined MRC systems.

## 2. Patients and Methods

### 2.1. Patients and treatment

The JALSG AML201 study was a multi-center phase 3 randomized study for newly diagnosed *de novo* adult AML patients, except for those with acute promyelocytic leukemia (UMIN Clinical Trials Registry C000000157, <http://www.umin.ac.jp/ctrj/>) [25, 26]. Detailed protocol is presented in Supplemental information.

Morphological diagnosis, the French-American-British (FAB) classification and karyotypes were reviewed and confirmed by the central review committees of the JALSG using the bone marrow (BM) samples obtained at diagnosis. The diagnosis of AML was based on the classification [27]. The AML201 study included 1057 patients, of whom 197 patients were available for the comprehensive genetic analysis, and their clinical and genetic data were used for this study.



We obtained informed consent from all patients to use their clinical data and their samples for banking and molecular analysis, and approval was obtained from the ethics committees of the participating institutes.

## *2.2 Cytogenetic and molecular analysis*

Cytogenetic G-banding analysis was performed using standard methods. We also examined 11 chimeric gene transcripts (Major *BCR-ABL1*, Minor *BCR-ABL1*, *PML-RARA*, *RUNX1-RUNX1T1*, *CBFB-MYH11*, *DEK-NUP214*, *NUP98-HOXA9*, *MLLT1-KMT2A*, *MLLT2-KMT2A*, *MLLT3-KMT2A*, *MLLT4-KMT2A*) by reverse transcriptase-mediated quantitative PCR (RQ-PCR) as previously reported [28].

Mutation analysis and results were reported previously [29]. To determine the allelic ratio of *FLT3*-ITD, exons 14 and 15 of the *FLT3* gene were amplified from DNA by PCR using a fluorescently labeled primer, and the products were analyzed by fragment analysis on the Genetic Analyzer 3500 (Applied

Biosystems, Foster City, CA).

### **3. Statistical analysis**

Differences in continuous variables were analyzed by the Kruskal-Wallis test. Analysis of frequencies was performed using Pearson's  $\chi^2$  test. Survival probabilities were estimated by the Kaplan-Meier method, and differences in the survival distributions were evaluated using the log-rank test. OS was defined as the time from the date of entry into the AML201 study to death due to any cause or last follow-up. The prognostic significance of the clinical variables was assessed using the Cox proportional hazards model. These statistical analyses were performed with Stata version 13.1 (StataCorp, College Station, TX). For all analyses, the *P*-values were two-tailed, and a *P*-value of less than 0.05 was considered significant.

## 4. Results

### *4.1. Risk stratification according to the 2017 ELN recommendation*

According to the ELN-2017 criteria, favorable, intermediate and adverse categories comprised 108 (54.8%), 43 (21.8%) and 46 (23.4%) patients, respectively (Table 1). In the ELN-2010 criteria, FR, IR-I, IR-II and AR consisted of 92 (47%), 35 (18%), 42 (21%) and 28 (14%) patients, respectively (Table 2), indicating that many patients were re-categorized into favorable or adverse risk groups in the ELN-2017 criteria. Based on the G-banding karyotype and chimeric transcript analyses, patients were assigned to favorable- (n=55, 28 %), intermediate- (n=119, 60 %) or adverse-risk (n=23, 12 %) groups according to the refined MRC criteria [4]. Patient distributions according to the refined MRC, ELN-2010 and ELN-2017 criteria are shown in Figure 1.

The ELN-2017 favorable group consisted of 90 FR, 6 IR-I and 12 IR-II patients according to the ELN-2010 criteria (Figure 2). All IR-I patients, who

were re-categorized into the favorable group in the ELN-2017, had mutated *NPM1* with *FLT3*-ITD<sup>low</sup>. Of 12 IR-II patients who were re-categorized into the favorable group, nine patients had mutated *NPM1* without *FLT3*-ITD and three had biallelic mutated *CEBPA*; however, all patients had cytogenetic abnormalities. The intermediate group consisted of 2 FR, 19 IR-I and 22 IR-II patients according to the ELN-2010 system (Figure 2). All FR patients who were re-categorized into the intermediate group in the ELN-2017 system had monoallelic mutated *CEBPA*. Of 19 IR-I patients who were re-categorized into the intermediate group, 10 patients had wild-type *NPM1* without *FLT3*-ITD, seven had wild-type *NPM1* with *FLT3*-ITD<sup>low</sup> and two had mutated *NPM1* with *FLT3*-ITD<sup>high</sup>. Of 22 IR-II patients who were re-categorized into the intermediate group, 12 patients had cytogenetic abnormalities not classified as favorable or adverse, nine had wild-type *NPM1* without *FLT3*-ITD or with *FLT3*-ITD<sup>low</sup> and one had *MLLT3-KMT2A*. The adverse group consisted of 10 IR-I, 8 IR-II and 28 AR patients according to the ELN-2010 system (Figure 2). Of 10 IR-I patients who were re-categorized into the adverse group, three patients had wild-type *NPM1* with *FLT3*-

ITD<sup>high</sup>, six patients had mutated *RUNX1*, and one patient had mutated *ASXL1*.

Of eight IR-II patients who were re-categorized into the adverse group, four patients had mutated *RUNX1*, two had monosormal karyotype, and two had mutated *ASXL1*.

#### *4.2. Patient characteristics according to the ELN-2017 categories*

Patient characteristics according to the ELN-2017 system are listed in Table

3. There were no significant differences in age distribution and WBC counts among three categories. As the favorable risk category includes CBF-AML, the FAB M2 and M4 subtypes were frequently observed in this category. Of the 197 patients, 98 and 99 patients were assigned to IDR or HiDNR arms for induction therapy, respectively. Allo-SCT was conducted for 105 patients; 23 patients at the first CR and 82 after the first relapse.

#### *4.3. Prognostic analysis according to the ELN-2017 system*

CR was achieved in 161 of 197 (81.7%) patients, and 80 and 77 patients were assigned to HiDAC or conventional consolidation therapies, respectively. The CR rate was significantly higher in the favorable risk groups (102/108; 94.4%) than in the intermediate (28/43; 65.1%) and adverse (31/46; 67.4%) groups (Table 4). Notably, 91 of the 102 (89.2%) patients in the favorable risk group achieved CR by one course of induction therapy, whereas 10 of the 28 (35.7%) in the intermediate risk and 9 of the 31 (29.0%) in the adverse risk groups required two courses of induction therapy. In the ELN-2010 criteria, the CR rates in the FR, IR-I, IR-II and AR groups were 93.5%, 77.1%, 69.0% and 67.9%, respectively. In the refined MRC criteria, the CR rates in the favorable, intermediate and adverse groups were 90.9%, 79.8% and 69.6%, respectively. These results indicated that the ELN-2017 system more clearly distinguished the risk groups for achieving CR.

Median follow-up time was 32.5 months for the analyzed 197 patients. Kaplan-Meier analyses for OS according to the refined MRC, ELN-2010 and ELN-2017 categories are shown in Figure 3. OSs at 5 years in the favorable,

intermediate and adverse groups with the ELN-2017 were 59.1% (95% CI, 47.7 - 68.8%), 32.6% (95% CI, 14.9 – 51.7%) and 22.6% (95% CI, 11.6 - 35.8 %), respectively (Figure 3A). On the other hand, OSs at 5 years in the FR, IR-I, IR-II and AR groups with the ELN-2010 were 64.8% (95% CI, 52.3 - 74.8%), 17.8% (95% CI, 6.1 – 34.5%), 38.1% (95% CI, 20.7 - 55.4%) and 24.1% (95% CI, 10.2 – 41.2%), respectively (Figure 3B). Therefore, the 3-risk category system of the ELN-2017 could clearly discriminate the OS in our cohort in comparison with the 4-risk category of the ELN-2010. However, the OS in the favorable group with the ELN-2017 was lower than that with the ELN-2010. We, therefore, compared the prognosis according to the ELN-2010 categories with each ELN-2017 category. In the favorable group with the ELN-2017, OS was significantly different among the FR, IR-I and IR-II groups with the ELN-2010 ( $P<0.0001$ ) (Figure 4A). Particularly, there was a significant difference between the FR and IR-I groups ( $P<0.0001$ ). All IR-I patients had mutated *NPM1* with *FLT3*-ITD<sup>low</sup>, and IR-II patients, all of whom were not cytogenetically normal and had mutated *NPM1* without *FLT3*-ITD or biallelic mutated *CEBPA*. These results indicated

that cytogenetic abnormalities may affect the prognosis of patients with mutated *NPM1* without *FLT3*-ITD and biallelic mutated *CEBPA* patients. However, further analysis is required to confirm this difference because of the low number of patients in the IR-I and IR-II groups.

Furthermore, from these results, we questioned whether the *FLT3*-ITD allelic ratio affected the prognosis in our cohort. We compared the prognosis of patients with *FLT3*-ITD according to the allelic ratio, but did not find any significant differences between *FLT3*-ITD<sup>high</sup> and *FLT3*-ITD<sup>low</sup> patients (Figure 5A). In addition, there were no significant differences in patients with CN-AML and those with wild-type and mutated *NPM1* (Figure 5B, C and D).

In the intermediate risk group with the ELN-2017, the patients categorized in the FR of the ELN-2010, all of whom were CN-AML with single mutated *CEBPA*, exhibited a better prognosis than those in the IR-I and IR-II groups, but a significant difference was not observed because of the small number of patients (Figure 4B). In the adverse risk group with the ELN-2017, there was no significant difference in OS among the IR-I, IR-II and AR groups with the ELN-



2010 (Figure 4C).

OSs at 5-yrs in the favorable, intermediate and adverse groups according to the refined MRC system were 71.0% (95% CI, 56.4 - 81.6%), 38.7% (95% CI, 28.3 - 49.0%), and 17.4% (95% CI, 5.4 - 35.0%), respectively (Figure 3C). These results indicated that favorable and adverse risk cytogenetics should be basically considered for risk stratification of adult AML.

In addition to the ELN and MRC systems, the Dutch–Belgian Cooperative Trial Group for Hematology/Oncology (HOVON) and the Southwest Oncology Group (SWOG) have also recommended risk stratification systems [30, 31]. Unfortunately, we could not evaluate the prognostic impact of the HOVON system on our cohort, because we did not examine all genetic status included in the HOVON system. OSs at 5-yrs in the favorable, intermediate and adverse groups according to the SWOG system were 67.2% (95% CI, 51.7 - 78.6%), 37.8% (95% CI, 26.1 - 49.4%), and 35.9% (95% CI, 21.1 - 51.0%), respectively (Supplemental Figure 1). Therefore, the SWOG system did not distinguish the prognosis of intermediate and adverse groups in our cohort.

We finally compared prognostic impacts of each risk category based on the ELN-2017, ELN-2010, refined MRC and SWOG systems. As shown in Table 5, the ELN-2017 system was more clearly distinguish the prognosis of intermediate and adverse risk groups than the ELN-2010 system in our cohort. However, the refined MRC system was also useful for the risk stratification.

#### *4.4. Association of mutations with therapeutic regimens*

In the JALSG AML201 study, patients were randomized to receive either the standard dose of IDR + Ara-C or HiDNR + Ara-C induction therapy, and the CR patients were again randomized to receive either three courses of HiDAC or four courses of conventional standard-dose multiagent consolidation therapy. Therefore, we analyzed whether the therapeutic regimens affected the CR rate, OS and DFS according to the ELN-2017 system, but no significant differences were observed among them.

In our cohort, 105 patients underwent allo-SCT during the treatments. We also evaluated the prognosis of each risk category when allo-SCT was censored. Although the ELN-2017 and the refined MRC systems could still clearly discriminate OSs even if allo-SCT was censored, the ELN-2010 system could not distinguish prognosis among IR-I, IR-II and AR groups (Figure 3).

## 5. Discussion

Prognostic risk assessment is the most important step in providing AML patients with an appropriate therapy. Although many risk factors for prognosis have been identified in AML patients, genetic alterations greatly affect the therapeutic strategies for patients who are eligible for intensive chemotherapy. In this study, we stratified the AML patients into each category according to the refined MRC, ELN-2010 and ELN-2017 risk categories, and validated each system for clinical application.

Since CBF-AML is frequently identified in Japanese patients, particularly in younger adults, our cohort included higher number of favorable risk patients according to the refined MRC system. To more precisely stratify the intermediate risk groups, particularly the CN-patients, the ELN-2010 included the mutation status of *FLT3*, *NPM1* and *CEBPA* genes, and stratified the patients into four risk groups. The ELN-2010 system could separate the favorable risk groups from the CN-AML patients; however, it has been reported that the long-

term prognosis in the IR-I group was not distinguishable from that in the IR-II and/or the AR groups [22]. We also reported that the long-term prognosis in the IR-I group was almost the same as that in the AR group in our cohort [29]. As we previously reported, *DNMT3A* and *RUNX1* mutations, and partial tandem duplication of the *MLL* gene (*MLL*-PTD) were identified as poor prognostic factors for OS in our cohort [29]. In the 35 IR-I group patients, 11 and six patients harbored *DNMT3A* mutation and *MLL*-PTD, respectively, and three patients harbored both mutations. Particularly, *MLL*-PTD was the poor prognostic factor also in the IR-I group [29]. Therefore, this mutation status reduced the OS of the FR-I group. Furthermore, *DNMT3A* mutation and *MLL*-PTD are not included in the ELN-2017 system. Multivariate analysis including these mutations and the adverse risk of the ELN-2017 showed that these were independent poor prognostic factors for OS (Table 6). Further analysis in a large-scale cohort is necessary to confirm the prognostic effects of these mutations in Japanese AML patients.

Although the ELN-2010 system could select favorable risk patients from the cytogenetically intermediate risk group, the AR groups did not increase because genetic status was not considered for the AR category. The adverse risk group with the ELN-2017 system additionally includes *RUNX1*, *ASXL1* and *TP53* mutations, and monosomal karyotype, resulting in the increase of this group to 23.4% from 14.2% in our cohort. As we previously confirmed the poor prognosis of patients with these genetic abnormalities in our cohort, the adverse risk group with the ELN-2017 is more clearly distinguished from the intermediate risk group than with the ELN-2010.

In the intermediate risk group with the ELN-2017, the patients categorized into the FR group with the ELN-2010 showed a better prognosis than those in the IR-I and IR-II groups (Figure 4B). All patients categorized in the FR group with the ELN-2010 are CN-AML with single mutated *CEBPA*. It has been reported that the single mutated *CEBPA* patients frequently acquire other genetic mutations [32]; however, those in our cohort did not have other mutations

associated with poor prognosis. Further studies are required to evaluate the prognostic relevance of single *CEBPA* mutation in patients with CN-AML.

The most controversial issue in our cohort was the genetic category for stratification into the favorable risk group with the ELN-2017. As shown in Figure 4A, the OS of the patients in the favorable risk group with the ELN-2017 was significantly different among the patients categorized into the FR, IR-I and IR-II groups with the ELN-2010. The IR-II patients were re-categorized into the favorable group with the ELN-2017 because the ELN-2017 does not consider cytogenetic abnormalities. As indicated above, the prognosis of CN-AML patients with single *CEBPA* mutation seems better in our cohort. These results indicated that the prognostic implications of the normal karyotype may be more precisely evaluated; however, there is a limitation in that cytogenetic abnormalities cannot be completely avoided using the conventional G-banding method, indicating that novel methods, such as next generation sequencing, may be necessary for evaluating the cytogenetic effects on the prognosis of AML patients. Of note is the poor prognosis of the IR-I patients who were categorized

into the favorable risk group with the ELN-2017. These patients were categorized into the favorable risk group because of the low *FLT3*-ITD allelic ratio. As shown in Figure 5, we were unable to distinguish the prognosis of patients with *FLT3*-ITD based on the allelic ratio. Although we analyzed the prognostic effects of *FLT3*-ITD ratio in other patients who were treated in the JALSG AML-87, -89 and -92 studies, we found no prognostic relevance for the *FLT3*-ITD allelic ratio (data not shown) [33]. At present, it is not clear why *FLT3*-ITD allelic ratio did not affect the prognosis in Japanese adult patients; however, this should be re-evaluated in patients treated with FLT3 inhibitors, because the combination of chemotherapy and a FLT3 inhibitor, midostaurin, reportedly improved the prognosis of the AML patients with *FLT3* mutation [34].

In conclusion, we demonstrated that the ELN-2017 risk stratification system for AML clearly distinguished long-term prognosis in Japanese adult patients with *de novo* AML. However, there are still controversial genetic categories in the favorable and intermediate risk groups. Further studies are required to confirm their prognostic relevance in Japanese AML patients.



## **Contributors**

Y.H., Y.I., H. Kiyoi, S. Ogawa, I.M., Y.M. and T.N. designed the study and interpreted the data; Y.H., Y.I. and H. Kiyoi wrote the manuscript; Y.H., Y.I., R.K. and Y.N. performed molecular analysis and interpreted the data; N.A., S. Ohtake, S.M., Y.M., T.S., Y.O., N.U., H. Kanamori, Y.I., K.I., Y.S., S.K., K.K., E.S., M.O., A.T., F.I., H.S., Y.K. and I.M. collected samples and clinical data, contributed to the interpretation of the data, and critically reviewed the manuscript; and all authors approved the final version submitted for publication.

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## References

- [1] E. Estey, H. Dohner, Acute myeloid leukaemia, *Lancet* 368(9550) (2006) 1894-907.
- [2] H. Dohner, E.H. Estey, S. Amadori, F.R. Appelbaum, T. Buchner, A.K. Burnett, H. Dombret, P. Fenaux, D. Grimwade, R.A. Larson, F. Lo-Coco, T. Naoe, D. Niederwieser, G.J. Ossenkoppele, M.A. Sanz, J. Sierra, M.S. Tallman, B. Lowenberg, C.D. Bloomfield, Diagnosis and management of acute myeloid leukemia in adults: recommendations from an international expert panel, on behalf of the European LeukemiaNet, *Blood* 115(3) (2010) 453-74.
- [3] D. Grimwade, H. Walker, F. Oliver, K. Wheatley, C. Harrison, G. Harrison, J. Rees, I. Hann, R. Stevens, A. Burnett, A. Goldstone, The importance of diagnostic cytogenetics on outcome in AML: analysis of 1,612 patients entered into the MRC AML 10 trial. The Medical Research Council Adult and Children's Leukaemia Working Parties, *Blood* 92(7) (1998) 2322-33.
- [4] D. Grimwade, R.K. Hills, A.V. Moorman, H. Walker, S. Chatters, A.H. Goldstone, K. Wheatley, C.J. Harrison, A.K. Burnett, Refinement of cytogenetic classification in acute myeloid leukemia: determination of prognostic significance of rare recurring chromosomal abnormalities among 5876 younger adult patients treated in the United Kingdom Medical Research Council trials, *Blood* 116(3) (2010) 354-65.
- [5] J.P. Patel, M. Gonen, M.E. Figueroa, H. Fernandez, Z. Sun, J. Racevskis, P. Van Vlierberghe, I. Dalgalev, S. Thomas, O. Aminova, K. Huberman, J. Cheng, A. Viale, N.D. Socci, A. Heguy, A. Cherry, G. Vance, R.R. Higgins, R.P. Ketterling, R.E. Gallagher, M. Litzow, M.R. van den Brink, H.M. Lazarus, J.M. Rowe, S. Luger, A. Ferrando, E. Paietta, M.S. Tallman, A. Melnick, O. Abdel-Wahab, R.L. Levine, Prognostic relevance of integrated genetic profiling in acute myeloid leukemia, *N Engl J Med* 366(12) (2012) 1079-89.
- [6] Y. Shen, Y.M. Zhu, X. Fan, J.Y. Shi, Q.R. Wang, X.J. Yan, Z.H. Gu, Y.Y. Wang, B. Chen, C.L. Jiang, H. Yan, F.F. Chen, H.M. Chen, Z. Chen, J. Jin, S.J. Chen, Gene mutation patterns and their prognostic impact in a cohort of 1185 patients with acute myeloid leukemia, *Blood* 118(20) (2011) 5593-603.
- [7] Y. Ofran, J.M. Rowe, Genetic profiling in acute myeloid leukaemia--where are we and what is its role in patient management, *Br J Haematol* 160(3) (2013) 303-20.

- [8] F. Delhommeau, S. Dupont, V. Della Valle, C. James, S. Trannoy, A. Masse, O. Kosmider, J.P. Le Couedic, F. Robert, A. Alberdi, Y. Lecluse, I. Plo, F.J. Dreyfus, C. Marzac, N. Casadevall, C. Lacombe, S.P. Romana, P. Dessen, J. Soulier, F. Viguie, M. Fontenay, W. Vainchenker, O.A. Bernard, Mutation in TET2 in myeloid cancers, *N Engl J Med* 360(22) (2009) 2289-301.
- [9] S.M. Langemeijer, R.P. Kuiper, M. Berends, R. Knops, M.G. Aslanyan, M. Massop, E. Stevens-Linders, P. van Hoogen, A.G. van Kessel, R.A. Raymakers, E.J. Kamping, G.E. Verhoef, E. Verburch, A. Hagemeijer, P. Vandenberghe, T. de Witte, B.A. van der Reijden, J.H. Jansen, Acquired mutations in TET2 are common in myelodysplastic syndromes, *Nat Genet* 41(7) (2009) 838-42.
- [10] E.R. Mardis, L. Ding, D.J. Dooling, D.E. Larson, M.D. McLellan, K. Chen, D.C. Koboldt, R.S. Fulton, K.D. Delehaunty, S.D. McGrath, L.A. Fulton, D.P. Locke, V.J. Magrini, R.M. Abbott, T.L. Vickery, J.S. Reed, J.S. Robinson, T. Wylie, S.M. Smith, L. Carmichael, J.M. Eldred, C.C. Harris, J. Walker, J.B. Peck, F. Du, A.F. Dukes, G.E. Sanderson, A.M. Brummett, E. Clark, J.F. McMichael, R.J. Meyer, J.K. Schindler, C.S. Pohl, J.W. Wallis, X. Shi, L. Lin, H. Schmidt, Y. Tang, C. Haipek, M.E. Wiechert, J.V. Ivy, J. Kalicki, G. Elliott, R.E. Ries, J.E. Payton, P. Westervelt, M.H. Tomasson, M.A. Watson, J. Baty, S. Heath, W.D. Shannon, R. Nagarajan, D.C. Link, M.J. Walter, T.A. Graubert, J.F. DiPersio, R.K. Wilson, T.J. Ley, Recurring mutations found by sequencing an acute myeloid leukemia genome, *N Engl J Med* 361(11) (2009) 1058-66.
- [11] T.J. Ley, L. Ding, M.J. Walter, M.D. McLellan, T. Lamprecht, D.E. Larson, C. Kandoth, J.E. Payton, J. Baty, J. Welch, C.C. Harris, C.F. Lichti, R.R. Townsend, R.S. Fulton, D.J. Dooling, D.C. Koboldt, H. Schmidt, Q. Zhang, J.R. Osborne, L. Lin, M. O'Laughlin, J.F. McMichael, K.D. Delehaunty, S.D. McGrath, L.A. Fulton, V.J. Magrini, T.L. Vickery, J. Hundal, L.L. Cook, J.J. Conyers, G.W. Swift, J.P. Reed, P.A. Alldredge, T. Wylie, J. Walker, J. Kalicki, M.A. Watson, S. Heath, W.D. Shannon, N. Varghese, R. Nagarajan, P. Westervelt, M.H. Tomasson, D.C. Link, T.A. Graubert, J.F. DiPersio, E.R. Mardis, R.K. Wilson, DNMT3A mutations in acute myeloid leukemia, *N Engl J Med* 363(25) (2010) 2424-33.
- [12] G. Nikoloski, S.M. Langemeijer, R.P. Kuiper, R. Knops, M. Massop, E.R. Tonnissen, A. van der Heijden, T.N. Scheele, P. Vandenberghe, T. de Witte, B.A. van der Reijden, J.H. Jansen, Somatic mutations of the histone methyltransferase gene EZH2 in myelodysplastic syndromes, *Nat Genet* 42(8) (2010) 665-7.

- [13] A.H. Shih, O. Abdel-Wahab, J.P. Patel, R.L. Levine, The role of mutations in epigenetic regulators in myeloid malignancies, *Nat Rev Cancer* 12(9) (2012) 599-612.
- [14] W.C. Chou, H.H. Huang, H.A. Hou, C.Y. Chen, J.L. Tang, M. Yao, W. Tsay, B.S. Ko, S.J. Wu, S.Y. Huang, S.C. Hsu, Y.C. Chen, Y.N. Huang, Y.C. Chang, F.Y. Lee, M.C. Liu, C.W. Liu, M.H. Tseng, C.F. Huang, H.F. Tien, Distinct clinical and biological features of de novo acute myeloid leukemia with additional sex comb-like 1 (ASXL1) mutations, *Blood* 116(20) (2010) 4086-94.
- [15] V. Grossmann, E. Tiacci, A.B. Holmes, A. Kohlmann, M.P. Martelli, W. Kern, A. Spanhol-Rosseto, H.U. Klein, M. Dugas, S. Schindela, V. Trifonov, S. Schnittger, C. Haferlach, R. Bassan, V.A. Wells, O. Spinelli, J. Chan, R. Rossi, S. Baldoni, L. De Carolis, K. Goetze, H. Serve, R. Peceny, K.A. Kreuzer, D. Oruzio, G. Specchia, F. Di Raimondo, F. Fabbiano, M. Sborgia, A. Liso, L. Farinelli, A. Rambaldi, L. Pasqualucci, R. Rabadan, T. Haferlach, B. Falini, Whole-exome sequencing identifies somatic mutations of BCOR in acute myeloid leukemia with normal karyotype, *Blood* 118(23) (2011) 6153-63.
- [16] M. Li, R. Collins, Y. Jiao, P. Ouillette, D. Bixby, H. Erba, B. Vogelstein, K.W. Kinzler, N. Papadopoulos, S.N. Malek, Somatic mutations in the transcriptional corepressor gene BCORL1 in adult acute myelogenous leukemia, *Blood* 118(22) (2011) 5914-7.
- [17] J.S. Welch, T.J. Ley, D.C. Link, C.A. Miller, D.E. Larson, D.C. Koboldt, L.D. Wartman, T.L. Lamprecht, F. Liu, J. Xia, C. Kandoth, R.S. Fulton, M.D. McLellan, D.J. Dooling, J.W. Wallis, K. Chen, C.C. Harris, H.K. Schmidt, J.M. Kalicki-Veizer, C. Lu, Q. Zhang, L. Lin, M.D. O'Laughlin, J.F. McMichael, K.D. Delehaunty, L.A. Fulton, V.J. Magrini, S.D. McGrath, R.T. Demeter, T.L. Vickery, J. Hundal, L.L. Cook, G.W. Swift, J.P. Reed, P.A. Alldredge, T.N. Wylie, J.R. Walker, M.A. Watson, S.E. Heath, W.D. Shannon, N. Varghese, R. Nagarajan, J.E. Payton, J.D. Baty, S. Kulkarni, J.M. Kline, M.H. Tomasson, P. Westervelt, M.J. Walter, T.A. Graubert, J.F. DiPersio, L. Ding, E.R. Mardis, R.K. Wilson, The origin and evolution of mutations in acute myeloid leukemia, *Cell* 150(2) (2012) 264-78.
- [18] K. Yoshida, M. Sanada, Y. Shiraishi, D. Nowak, Y. Nagata, R. Yamamoto, Y. Sato, A. Sato-Otsubo, A. Kon, M. Nagasaki, G. Chalkidis, Y. Suzuki, M. Shiosaka, R. Kawahata, T. Yamaguchi, M. Otsu, N. Obara, M. Sakata-Yanagimoto, K. Ishiyama, H. Mori, F. Nolte, W.K. Hofmann, S. Miyawaki, S. Sugano, C. Haferlach, H.P. Koeffler, L.Y. Shih, T. Haferlach, S. Chiba, H. Nakauchi, S. Miyano, S. Ogawa, Frequent pathway mutations of splicing machinery in myelodysplasia, *Nature* 478(7367) (2011) 64-9.

- [19] T. Naoe, H. Kiyoi, Gene mutations of acute myeloid leukemia in the genome era, *International journal of hematology* 97(2) (2013) 165-74.
- [20] C. Mazumdar, R. Majeti, The role of mutations in the cohesin complex in acute myeloid leukemia, *International journal of hematology* 105(1) (2017) 31-36.
- [21] C. Rollig, M. Bornhauser, C. Thiede, F. Taube, M. Kramer, B. Mohr, W. Aulitzky, H. Bodenstern, H.J. Tischler, R. Stuhlmann, U. Schuler, F. Stolz, M. von Bonin, H. Wandt, K. Schafer-Eckart, M. Schaich, G. Ehninger, Long-term prognosis of acute myeloid leukemia according to the new genetic risk classification of the European LeukemiaNet recommendations: evaluation of the proposed reporting system, *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 29(20) (2011) 2758-65.
- [22] K. Mrozek, G. Marcucci, D. Nicolet, K.S. Maharry, H. Becker, S.P. Whitman, K.H. Metzeler, S. Schwind, Y.Z. Wu, J. Kohlschmidt, M.J. Pettenati, N.A. Heerema, A.W. Block, S.R. Patil, M.R. Baer, J.E. Kolitz, J.O. Moore, A.J. Carroll, R.M. Stone, R.A. Larson, C.D. Bloomfield, Prognostic significance of the European LeukemiaNet standardized system for reporting cytogenetic and molecular alterations in adults with acute myeloid leukemia, *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 30(36) (2012) 4515-23.
- [23] K.H. Metzeler, K. Maharry, M.D. Radmacher, K. Mrozek, D. Margeson, H. Becker, J. Curfman, K.B. Holland, S. Schwind, S.P. Whitman, Y.Z. Wu, W. Blum, B.L. Powell, T.H. Carter, M. Wetzler, J.O. Moore, J.E. Kolitz, M.R. Baer, A.J. Carroll, R.A. Larson, M.A. Caligiuri, G. Marcucci, C.D. Bloomfield, TET2 mutations improve the new European LeukemiaNet risk classification of acute myeloid leukemia: a Cancer and Leukemia Group B study, *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 29(10) (2011) 1373-81.
- [24] H. Dohner, E. Estey, D. Grimwade, S. Amadori, F.R. Appelbaum, T. Buchner, H. Dombret, B.L. Ebert, P. Fenaux, R.A. Larson, R.L. Levine, F. Lo-Coco, T. Naoe, D. Niederwieser, G.J. Ossenkoppele, M. Sanz, J. Sierra, M.S. Tallman, H.F. Tien, A.H. Wei, B. Lowenberg, C.D. Bloomfield, Diagnosis and management of AML in adults: 2017 ELN recommendations from an international expert panel, *Blood* 129(4) (2017) 424-447.
- [25] S. Ohtake, S. Miyawaki, H. Fujita, H. Kiyoi, K. Shinagawa, N. Usui, H. Okumura, K. Miyamura, C. Nakaseko, Y. Miyazaki, A. Fujieda, T. Nagai, T. Yamane, M. Taniwaki, M. Takahashi, F. Yagasaki, Y. Kimura, N. Asou, H. Sakamaki, H. Handa, S. Honda, K.

Ohnishi, T. Naoe, R. Ohno, Randomized study of induction therapy comparing standard-dose idarubicin with high-dose daunorubicin in adult patients with previously untreated acute myeloid leukemia: the JALSG AML201 Study, *Blood* 117(8) (2011) 2358-65.

[26] S. Miyawaki, S. Ohtake, S. Fujisawa, H. Kiyoi, K. Shinagawa, N. Usui, T. Sakura, K. Miyamura, C. Nakaseko, Y. Miyazaki, A. Fujieda, T. Nagai, T. Yamane, M. Taniwaki, M. Takahashi, F. Yagasaki, Y. Kimura, N. Asou, H. Sakamaki, H. Handa, S. Honda, K. Ohnishi, T. Naoe, R. Ohno, A randomized comparison of 4 courses of standard-dose multiagent chemotherapy versus 3 courses of high-dose cytarabine alone in postremission therapy for acute myeloid leukemia in adults: the JALSG AML201 Study, *Blood* 117(8) (2011) 2366-72.

[27] J.M. Bennett, D. Catovsky, M.T. Daniel, G. Flandrin, D.A. Galton, H.R. Gralnick, C. Sultan, Proposed revised criteria for the classification of acute myeloid leukemia. A report of the French-American-British Cooperative Group, *Annals of internal medicine* 103(4) (1985) 620-5.

[28] K. Osumi, T. Fukui, H. Kiyoi, M. Kasai, Y. Kodaera, K. Kudo, K. Kato, T. Matsuyama, K. Naito, M. Tanimoto, H. Hirai, H. Saito, R. Ohno, T. Naoe, Rapid screening of leukemia fusion transcripts in acute leukemia by real-time PCR, *Leuk Lymphoma* 43(12) (2002) 2291-9.

[29] R. Kihara, Y. Nagata, H. Kiyoi, T. Kato, E. Yamamoto, K. Suzuki, F. Chen, N. Asou, S. Ohtake, S. Miyawaki, Y. Miyazaki, T. Sakura, Y. Ozawa, N. Usui, H. Kanamori, T. Kiguchi, K. Imai, N. Uike, F. Kimura, K. Kitamura, C. Nakaseko, M. Onizuka, A. Takeshita, F. Ishida, H. Suzushima, Y. Kato, H. Miwa, Y. Shiraishi, K. Chiba, H. Tanaka, S. Miyano, S. Ogawa, T. Naoe, Comprehensive analysis of genetic alterations and their prognostic impacts in adult acute myeloid leukemia patients, *Leukemia* 28(8) (2014) 1586-95.

[30] M.L. Slovak, K.J. Kopecky, P.A. Cassileth, D.H. Harrington, K.S. Theil, A. Mohamed, E. Paietta, C.L. Willman, D.R. Head, J.M. Rowe, S.J. Forman, F.R. Appelbaum, Karyotypic analysis predicts outcome of preremission and postremission therapy in adult acute myeloid leukemia: a Southwest Oncology Group/Eastern Cooperative Oncology Group Study, *Blood* 96(13) (2000) 4075-83.

[31] F. Damm, M. Heuser, M. Morgan, K. Wagner, K. Gorlich, A. Grosshennig, I. Hamwi, F. Thol, E. Surdziel, W. Fiedler, M. Lubbert, L. Kanz, C. Reuter, G. Heil, R. Delwel, B. Lowenberg, P.J. Valk, J. Krauter, A. Ganser, Integrative prognostic risk score in acute

myeloid leukemia with normal karyotype, *Blood* 117(17) (2011) 4561-8.

[32] A. Fasan, C. Haferlach, T. Alpermann, S. Jeromin, V. Grossmann, C. Eder, S. Weissmann, F. Dicker, A. Kohlmann, S. Schindela, W. Kern, T. Haferlach, S. Schnittger, The role of different genetic subtypes of CEBPA mutated AML, *Leukemia* 28(4) (2014) 794-803.

[33] H. Kiyoi, T. Naoe, Y. Nakano, S. Yokota, S. Minami, S. Miyawaki, N. Asou, K. Kuriyama, I. Jinnai, C. Shimazaki, H. Akiyama, K. Saito, H. Oh, T. Motoji, E. Omoto, H. Saito, R. Ohno, R. Ueda, Prognostic implication of FLT3 and N-RAS gene mutations in acute myeloid leukemia, *Blood* 93(9) (1999) 3074-80.

[34] R.M. Stone, S.J. Mandrekar, B.L. Sanford, K. Laumann, S. Geyer, C.D. Bloomfield, C. Thiede, T.W. Prior, K. Dohner, G. Marcucci, F. Lo-Coco, R.B. Klisovic, A. Wei, J. Sierra, M.A. Sanz, J.M. Brandwein, T. de Witte, D. Niederwieser, F.R. Appelbaum, B.C. Medeiros, M.S. Tallman, J. Krauter, R.F. Schlenk, A. Ganser, H. Serve, G. Ehninger, S. Amadori, R.A. Larson, H. Dohner, Midostaurin plus Chemotherapy for Acute Myeloid Leukemia with a FLT3 Mutation, *N Engl J Med* 377(5) (2017) 454-464.



**Table 1. Distribution of genetic abnormalities according to the ELN-2017 risk stratification system**

Risk category	Genetic abnormality	Number (%)
Favorable	t(8;21)(q22;q22): <i>RUNX1-RUNX1T1</i>	41 (20.8)
	inv(16)(p13.1q22) or t(16;16)(p13.1;q22): <i>CBFB-MYH11</i>	14 (7.1)
	Mutated <i>NPM1</i> without <i>FLT3</i> -ITD or with <i>FLT3</i> -ITD <sup>low</sup>	36 (18.3)
	Biallelic mutated <i>CEBPA</i>	17 (8.6)
	Total	108 (54.8)
Intermediate	Mutated <i>NPM1</i> and <i>FLT3</i> -ITD <sup>high</sup>	2 (1.0)
	Wild type <i>NPM1</i> without <i>FLT3</i> -ITD or with <i>FLT3</i> -ITD <sup>low</sup>	28 (14.2)
	t(9;11)(p21.3;q23.3); <i>MLLT3-KMT2A</i>	1 (0.5)
	Cytogenetic abnormalities not classified as favorable or adverse	12 (6.1)
	Total	43 (21.8)
Adverse	t(6;9)(p23;q34): <i>DEK-NUP214</i>	3 (1.5)
	t(v;11)(v;q23): <i>KMT2A</i> rearranged	6 (3.0)
	t(9;22)(q34.1;q11.2); <i>BCR-ABL1</i>	2 (1.0)
	-7	1 (0.5)
	Complex karyotype	16 (8.1)
	monosormal karyotype	2 (1.0)
	Wild type <i>NPM1</i> and <i>FLT3</i> -ITD <sup>high</sup>	3 (1.5)
	Mutated <i>RUNX1</i>	10 (5.1)
	Mutated <i>ASXL1</i>	3 (1.5)
	Total	46 (23.4)

**Table 2. Distribution of genetic abnormalities according to the ELN-2010 risk stratification system**

Risk category	Genetic abnormality	Number (%)
Favorable	t(8;21)(q22;q22); <i>RUNX1-RUNX1T1</i>	41 (20.8)
	Inv(16)(p13.1q22) or t(16;16)(8p13.1;q22); <i>CBFB-MYH11</i>	14 (7.1)
	Mutated <i>NPM1</i> /Wild type <i>FLT3</i> (normal karyotype)	21 (10.7)
	Mutated <i>CEBPA</i> (normal karyotype)	16 (8.1)
	Total	92 (46.7)
Intermediate-I	Mutated <i>NPM1/FLT3</i> -ITD (normal karyotype)	8 (4.1)
	Wild type <i>NPM1/FLT3</i> -ITD (normal karyotype)	13 (6.6)
	Wild type <i>NPM1</i> /Wild type <i>FLT3</i> (normal karyotype)	14 (7.1)
	Total	35 (17.8)
Intermediate-II	t(9;11)(p21.3;q23.3); <i>MLLT3-KMT2A</i>	1 (0.5)
	Cytogenetic abnormalities not classified as favorable or adverse	41 (20.8)
	Total	42 (21.3)
Adverse	t(6;9)(q23;q34); <i>DEK-NUP214</i>	3 (1.5)
	t(v;11)(v;q23); <i>MLL</i> rearranged	6 (3.0)
	-7	1 (0.5)
	Complex karyotype	18 (9.1)
	Total	28 (14.2)

**Table 3. Characteristics of the 197 patients according to the ELN-2017 risk stratification system**

	Total (n=197)	Favorable (n=108)	Intermediate (n=43)	Adverse (n=46)	
	Number (%)	Number (%)	Number (%)	Number (%)	P-value
Age (yr)					0.222
15 - 19	6 (3.0%)	4 (3.7%)	1 (2.3%)	1 (2.2%)	
20 - 29	32 (16.2%)	14 (13.0%)	12 (27.9%)	6 (13.0%)	
30 - 39	35 (17.8%)	20 (18.5%)	7 (16.3%)	8 (17.4%)	
40 - 49	33 (16.8%)	22 (20.4%)	3 (7.0%)	8 (17.4%)	
50 - 59	69 (35.0%)	40 (37.0%)	15 (34.9%)	14 (30.4%)	
60 - 64	22 (11.2%)	8 (7.4%)	5 (11.6%)	9 (19.6%)	
FAB-type					0.008
M0	7 (3.6%)	0 (0%)	4 (9.3%)	3 (6.5%)	
M1	36 (18.3%)	13 (12.0%)	12 (27.9%)	11 (23.9%)	
M2	89 (45.2%)	59 (54.6%)	15 (34.9%)	15 (32.6%)	
M4	43 (21.8%)	26 (24.1%)	8 (18.6%)	9 (19.6%)	
M5	21 (10.7%)	10 (9.3%)	4 (9.3%)	7 (15.2%)	
M6	1 (0.5%)	0 (0%)	0 (0%)	1 (2.2%)	
Induction therapy					0.102
IDR + AraC	98 (49.7%)	61 (56.5%)	19 (44.2%)	18 (39.1%)	
DNR + AraC	99 (50.3%)	47 (43.5%)	24 (55.8%)	28 (60.9%)	
Consolidation therapy					0.797
High-dose Ara-C	80 (40.6%)	51 (47.2%)	15 (34.9%)	14 (30.4%)	
Multiagent CT	77 (39.1%)	50 (46.3%)	11 (25.6%)	16 (34.8%)	
WBC count (x 10 <sup>9</sup> /L)					0.797
median	17.2	16.6	19.7	13.0	
range	0.05 – 367	0.05 – 367	0.50 – 323	0.08 – 200	

**Table 4. Comparison of the CR rates in each risk category among the refined MRC, ELN-2010 and ELN-2017**

		Favorable	Intermediate		Adverse
		number	55	119	23
		CR	50 (90.9%)	95 (79.8%)	16 (69.6%)
Refined	No. of induction courses				
MRC	1	46 (83.6%)	72 (60.5%)	13 (56.5%)	
	2	4 (7.3%)	23 (19.3%)	3 (13.0%)	
	non CR	5 (9.1%)	24 (20.2%)	7 (30.4%)	
			Intermediate-I	Intermediate-II	
		number	92	35	42
		CR	86 (93.5%)	27 (77.1%)	29 (69.0%)
ELN-2010	No. of induction courses				
	1	76 (82.6%)	17 (48.6%)	24 (57.1%)	14 (50.0%)
	2	10 (10.9%)	10 (28.6%)	5 (11.9%)	5 (17.9%)
	non CR	6 (6.5%)	8 (22.9%)	13 (31.0%)	9 (32.1%)
		number	108	43	46
		CR	102 (94.4%)	28 (65.1%)	31 (67.4%)
ELN-2017	No. of induction courses				
	1	91 (84.3%)	18 (41.9%)	22 (47.8%)	
	2	11 (10.2%)	10 (23.3%)	9 (19.6%)	
	non CR	6 (5.6%)	15 (34.9%)	15 (32.6%)	

**Table 6. Prognostic impact of each category**

	HR	95% CI	<i>P</i> -value
ELN2017			
Favorable	0.362	0.241 – 0.543	<0.001
Intermediate	1.466	0.931 – 2.310	0.099
Adverse	2.632	1.736 – 3.988	<0.001
ELN2010			
Favorable	0.295	0.190 – 0.459	<0.001
Intermediate-I	2.404	1.539 – 3.755	<0.001
Intermediate-II	1.247	0.781 – 1.990	0.356
Adverse	2.371	1.461 – 3.850	<0.001
Refined MRC			
Favorable	0.359	0.207 – 0.622	<0.001
Intermediate	1.311	0.866 – 1.985	0.201
Adverse	2.947	1.779 – 4.881	<0.001
SWOG			
Favorable	0.438	0.257 – 0.749	0.003
Intermediate	1.271	0.856 – 1.887	0.235
Unfavorable	1.578	1.002 – 2.485	0.049
Unknown	1.186	0.482 – 2.920	0.710

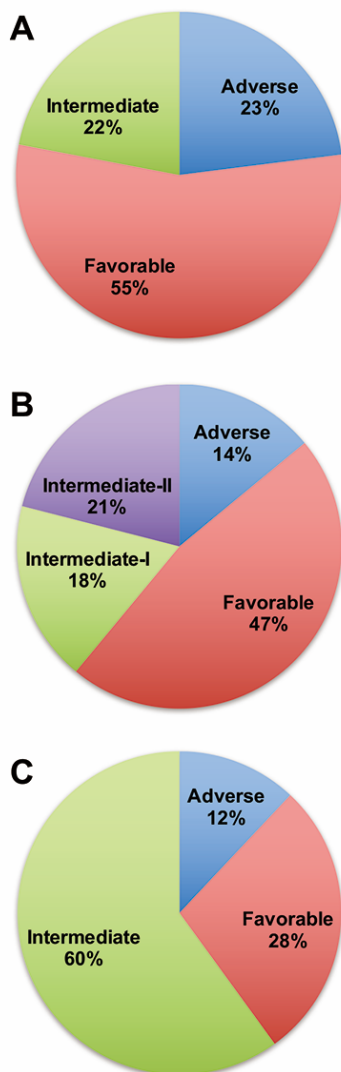
**Table 6. Multivariate analysis for overall survival**

Risk	HR	95% CI	<i>P</i> -value
ELN2017-Adverse	2.772	1.735 - 4.804	<0.001
<i>MLL-PTD</i>	2.450	1.188 - 5.050	0.015
<i>DNMT3A</i>	2.187	1.322 - 3.619	0.002

## Figure legends

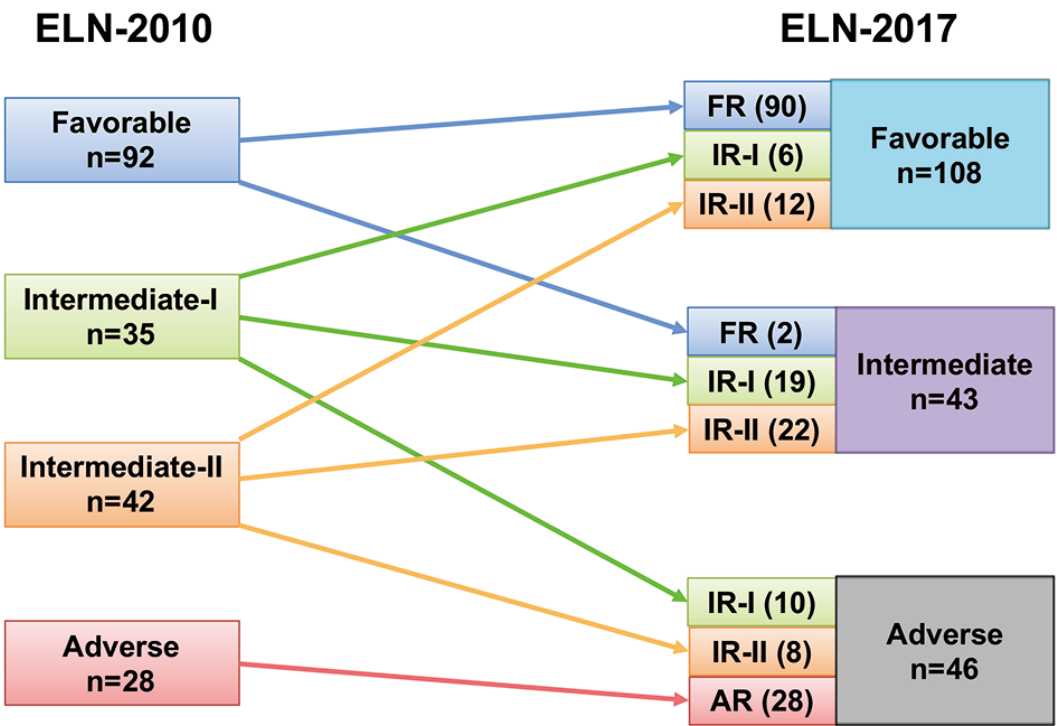
**Figure 1. Distribution of patients according to the refined MRC, ELN-2010 and ELN-2017 systems.**

Distribution of patients according to the refined MRC (A), ELN-2010 (B) and ELN-2017 (C). The patient numbers categorized into the favorable and adverse groups increased with the ELN-2017 system.



**Figure 2. Changes in the risk categories between the ELN-2010 and the ELN-2017.**

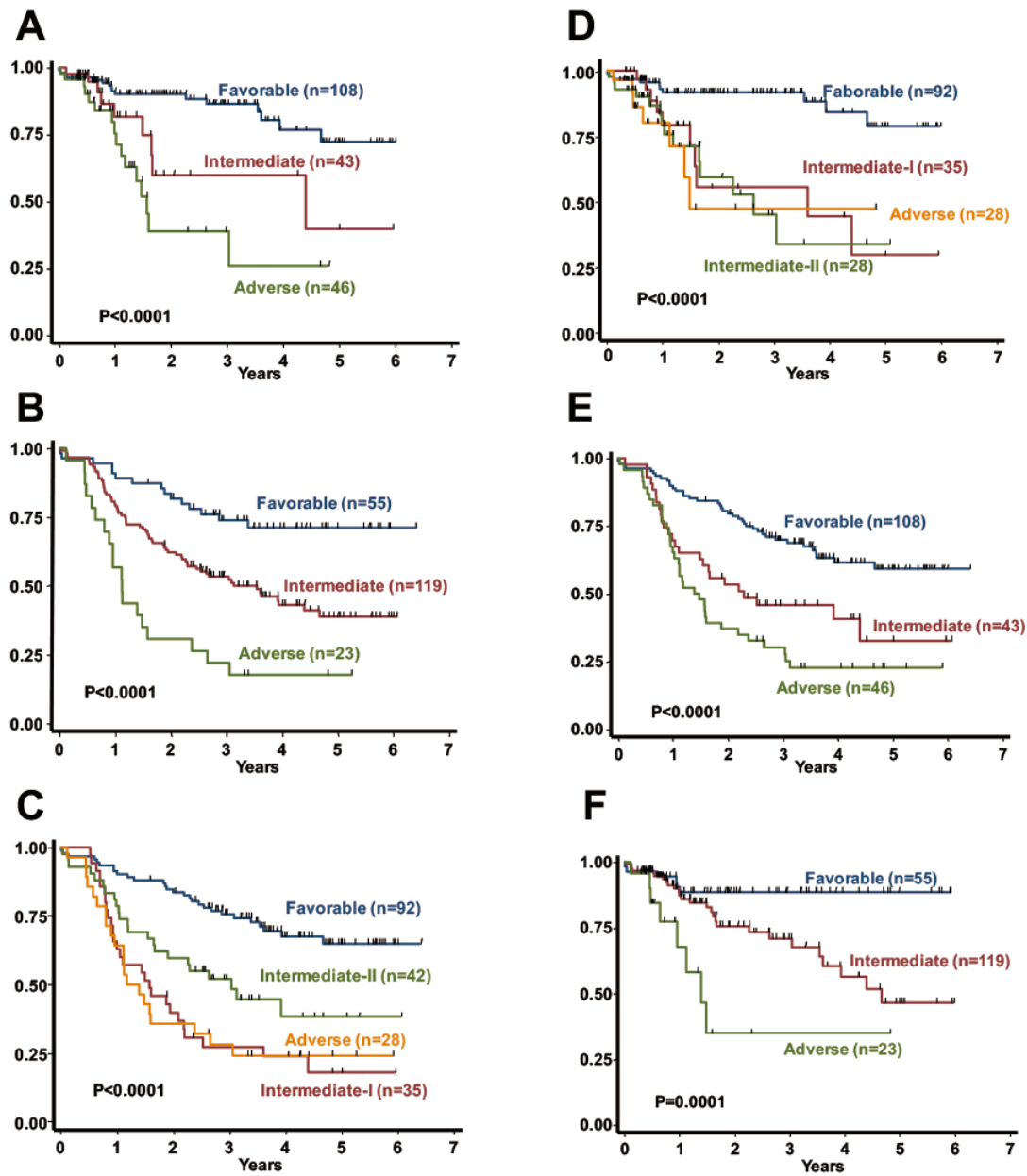
With the ELN-2017 system, the patient numbers in the favorable and adverse risk group increased by the changes in the risk categories based on genetic status.





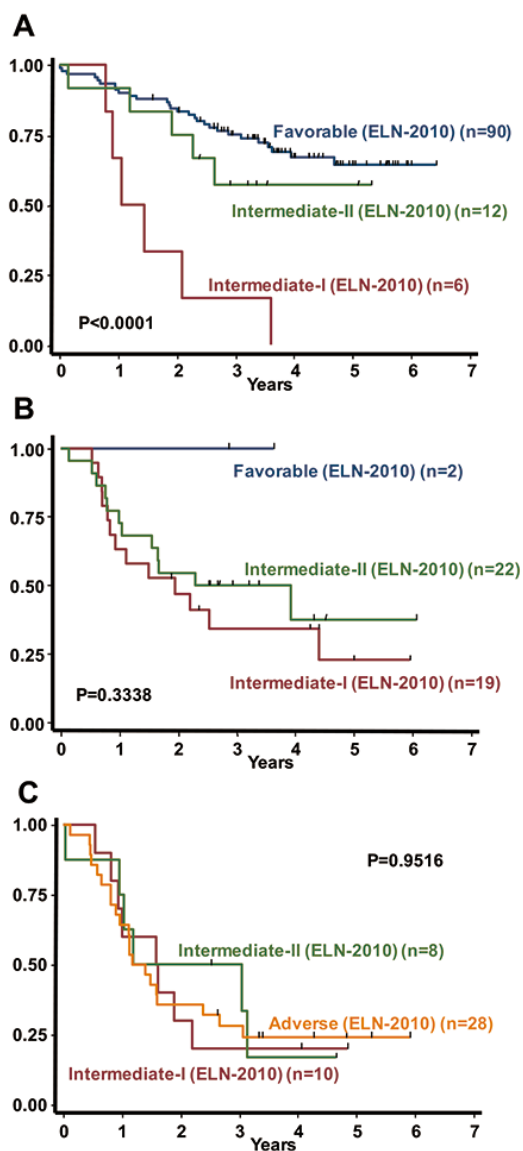
**Figure 3. Overall survivals according to the ELN-2017, ELN-2010 and refined MRC risk categories.**

Overall survivals according to the ELN-2017 (A, D), ELN-2010 (B, E) and refined MRC (C, F) risk categories are shown. D, E and F are results when allo-SCT are censored.



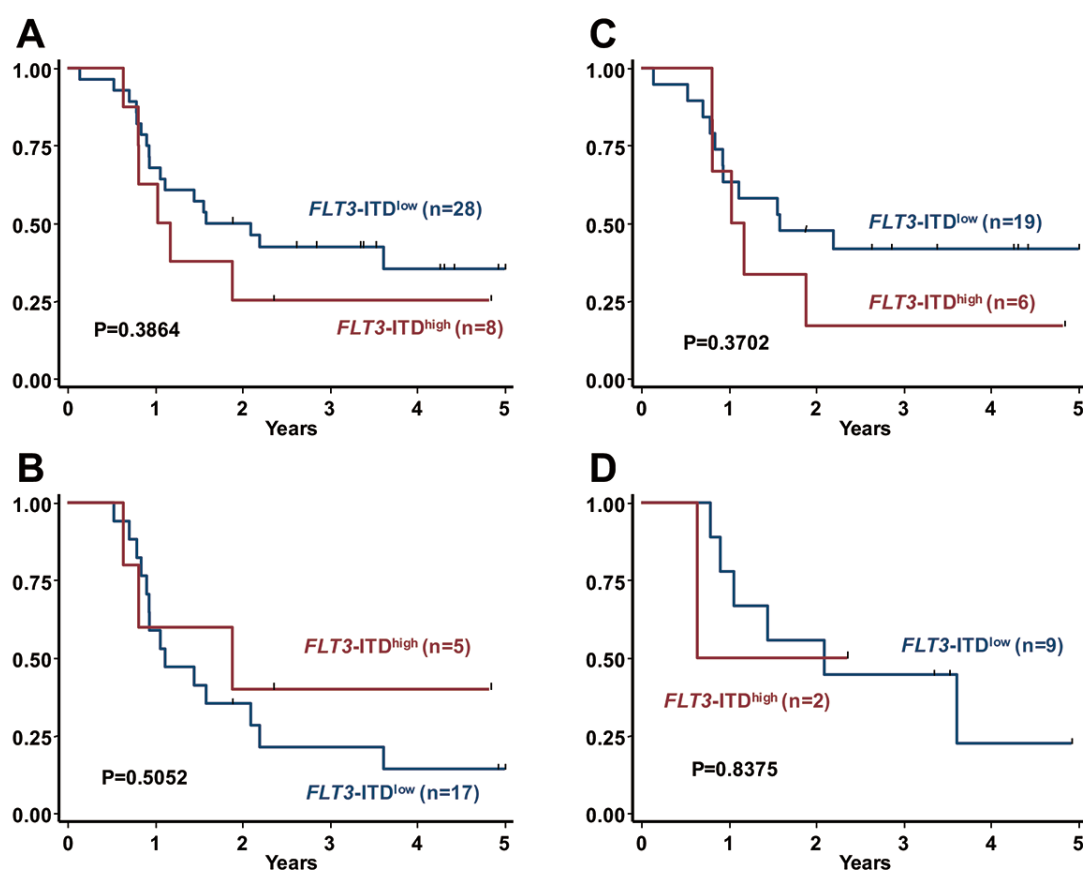
**Figure 4. Overall survivals according to the 2010 ELN risk categories in the 2017 ELN categories.**

(A) In the favorable risk groups, prognosis of the patients categorized into the IR-I and IR-II by the ELN-2010 was relatively poor. Particularly, there was a significant difference between the FR and IR-I groups ( $P<0.0001$ ). (B) In the intermediate group, prognosis of the patients categorized into the FR by the ELN-2010 is relatively better. (C) In the adverse group, there was no prognostic difference among the risk categories with the ELN-2010.



**Figure 5. Overall survivals according to the *FLT3*-ITD allelic ratio.**

The allelic ratio of *FLT3*-ITD did not affect the prognosis in patients with *FLT3*-ITD (A), those with CN-AML (B), those with wild-type *NPM1* (C) and those with mutated *NPM1* (D).

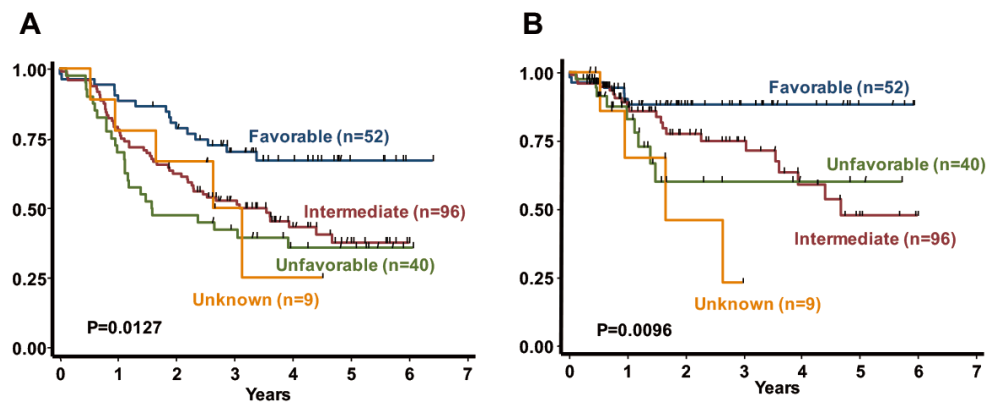


## Supplemental figure legend

### Supplemental Figure 1

Overall survivals according to the SWOG system.

(A) Allo-SCT was not censored. (B) Allo-SCT was censored.



## **Supplemental information**

### **Protocol of the JALSG AML201 study**

The patients were prospectively registered and randomly assigned to receive either idarubicin (IDR) (12 mg/m<sup>2</sup> daily for 3 days) or high-dose daunorubicin (HiDNR) (50 mg/m<sup>2</sup> daily for 5 days) in combination with 100 mg/m<sup>2</sup> of cytarabine (Ara-C) by continuous infusion daily for 7 days as induction therapy, and those who achieved CR were again randomized to receive either 4 courses of conventional consolidation therapy (course 1: mitoxantrone 7 mg/m<sup>2</sup> days 1-3 and Ara-C 200 mg/m<sup>2</sup> by 24-hour continuous infusion days 1-5; course 2 daunorubicin 50 mg/m<sup>2</sup> days 1-3 and Ara-C 200 mg/m<sup>2</sup> by 24-hour continuous infusion days 1-5); course 3: aclarubicin 20 mg/m<sup>2</sup> days 1-5) and Ara-C 200 mg/m<sup>2</sup> by 24-hour continuous infusion days 1-5; course 4: Ara-C 200 mg/m<sup>2</sup> by 24-hour continuous infusion days 1-5, etoposide 100 mg/m<sup>2</sup> days 1-5, vincristine 0.8 mg/m<sup>2</sup> day 8, and vindesine 2 mg/m<sup>2</sup> day 10) or 3 courses of high-dose cytarabine (HiDAC) therapy (2 g/m<sup>2</sup> twice daily for 5 days). Allo-SCT was offered during the first CR to patients 50 years of age or younger and with a

histocompatible donor in the intermediate or adverse cytogenetic risk groups.