

1 **The origin and genetic variability of vegetatively propagated clones identified from**
2 **old planted trees and plantations of *Thujaopsis dolabrata* var. *hondae* in Ishikawa**
3 **Prefecture, Japan**

4 Torazo Ikeda^{a, b, c}, Kentaro Mishima^d, Katsuhiko Takata^e and Nobuhiro Tomaru^b

5 ^a *Department of Agriculture, Forestry and Fisheries, Ishikawa Prefectural Government,*
6 *Ishikawa, Japan*

7 ^b *Graduate School of Bioagricultural Sciences, Nagoya University, Nagoya, Japan*

8 ^c *Present address: Forestry Research Institute, Hokkaido Research Organization,*
9 *Hokkaido, Japan*

10 ^d *Forest Tree Breeding Center, Forestry and Forest Products Research Institute, Forest*
11 *Research and Management Organization, Ibaraki, Japan*

12 ^e *Institute of Wood Technology, Akita Prefectural University, Akita, Japan*

13

14 Corresponding author: Torazo Ikeda

15 Forestry Research Institute, Hokkaido Research Organization, Higashiyama, Koshunai-
16 cho, Bibai, Hokkaido 079-0198, Japan

17 Telephone: 0126-63-4164

18 Fax: 0126-63-4166

19 E-mail: ikeda-torazo@hro.or.jp

20

21 **Acknowledgments**

22 We thank Dr. Jiro Kodani, Ms. Tomomi Hashiba, Ms. Noriko Tokuda, Ms. Moeko
23 Kawasaki, Mr. Yo Sengi, and other members of Forestry Experiment Station, Ishikawa
24 Agriculture and Forest Research Center, for research support and helpful advice. We also
25 thank Associate Professor Tatsuro Hamada of Ishikawa Prefectural University and Dr.
26 Masako Komaki of Agricultural Experiment Station, Ishikawa Agriculture and Forest
27 Research Center, for technical support.

28 **Abstract**

29 Clonal plantations of *Thujopsis dolabrata* var. *hondae* have been established in Ishikawa
30 Prefecture, Japan, since at least the 1800s. Historical planting of the species has led to the
31 development of vegetatively propagated local cultivars, which originated from ‘donor’
32 trees that have often been conserved in sacred groves or avenues at shrines and temples.
33 These donor trees must have been selected from natural populations. In this study we
34 estimated the origin and genetic variability of clones identified among old planted trees
35 and clonal plantations of *T. dolabrata* var. *hondae*, using 19 microsatellite markers. We
36 discovered 12 clones among old planted trees, including five identical to members of a
37 set of 14 we previously identified in plantations (giving 21 clones in total). Based on
38 analyses combining assignment and exclusion tests, we inferred origins of eight of those
39 21 clones: six may have originated from a natural population distributed in Ishikawa, one
40 from Hokkaido & Aomori, and the other from Iwate & Yamagata, suggesting the clones
41 constituting cultivars have multiple origins. The clones identified in plantations have
42 significantly lower genetic variability, and higher relatedness, indicating that clones of
43 cultivars have a much narrower genetic base than those of natural populations. We
44 suggest new clones selected from natural populations elsewhere, as well as Ishikawa, are
45 needed for future breeding of *T. dolabrata* var. *hondae* to develop clonal forestry for this
46 species.

47 **Keywords:** assignment test; clonal forestry; genetic diversity; local cultivar;
48 microsatellite; natural population

49 **Introduction**

50 Clonal forestry refers to extensive deployment of relatively few clones with proven
51 phenotypic superiority (White et al. 2007). Clonal forestry plantations have been
52 established with various tree species, such as *Cryptomeria japonica* (Miyajima 1989;
53 Ohba 1993) and *Chamaecyparis obtusa* (Sato and Miyajima 1956; Miyajima 1989) in
54 Japan, *Picea abies* in Europe (Bentzer 1993), *Pinus radiata* in New Zealand (Arnold and
55 Gleed 1985) and species and interspecific hybrids of *Eucalyptus* (e.g., Van Wyk 1985),
56 *Populus* and *Salix* (Zsuffa et al. 1993) in many countries. In clonal forestry of *C. japonica*
57 in Japan, individuals used for plantations have been spontaneously or instinctively
58 selected during the domestication process over a long period. This has led to the
59 development of many local cutting cultivars, e.g., Measa, Hon-sugi, Aya-sugi and
60 Yabukuguri in the Kyushu region, Okinoyama-sugi in Tottori Prefecture, Boka-sugi in
61 Toyama Prefecture and Sanbu-sugi in Chiba Prefecture (Miyajima 1989; Ohba 1993).

62 *Thujopsis dolabrata* var. *hondae* is a coniferous species, distributed from southern
63 Hokkaido to northern Honshu in Japan (Hayashi 1960). Its wood has excellent strength,
64 durability and moisture resistance (Saito 1972), so it has become one of the most
65 important forestry species in Japan. It is mainly harvested in three prefectures: Aomori,
66 Niigata and Ishikawa. In Aomori and Niigata, its wood is mostly produced from natural
67 populations. However, in Ishikawa the wood comes from clonal plantations. These were
68 established in the past from saplings created by layering or unrooted cuttings planted
69 directly in the forests and, in recent years, from rooted cuttings or saplings created by air
70 layering. In Ishikawa Prefecture, the species is known as ‘ate’ and has been planted
71 (mainly on the Noto Peninsula) since at least the 1800s (Saito 1972). Such historical
72 planting has led to the development of more than 20 vegetatively propagated local
73 cultivars, most importantly Ma-ate, Kusa-ate, Eso-ate (Suzu-ate) and Kana-ate. Ma-ate

74 consists of multiple clones, but a single clone predominates in plantations of each of the
75 other three cultivars (Nakano 1990; Ikeda et al. 2018). A twisted trunk is one of several
76 important traits for identifying the Ma-ate cultivar, although Kana-ate trees may also have
77 a twisted trunk (Nakano 1990; Forest Experiment Station, Ishikawa Agriculture and
78 Forest Research Center 2016). Ikeda et al. (2018) identified clones in plantations using
79 microsatellite markers and found that present plantations had been established from at
80 least 14 clones. The original (donor) trees that gave rise to the clones have often been
81 conserved in sacred groves or avenues at shrines and temples (Saito 1972). People seem
82 to have taken cuttings from these holy trees and distributed them in their local areas.
83 These old planted trees must have been selected from natural populations. The clones
84 used for clonal forestry may have hypothetically originated from natural populations
85 distributed in Ishikawa. An alternative hypothesis, based on oral traditions, is that
86 planting material was brought from natural populations in the Tohoku region (which
87 encompasses Aomori, Iwate and Yamagata Prefectures) about 300, 400 or 800 years ago
88 (Agriculture, Forestry and Fisheries Department, Ishikawa Prefectural Government 1997;
89 Forest Experiment Station, Ishikawa Agriculture and Forest Research Center, 2016). It is
90 still not clear which hypothesis is correct.

91 Microsatellite marker-based genetic analyses of vegetatively propagated cultivars of
92 *C. japonica*, *C. obtusa* and *T. dolabrata* var. *hondae* used for clonal forestry in Japan
93 have provided valuable information (Matsui et al. 2013; Goto et al. 2008; Ikeda et al.
94 2018). High genetic variability has been found among 19 major cutting cultivars of *C.*
95 *japonica* in the Kyushu region, but generally close genetic relationships between clones
96 within the cultivars (Matsui et al. 2013). Clones of a local cutting cultivar of *C. obtusa*,
97 Nango-hi appear to be more closely related to each other, and have lower genetic
98 variability, than individuals in natural populations (Goto, et al. 2008). Clones of the Ma-

99 ate cultivar of *T. dolabrata* var. *hondae* also tend to have close relationships (Ikeda et al.
100 2018). All these studies indicate that clones within cultivars tend to be genetically closely
101 related. This may be problematic when crossing clones of conifer cultivars to create new
102 cultivars, because of risks of severe inbreeding depression (Williams and Savolainen
103 1996). Thus, there are clear needs to raise and maintain higher genetic variability of
104 breeding populations. The natural populations from which clones probably originated are
105 very important genetic resources, and recurrent selection from them would be an efficient
106 way of increasing breeding populations' genetic variability. Identifying such ancestral
107 populations is far from straightforward, because clones originated from various natural
108 populations throughout a long history of domestication (Testolin et al. 2000; Honjo et al.
109 2008). However, assignment tests based on microsatellite genotypes have proven utility
110 for inferring source populations of individuals of several species (Honjo et al. 2008; Goto
111 et al. 2008; Caldera et al. 2008).

112 Thus, in this study we used microsatellite markers to evaluate the genetic variability of
113 vegetatively propagated clones identified among old planted trees and plantations of *T.*
114 *dolabrata* var. *hondae* in Ishikawa Prefecture. We compared the variability of the clones
115 and natural populations of the species. We also attempted to identify the clones' origins,
116 in potential natural source populations, using assignment tests based on microsatellite
117 genotypes. Finally, we discuss implications of the findings for future breeding of *T.*
118 *dolabrata* var. *hondae*.

119

120 **Materials and methods**

121 **Plant materials**

122 We collected leaves from 24 old planted *T. dolabrata* var. *hondae* trees at 12 shrines, four
123 temples and one private garden (of the Izumi family) in Ishikawa Prefecture (Table 1, Fig.

124 S1). We also examined the trunk twist of each tree. The two trees in the private garden
125 (old tree nos. 16 and 17), which were believed to be the origin of ate, were the oldest
126 (more than 450 years old) of all the old planted trees. The two old planted trees at
127 Hinomiya Shrine (nos. 13 and 14) were reportedly origins of the Kusa-ate cultivar (Saito
128 1972). Ikeda et al. (2018) identified 14 clones (designated C-1, -6, -7 and -14, and C-44
129 to -53) in *T. dolabrata* var. *hondae* plantations in Ishikawa, based on 12 microsatellite
130 genotypes, which we used as genotype data in assignment tests for this study. As potential
131 source populations in assignment tests for clones among old planted trees and plantations,
132 we selected 17 natural populations from the entire distribution range of *T. dolabrata* var.
133 *hondae* and collected leaves from a total of 410 individuals belonging to the natural
134 populations (Fig. 1 and Table S1).

135

136 **DNA extraction and genotyping**

137 We extracted genomic DNA from the leaves collected from old planted trees and
138 individuals in natural populations and determined genotypes at 19 microsatellite loci
139 (Tdest 1, Tdest 3, Tdest 11, Tdest 14, Tdest 17, Tdest 21, Tdest 24, Tdest 29, Tdest 35,
140 Tdest 37, Tdest 38, Tdest 39, Tdest 42, Tdest 43, Tdest 45, Tdest 49, Tdest 53, Tdest 56
141 and Tdest 58) developed for *T. dolabrata* var. *hondae* (Sato et al. 2015). We also
142 determined genotypes of the 14 clones from plantations examined by Ikeda et al. (2018)
143 at seven additional microsatellite loci. The methods used for DNA extraction and
144 microsatellite genotyping were as in the previous study (Ikeda et al. 2018).

145

146 **Data analysis**

147 **Clone identification and analysis of clones' genetic relationships**

148 We assigned the sampled old planted trees to clones, based on the genotypes at 19
149 microsatellite loci with reference to those of the 14 clones from plantations (Ikeda et al.
150 2018). When old planted trees shared identical multilocus genotypes with known clones
151 from plantations, with no mismatched loci, the old planted trees were considered to be
152 known clones. If the multilocus genotypes of old planted trees did not match those of any
153 known clones from plantations, the old planted trees were considered to be new clones.

154 To estimate genetic relationships between clones, we calculated the coefficient of
155 relatedness (r , the fraction of alleles identical by descent shared by individuals; Blouin,
156 2003), using ML-Relate software (Kalinowski et al. 2006). For this, we applied allele
157 frequencies based on data for all sampled individuals in the 17 natural populations
158 included in the study (Fig. 1, Table S1). This coefficient is 1.000 for individuals of the
159 same clone, 0.5 for parents and offspring or full siblings, 0.250 for half siblings, and 0.125
160 for first cousins. We also calculated values of r among individuals in each natural
161 population to compare with those among clones in the old planted trees and plantations.

162

163 **Analysis of genetic diversity and structure in natural populations**

164 To estimate genetic diversity within each natural population, we calculated the number
165 of alleles per locus (A), allelic richness based on 12 diploid individuals (A_R), observed
166 heterozygosity (H_O), gene diversity (H_E) and inbreeding coefficient ($F_{IS} = 1 - H_O / H_E$)
167 over all loci in each population. The significance of departures from Hardy-Weinberg
168 equilibrium (HWE) over all loci in each population was evaluated by randomization tests.

169 To estimate genetic diversity across all natural populations, we calculated the total
170 number of alleles detected (TN), H_O , average gene diversity within populations and gene
171 diversity in the total population (H_S and H_T , respectively; Nei 1987), and inbreeding
172 coefficient and genetic differentiation measure (F_{IS} and F_{ST} , respectively; Weir and

173 Cockerham 1984) across all populations at each locus and over all loci. The standardized
174 genetic differentiation measure (G'_{ST} ; Hedrick 2005) was also manually calculated. The
175 significance of departures from HWE and genetic differentiation among populations at
176 each locus and over all loci was evaluated by randomization tests. The above calculations,
177 apart from G'_{ST} calculation, were performed using FSTAT version 2.9.4 (Goudet 1995).

178 To assess the individual-based genetic structure among natural populations of *T.*
179 *dolabrata* var. *hondae*, the Bayesian clustering method was applied, using STRUCTURE
180 version 2.3.4 software (Pritchard et al. 2000). Simulations were run 10 times for each
181 value of K (number of genetic clusters) from 1 to 10 for 10,000 iterations after a burn-in
182 period of 10,000 iterations with an admixture model, an allele frequency correlated model
183 (Falush et al. 2003) and a LOCPRIOR model (Hubisz et al. 2009). The optimal value of
184 K was determined based on the log probability of data and the ΔK for each K (Evanno et
185 al. 2005) using STRUCTURE HARVESTER software (Earl and VonHoldt, 2012). To
186 facilitate interpretation of the results from STRUCTURE analysis, we used CLUMPP
187 version 1.1.2 (Jakobsson and Rosenberg, 2007). We also used DISTRUCT version 1.1
188 (Rosenberg, 2004) to visualize the results. Analysis of molecular variance (AMOVA)
189 was performed with GenAlEx version 6.502 (Peakall and Smouse, 2012), based on the
190 genetic structure among natural populations detected by STRUCTURE, which separated
191 the 17 natural populations into four regional groups. These regions were: Hokkaido &
192 Aomori, Iwate & Yamagata, Niigata, and Ishikawa (for details, see ‘Genetic diversity and
193 structure in natural populations’ in the Results section). Genetic variation of the natural
194 populations was hierarchically divided into three layers (the regions inferred by
195 STRUCTURE analysis, populations and individuals) and variance components for each
196 layer and related Φ -statistics were calculated. The significance of each Φ -statistic was
197 evaluated by a permutation test implemented in GenAlEx.

198

199 **Comparison of genetic variability among clones and among individuals in natural**
200 **populations**

201 To estimate genetic variability among clones, we calculated A , A_R , H_O and H_E for clones
202 identified among old planted trees and in plantations, using FSTAT. Similarly, to estimate
203 genetic variability among individuals within natural populations in the regions inferred
204 by STRUCTURE analysis as well as that across all the populations, we averaged the
205 values of A_R and H_E over populations in each region and over all populations. The A_R and
206 H_E values for the clones detected among old planted trees and in plantation were
207 compared with the average values for all the natural populations, using a Wilcoxon's
208 signed-rank test.

209

210 **Inferring origins of clones**

211 To infer the origin of each clone, we applied a Bayesian-based assignment test (Rannala
212 and Mountain 1997) with the 'leave one out' procedure implemented in GeneClass 2
213 software (Piry et al. 2004), using microsatellite genotype data. This test estimates the
214 likelihood that an individual belongs to a population and assigns the population with the
215 highest likelihood as the origin of the individual. We set the four regions inferred by
216 STRUCTURE analysis as units of origins of clones. To confirm that there was sufficient
217 information for estimating the origin of each clone, we applied a self-assignment test for
218 individuals from natural populations in the four regions. Next we applied an assignment
219 test for each clone, regarding the natural populations in the four regions as potential
220 source populations. To confirm that individuals belong to potential source populations,
221 in addition to an assignment test an exclusion test can be applied (Cornuet et al. 1999).
222 Combinations of assignment and exclusion tests have been used effectively to determine

223 original populations of individuals in several species (Craig et al. 2000, Hare et al. 2006,
224 Frantz et al. 2006). Thus, we also applied an exclusion test using the Monte Carlo
225 resampling method (Paetkau et al. 2004) with 10,000 replicates implemented in
226 GeneClass2. When the probability that populations in a region were sources of a clone
227 was less than a given threshold ($\alpha = 0.01$) in the exclusion test, we concluded that
228 populations in that region were not sources for the tested clone.

229

230 **Results**

231 **Clone identification and genetic relationships between clones**

232 We identified 12 clones (C-1, -6, -7, -14, -46 and C-54 to -60; Table 1) among old planted
233 trees, five of which (C-1, -6, -7, -14 and -46) were the same as clones found in plantations
234 (Ikeda et al. 2018), while the other seven (C-54 to -60) were newly discovered. Four of
235 the first five clones (C-1, -6, -7 and -14) are commonly used and represent major clones
236 in plantations (Ikeda et al. 2018). Half of the 24 old planted trees were identical to one of
237 the five clones identified in plantations.

238 Values of the coefficient of relatedness (r) among the 21 clones detected in plantations
239 and old planted trees ranged from 0.000 to 0.799 (Table 2), and were particularly high
240 (>0.5) among C-44, C-53 and C-54, as well as between C-46 & C-49, C-46 & C-59, and
241 C-49 & C-50. Average values of r among clones in old planted trees and plantations were
242 0.102 and 0.183, respectively; higher than those among individuals within each natural
243 population (0.015 to 0.060; on average, 0.031; Table S1).

244

245 **Genetic diversity and structure in natural populations**

246 The allelic richness (A_R) and gene diversity (H_E) within each natural population ranged

247 from 5.03 to 6.55 and from 0.567 to 0.693, with averages of 5.88 and 0.652, respectively
248 (Table S1). Inbreeding coefficient (F_{IS}) ranged from -0.112 to 0.093. Two of the 17
249 populations showed significant deviation from Hardy-Weinberg equilibrium (HWE).
250 Over the 19 loci across the 17 natural populations, the average values of the total number
251 of alleles detected (TA), average gene diversity within populations (H_S) and gene
252 diversity in the total population (H_T) were 7.2, 0.652 and 0.677, respectively (Table S2).
253 Values for F_{IS} and two genetic differentiation measures (F_{ST} and G'_{ST}) over the 19 loci
254 were 0.018, 0.039 and 0.114, respectively. Significant deviation from HWE was detected
255 at two of the 19 loci and over all loci. Genetic differentiation among populations were
256 significant at 17 of the 19 loci and over all loci.

257 According to STRUCTURE analysis, log probabilities of the data increased with
258 increasing K up to a plateau at $K = 4$ (Fig. S2), while ΔK values were highest at $K = 2$
259 (Fig. S3). We therefore examined genetic structure among natural populations at $K = 2$,
260 3 and 4. At $K = 2$, the proportions of cluster 2-I in populations were highest in Hokkaido
261 & Aomori, lowest in Niigata & Ishikawa and intermediate in Iwate & Yamagata (Fig. 2).
262 At $K = 3$, clusters 3-I, 3-III and 3-II dominated in Hokkaido & Aomori, Iwate & Yamagata,
263 and Niigata & Ishikawa, respectively, although populations 10 in Aomori and 15 in
264 Niigata also had relatively high proportions of cluster 3-III. At $K = 4$, similar patterns
265 appeared as at $K = 3$: clusters 4-I, 4-III and 4-II dominated in Hokkaido & Aomori, Iwate
266 & Yamagata, and Niigata & Ishikawa, respectively, although most populations contained
267 mixtures of clusters. Based on the genetic structure among populations detected by
268 STRUCTURE analysis, we separated the 17 natural populations into four regional
269 groups: Hokkaido & Aomori, Iwate & Yamagata, Niigata, and Ishikawa. At $K = 4$, the
270 values of F_{ST} were about twice as high for clusters 4-II and 4-IV (0.0817 and 0.0675,
271 respectively) as for clusters 4-I and 4-III (0.0360 and 0.0412, respectively) (Fig. 2).

272 AMOVA results indicated that 8.7, 5.9 and 85.4% of detected genetic variation resided
273 among regions, among populations within regions, and among individuals, respectively
274 (Table S3). All Φ -statistics were highly significant ($P < 0.001$).

275

276 **Comparison of genetic variability among clones and among individuals in natural** 277 **populations**

278 Across natural populations in the regions inferred by STRUCTURE analysis, the average
279 values of A_R decreased from north to south, from 5.97 for populations in Hokkaido &
280 Aomori to 5.39 for populations in Ishikawa (Table 3). Average values of H_E decreased
281 from 0.668 to 0.586 in a similar fashion. Average values of A_R and H_E over all the natural
282 populations were 5.88 and 0.652, respectively. The A_R and H_E values for the 12 clones
283 detected among old planted trees were 5.58 and 0.647, respectively; not significantly
284 lower than the averages for all the natural populations. However, values for the 14 clones
285 detected in plantations were significantly lower: 4.90 and 0.616 ($P < 0.01$ and $P < 0.05$,
286 respectively; Wilcoxon's signed-ranks test).

287

288 **Inferring origins of clones**

289 To confirm that the dataset for reference populations included sufficient information to
290 allow estimation of the origins of clones, we applied a self-assignment test for individuals
291 from natural populations in four regions. This resulted in correct assignment of 362
292 individuals (88.3% of the total) to their respective sampling regions (Table S4). To infer
293 the clones' origins, we applied a combination of assignment and exclusion tests.
294 Probabilities obtained for the first, second and third most likely source regions for the 21
295 clones are shown in Table 4. Criteria for assignment to a single region were probability
296 higher than 0.01 for the first region and lower than 0.01 for the second and third regions.

297 Eight clones were assigned only to single regions (C-1, -6, -45, -48, -55 and -59 to
298 Ishikawa; C-7 to Hokkaido & Aomori; C-52 to Iwate & Yamagata), but four (C-14, -46,
299 -49 and -60) were not assigned to any of the four regions and the other 9 were assigned
300 to more than one region.

301

302 **Discussion**

303 **Original trees that gave rise to vegetatively propagated local cultivars**

304 Historical planting of *T. dolabrata* var. *hondae* has led to the development of more than
305 20 vegetatively propagated local cultivars. The four major cultivars are Ma-ate, Kusa-ate,
306 Eso-ate (Suzu-ate) and Kana-ate (Forest Experiment Station, Ishikawa Agriculture and
307 Forest Research Center, 2016). The original (donor) trees of cultivars have often been
308 conserved as sacred trees or in groves in the grounds of shrines and temples (Saito 1972).
309 Ikeda et al. (2018) identified clones of three cultivars (Ma-ate, Kusa-ate and Eso-ate)
310 using microsatellite markers and demonstrated that the Ma-ate cultivar consists of
311 multiple clones (including the three most frequently used in plantations: C-1, -6 and -7).
312 We found that the oldest planted trees (older than 450 years), which were believed to be
313 the origin of ate, were identical to clone C-7 of the Ma-ate cultivar. The other two clones,
314 C-1 and -6, also apparently originated from old planted trees. All the old planted trees of
315 clones C-1, -6 and -7 had a dextrally twisted trunk (Table 1) and bark resembling that of
316 *C. obtusa*, which are characteristics of the Ma-ate cultivar.

317 Ikeda et al. (2018) also demonstrated that plantations of the Kusa-ate cultivar are
318 dominated by a single clone (C-14), which we confirmed originated from old planted
319 trees previously regarded as the cultivar's origin. In addition, Ikeda et al. (2018) argued
320 that two clones detected in plantations (C-44 and -53) were probably trees of the Eso-ate
321 cultivar. However, genotypes of these two clones were not found among the old planted

322 trees and they are closely related to the Ma-ate clone (C-1), according to coefficients of
323 relatedness (r) estimated in this study. Clone C-54, identified among old planted trees,
324 had high values of r with C-44 and C-53 (0.799 and 0.569, respectively). These values
325 indicate that C-54 has a closer than parent-offspring or full sibling relationship with C-
326 44, and relationship as close as parent-offspring or full sibling with C-53. Clone C-54
327 may therefore be the original clone of the Eso-ate cultivar, and its derivative clones (C-
328 44 and -53) may have originated from seedlings of C-54 and been used for establishing
329 plantations as the Eso-ate cultivar. Accordingly, this study demonstrates that the original
330 trees of the Ma-ate, Kusa-ate and Eso-ate clones (C-1, -6, -7, -14 and -54) have been
331 conserved as old planted trees at shrines or temples. A similar domestication history has
332 been proposed for vegetatively propagated local cultivars of *C. japonica* in the Kyushu
333 region, Japan. For example, original trees of one of the cultivars, Measa, may have been
334 selected from natural populations, preserved as sacred trees at shrines, and spread to
335 plantations in the Kyushu region (Miyajima 1989; Ohba 1993).

336

337 **The origins of clones**

338 The STRUCTURE analysis provided indications of four regional groups (Hokkaido &
339 Aomori, Iwate & Yamagata, Niigata, or Ishikawa) of natural populations within the
340 species, *T. dolabrata* var. *hondae*. Therefore, we conducted assignment tests with the four
341 regions inferred by STRUCTURE analysis as units of origins of clones. In the self-
342 assignment test, 88.3% of individuals from natural populations were correctly assigned
343 to their respective sampling regions. According to AMOVA ($\Phi_{RT} = 0.065$), only 5.9% of
344 genetic variation resided among regions, indicating that genetic differentiation among
345 regions is low. Because of this low genetic differentiation among regions, the proportion
346 of individuals assigned to their regions of origin was not particularly high. Use of an

347 exclusion test in addition to an assignment tests increased the accuracy of assigned origins
348 for some of the clones. Nevertheless, origins of 14 of the 21 clones could not be inferred
349 from results of combined assignment and exclusion tests, either because of the low
350 genetic differentiation among regions or because source populations were not included in
351 the analysis. Goto et al. (2008) also found that origins of eight of 21 clones of a local
352 cutting cultivar of *C. obtusa*, Nango-hi, could not be inferred using a combination of
353 assignment and exclusion tests, because of low genetic differentiation among candidate
354 natural populations ($G_{ST} = 0.039$; Tsumura et al. 2007). Several hypervariable markers
355 would be needed to assign more clones of *T. dolabrata* var. *hondae* cultivars to their
356 source populations. However, we were able to infer origins of seven of the 21 clones: five
357 were from Ishikawa, one from Hokkaido & Aomori, and the other from Iwate &
358 Yamagata, suggesting that clones of *T. dolabrata* var. *hondae* cultivars are derived from
359 multiple origins. Thus, results of this study support both hypotheses about the origin of
360 clones of this species: some appear to have originated from within Ishikawa Prefecture
361 and others from the Tohoku region.

362 The three most frequently used clones of the Ma-ate cultivar (C-1, -6 and -7) were
363 inferred to have originated from either inside or outside Ishikawa Prefecture. C-7's
364 inferred origins were natural populations in the Hokkaido & Aomori region. This is
365 consistent with oral traditions that old planted trees were brought from the Tohoku region,
366 which includes Aomori, Iwate and Yamagata Prefectures, in the 12th or 16th century
367 (Nihei and Tsuzi 1917). Inferred origins of the other two Ma-ate clones, C-1 and -6, were
368 natural populations in Ishikawa Prefecture. This suggests that trees of the two clones may
369 be classified as Ma-ate because of phenotypic similarities (in trunk twisting and bark) to
370 the oldest planted C-7 trees, although they have different origins from C-7 and the Ma-
371 ate clone complex has multiple origins. Origins of the Kusa-ate clone (C-14) and Eso-ate

372 clones (C-44, -53 and -54) could not be inferred in this study. However, Nagahama et al.
373 (1996) suggested that the Kusa-ate cultivar originated from Aomori Prefecture, because
374 the composition of its leaf oil is the same as in individuals from Ohata in Aomori
375 Prefecture.

376

377 **Genetic variability and relationships among clones**

378 The clones identified in plantations and old trees tended to have lower genetic variability
379 than natural populations. There may be two reasons for this. The first is phenotypic
380 selection during local cultivars' establishment. As a result of selection, the clones of
381 cultivars may have tended to have similar desirable characteristics, such as height,
382 diameter, volume, stem form, crown shape and bark traits. Hence, the genetic variability
383 of the clones selected for cultivars may have declined. Accordingly, selected individuals
384 generally have lower genetic variability than those in natural populations, as reported in
385 other coniferous species, such as *C. obtusa* (Goto et al. 2008), *Pinus radiata* (Moran and
386 Bell 1987) and *Picea glauca* (Rajora 1999). Another reason is that most of the clones
387 may have originated from natural populations in Ishikawa Prefecture, which have lower
388 genetic variability than natural populations in other regions. Six of the eight clones (C-1,
389 -6, -45, -48, -55 and -59) that were only assigned to single regions were inferred to have
390 originated from Ishikawa. The current natural populations in Ishikawa are small, located
391 at the southern limit of *T. dolabrata* var. *hondae*'s range on the Japan Sea side, and
392 geographically isolated from other natural populations. Low population size may lead to
393 loss of genetic variability through genetic drift and inbreeding, while isolation from other
394 populations may limit gene flow and accompanying increases in genetic variability
395 (Frankham 2005). A high F_{ST} value was obtained for the dominant cluster in natural
396 populations in Ishikawa detected by STRUCTURE analysis, suggesting they experienced

397 considerable genetic drift after divergence from the ancestral population (Falush,
398 Stephens, and Pritchard 2003). Therefore, the natural populations in Ishikawa may have
399 experienced substantial genetic drift without effective gene flow, resulting in substantial
400 reductions in genetic diversity within the populations.

401 The clones detected in plantations and old planted trees had closer genetic relationships
402 than individuals in natural populations. Similar patterns have been observed in other
403 species. For example, Goto et al. (2008) found that clones of a local cutting cultivar of *C.*
404 *obtusata*, Nango-hi, had closer relationships than conspecific individuals in natural
405 populations. Genotypes of cultivars have remained constant for hundreds of years through
406 vegetative propagation, and new cultivars tend to be produced by crossings between
407 cultivars (Sefc et al. 2000; Goto et al. 2008). Some of the clones of *T. dolabrata* var.
408 *hondae* cultivars may therefore be families, each consisting of a parent and its offspring,
409 as discussed by Ikeda et al. (2018), or generated by crossing between clones either of the
410 same or different cultivars. Such production of clones in cultivars has putatively occurred
411 in the history of the *C. obtusata* cultivar Nango-hi (Goto et al. 2008).

412

413 **Conclusions**

414 Our results suggest that clones of the Ma-ate, Kusa-ate and Eso-ate cultivars used in *T.*
415 *dolabrata* var. *hondae* clonal forestry originated from old planted trees, many of which
416 have been conserved. It also suggests that clones of cultivars used for clonal forestry
417 originated from natural populations in Ishikawa Prefecture and others in the Tohoku
418 region. The clones identified in current plantations have lower genetic variation, and
419 closer genetic relationships, than the natural populations. Thus, the clones of cultivars
420 used in clonal forestry have a much narrower genetic base. The close genetic relationships
421 between clones may be problematic when crossing clones of cultivars to create new

422 cultivars because such crossings cause severe inbreeding depression (Williams and
423 Savolainen 1996). Recurrent selection from the natural populations from which these
424 clones originated would be an efficient strategy for maintaining and increasing the genetic
425 variability of breeding populations (Zobel and Talbert 1984). Therefore, new clones from
426 natural populations elsewhere, as well as Ishikawa Prefecture, should be included in
427 future *T. dolabrata* var. *hondae* breeding programs to enhance the sustainability of clonal
428 forestry of the species in the Prefecture. It is also essential to conserve natural populations
429 of *T. dolabrata* var. *hondae* as genetic resources for future breeding of the species.

430

431 **Data Archiving Statement**

432 The microsatellite genotype data used in this study have been deposited in the Dryad
433 Digital Repository: [http:](http://) (If the manuscript is accepted, we will submit them to the
434 repository).

435

436 **Disclosure statement**

437 The authors declare that they have no conflicts of interest.

438 **References**

- 439 Agriculture, Forestry and Fisheries Department, Ishikawa Prefectural Government
440 (1997) Ate (*Thujopsis dolabrata* var. *hondae*) in Noto. Ishikawa Prefectural
441 Government, Kanazawa (in Japanese)
- 442 Arnold R, Gleed JA (1985) Raising and managing radiata pine cuttings for production
443 forests. *Aust For* 48:199–206
- 444 Bentzer BG (1993) Strategies for clonal forestry with Norway spruce. In: Ahuja MR,
445 Libby WJ (eds) *Clonal forestry II, Conservation and application*. Springer-Verlag,
446 Berlin Heidelberg, pp 120–138
- 447 Blouin MS (2003) DNA-based methods for pedigree reconstruction and kinship
448 analysis in natural populations. *Trends Ecol Evol* 18:503–511
- 449 Caldera EJ, Ross KG, DeHeer CJ, Shoemaker DDW (2008) Putative native source of
450 the invasive fire ant *Solenopsis invicta* in the USA. *Biol Invasions* 10:1457–1479
- 451 Cornuet JM, Piry S, Luikart G, Estoup A, Solignac M (1999) New methods employing
452 multilocus genotypes to select or exclude population as origins of individuals.
453 *Genetics* 153:1989–2000
- 454 Craig R. P, Koskinen MT, Piironen J (2000) The one that did not get away: individual
455 assignment using microsatellite data detects a case of fishing competition fraud.
456 *Proc R Soc B Biol Sci* 267:1699–1704
- 457 Earl DA, VonHoldt BM (2012) STRUCTURE HARVESTER: a website and program
458 for visualizing STRUCTURE output and implementing the Evanno method.
459 *Conserv Genet Resour* 4:359–361

460 Evanno G, Regnaut S, Goudet J (2005) Detecting the number of clusters of individuals
461 using the software STRUCTURE: a simulation study. *Mol Ecol* 14:2611–2620

462 Falush D, Stephens M, Pritchard JK (2003) Inference of population structure using
463 multilocus genotype data: linked loci and correlated allele frequencies. *Genetics*
464 164:1567–1587

465 Forest Experiment Station, Ishikawa Agriculture and Forest Research Center (2016) *Ate*
466 (*Thujopsis dolabrata* var. *hondae*) in Noto. Ishikawa Prefectural Government,
467 Kanazawa (in Japanese)

468 Frankham R (2005) Genetics and extinction. *Biol Conserv* 126:131–140

469 Frantz AC, Pourtois JT, Heuertz M, Schley L, Flamand MC, Krier A, Bertouille S,
470 Chaumont F, Burke T (2006) Genetic structure and assignment tests demonstrate
471 illegal translocation of red deer (*Cervus elaphus*) into a continuous population.
472 *Mol Ecol* 15:3191–3203

473 Goto S, Takahashi M, Matsumoto A, Ieiri R, Tsumura Y (2008) Genetic relationships
474 and origin of commercial clones of Nangouhi, a vegetatively propagated cultivar of
475 hinoki cypress (*Chamaecyparis obtusa*). *Breed Sci* 58:411–418

476 Goudet J (1995) FSTAT (version 1.2): a computer program to calculate *F*-statistics. *J*
477 *Hered* 86: 485–86

478 Hare MP, Allen SK, Bloomer P, Camara MD, Carnegie RB, Murfree J, Luckenbach M,
479 Meritt D, Morrison C, Paynter K, Reece KS, Rose CG (2006) A genetic test for
480 recruitment enhancement in Chesapeake Bay oysters, *Crassostrea virginica*, after
481 population supplementation with a disease tolerant strain. *Conserv Genet* 7:717–
482 734

- 483 Hayashi Y (1960) Taxonomical and phytogeographical study of Japanese conifers.
484 Norin-Shuppan, Tokyo (in Japanese)
- 485 Hedrick PW (2005) A standardized genetic differentiation measure. *Evolution* 59:
486 1633–1638.
- 487 Honjo M, Ueno S, Tsumura Y, Handa T, Washitani I, Ohsawa R (2008) Tracing the
488 origins of stocks of the endangered species *Primula sieboldii* using nuclear
489 microsatellites and chloroplast DNA. *Conserv Genet* 9:1139–1147
- 490 Hubisz MJ, Falush D, Stephens M, Pritchard JK (2009) Inferring weak population
491 structure with the assistance of sample group information. *Mol Ecol Resour*
492 9:1322–1332
- 493 Ikeda T, Tokuda N, Tomaru N (2018) Identification of *Thujopsis dolabrata* var. *hondae*
494 clones and their distribution across plantations in Ishikawa prefecture. *J For Res*
495 23:105–111
- 496 Jakobsson M, Rosenberg NA (2007) CLUMPP: a cluster matching and permutation
497 program for dealing with label switching and multimodality in analysis of
498 population structure. *Bioinformatics* 23:1801–1806
- 499 Kalinowski ST, Wagner AP, Taper ML (2006) ML-RELATE: a computer program for
500 maximum likelihood estimation of relatedness and relationship. *Mol Ecol Notes*
501 6:576–579
- 502 Matsui Y, Ieiri R, Moriguchi Y, Matsumoto A, Takahashi M, Tsumura Y (2013) Clone
503 identification of major cutting cultivars in *Cryptomeria japonica* D. Don and
504 evaluation of their genetic relationship in Kyushu. *J Jpn Soc* 95:220-226 (In
505 Japanese with English summary)

- 506 Miyajima H (1989) Sugi (*Cryptomeria japonica*) and hinoki (*Chamaecyparis obtusa*) in
507 Kyushu. Kyushu University Press, Fukuoka (in Japanese)
- 508 Moran GF, Bell JC (1987) The origin and genetic diversity of *Pinus radiata* in
509 Australia. *Theor Appl Genet* 73:616–622
- 510 Nagahama S, Tajima M, Ishimura K (1996) Chemotaxonomy of ate (*Thujopsis*
511 *dolabrata*) of Noto, Ishikawa Prefecture. *Mokuzai Gakkaishi* 42:698-702 (In
512 Japanese with English summary)
- 513 Nakano T (1990) External characteristic and zymograms of *Thujopsis dolabrata* var.
514 *hondae* cultivars. *Res Rep Ishikawa For Exp Stn* 21:14-21 (in Japanese)
- 515 Nei M (1987) *Molecular evolutionary genetics*. Columbia University Press, New York
- 516 Nihei H, Tsuzi K (1917) *Ate: rakanhaku*. Ishikawa Prefecture Forest Society, Kanazawa
517 (in Japanese)
- 518 Ohba K (1993) Clonal forestry with sugi (*Cryptomeria japonica*). In: Ahuja MR, Libby
519 WJ (eds) *Clonal forestry II, conservation and application*. Springer-Verlag, Berlin
520 Heidelberg, pp 66–90
- 521 Paetkau D, Robert S, Michael B, Arnaud E (2004) Genetic assignment methods for the
522 direct, real-time estimation of migration rate: a simulation-based exploration of
523 accuracy and power. *Mol Ecol* 13:55–65
- 524 Peakall R, Smouse PE (2012) GenALEX 6.5: genetic analysis in Excel. Population
525 genetic software for teaching and research-an update. *Bioinformatics* 28:2537–
526 2539

- 527 Piry S, Alapetite A, Cornuet JM, Paetkau D, Baudouin L, Estoup A (2004)
528 GENECLASS2: a software for genetic assignment and first-generation migrant
529 detection. *J Hered* 95:536–539
- 530 Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using
531 multilocus genotype data. *Genetics* 155:945–959
- 532 Rajora OP (1999) Genetic biodiversity impacts of silvicultural practices and phenotypic
533 selection in white spruce. *Theor Appl Genet* 99:954–961
- 534 Rannala B, Mountain JL (1997) Detecting immigration by using multilocus genotypes.
535 *Proc Natl Acad Sci* 94:9197–9201
- 536 Rosenberg NA (2004) DISTRUCT: a program for the graphical display of population
537 structure. *Mol Ecol Notes* 4:137–138
- 538 Saito K (1972) Silvicultural history of ate (*Thujopsis dolabrata* var. *hondae*). Ishikawa
539 Forestry Experiment Station, Hakusan (in Japanese)
- 540 Sato K, Miyajima H (1956) On a cultivar, Nango-hi, of *Chamaecyparis obtusa*. *Warm*
541 *Temp* 9:4-8 (in Japanese)
- 542 Sato M, Hasegawa Y, Mishima K, Takata K (2015) Isolation and characterization of 22
543 EST-SSR markers for the genus *Thujopsis* (Cupressaceae). *Appl Plant Sci*
544 3:11400101
- 545 Sefc KM, Lopes MS, Lefort F, Botta R, Roubelakis-Angelakis KA, Ibáñez J, Pejić I,
546 Wagner HW, Glössl J, Steinkellner H (2000) Microsatellite variability in
547 grapevine cultivars from different European regions and evaluation of assignment
548 testing to assess the geographic origin of cultivars. *Theor Appl Genet* 100:498–505

549 Testolin R, Marrazzo T, Cipriani G, Quarta R, Verde I, Dettori MT, Panclidi M,
550 Sansavini S (2000) Microsatellite DNA in peach (*Prunus persica* L. Batsch) and its
551 use in fingerprinting and testing the genetic origin of cultivars. *Genome* 43:512–
552 520

553 Tsumura Y, Matsumoto A, Tani N, Ujino-Ihara T, Kado T, Iwata H, Uchida K (2007).
554 Genetic diversity and the genetic structure of natural populations of
555 *Chamaecyparis obtusa*: implications for management and conservation. *Heredity*
556 99: 161–172

557 Van Wyk G (1985) Tree breeding in support of vegetative propagation of *Eucalyptus*
558 *grandis* (Hill) Maiden. *South African For J* 135:33–39

559 Weir BS, Cockerham CC (1984) Estimating *F*-statistics for the analysis of population
560 structure. *38*: 1358–1370

561 White TL, Adams WT, Neale DB (2007) *Forest genetics*. CABI International,
562 Wallingford

563 Williams CG, Savolainen O (1996) Inbreeding depression in conifers: implications for
564 breeding strategy. *For Sci* 42:102–117

565 Zobel BJ, Talbert JT (1984) *Applied forest tree improvement*. John Wiley & Sons, Inc,
566 New York

567 Zsuffa L, Sennerby-Forsse L, Weisgerber H, Hall R (1993) Strategies for clonal forestry
568 with poplars, aspens, and willows. In: Ahuja MR, Libby WJ (eds) *Clonal forestry*
569 II, Conservation and application. Springer-Verlag, Berlin Heidelberg, pp 91–119
570

571 **Titles and legends to figures**

572

573 **Fig. 1** Locations of 17 natural *Thujopsis dolabrata* var. *hondae* populations used in an
574 assignment test as potential source populations for the clones identified from old planted
575 trees and plantations. Numbers on the map correspond to the population numbers in Table
576 S1

577

578 **Fig. 2** Distribution of genetic clusters in each individual, at numbers of clusters (K) = 2,
579 3 and 4, for the 17 natural populations of *Thujopsis dolabrata* var. *hondae*, obtained by
580 STRUCTURE analysis. F_{ST} values for the clusters indicate their genetic divergence
581 from the ancestral population. Numbers below the chart correspond to the population
582 numbers in Table S1.

583 **Table 1** Designations, locations, diameter at breast height (DBH) and trunk twist
584 characteristics of 24 old planted *Thujaopsis dolabrata* var. *hondae* trees sampled from 12
585 shrines, four temples and one private garden in Ishikawa Prefecture, Japan

Old tree no.	Place planted	DBH	Trunk twist	Clone name
1	Yahata Shrine, Sasanami, Suzu	76	No	C-54
2		85	No	C-54
3	Komashihiko Shrine, Kyounen, Wakayama, Suzu	93	No	C-54
4		118	Sinistral	C-55
5	Ohhirume Shrine, Mawatari, Horyu, Suzu	102	No	C-56
6	Kumano Shrine, Omou, Noto, Hosu	83	No	C-54
7		88	No	C-54
8	Hakusan Shrine, Yanagida, Noto, Hosu	84	No	C-57
9	Koumyou Temple, Kitakawachi, Noto, Hosu	98	Dextral	C-7 ^a
10		98	No	C-57
11	Kasuga Shrine, Soyama, Anamizu, Hosu	92	Dextral	C-7 ^a
12		95	Dextral	C-7 ^a
13	Hinomiya Shrine, Hajikashi, Anamizu, Hosu	45	No	C-14 ^a
14		55	No	C-14 ^a
15	Sugawara Shrine, Tenjindani, Anamazu, Hosu	102	No	C-58
16	Private garden of Izumi family, Urakami, Monzen, Wajima	>150	Dextral	C-7 ^a
17		>150	Dextral	C-7 ^a
18	Isurugi Shrine, Hirose, Monzen, Wajima	120	Dextral	C-1 ^a
19	Ryugo Temple, Sakami, Shika, Hakui	58	Dextral	C-6 ^a
20	Jonen Temple, Notobe, Nakanoto, Kashima	56	Dextral	C-59
21	Tenpyou Temple, Mt. Sekido, Nakanoto, Kashima	73	Dextral	C-6 ^a
22	Tehayahime Shrine, Azuma, Hodatsushimizu, Hakui	94	Dextral	C-46 ^a
23	Hakusan Shrine, Kamiohta, Tsubata, Kahoku	82	Dextral	C-6 ^a
24	Ohkuninushi Shrine, Torigoe, Tsubata, Kahoku	102	No	C-60

586

587 ^aThese clones are the same as clones previously detected in plantations (Ikeda, Tokuda,
588 and Tomaru 2018)

589 **Table 2** Values of the coefficient of relatedness (r) among 21 *Thujaopsis dolabrata* var. *hondae* clones identified in old planted trees and plantations

	C-1	C-6	C-7	C-14	C-44	C-45	C-46	C-47	C-48	C-49	C-50	C-51	C-52	C-53	C-54	C-55	C-56	C-57	C-58	C-59	C-60
C-1																					
C-6	0.269																				
C-7	0.089	0.163																			
C-14	0.176	0.084	0.044																		
C-44	0.265	0.172	0.051	0.077																	
C-45	0.206	0.226	0.054	0.055	0.390																
C-46	0.248	0.191	0.035	0.167	0.116	0.071															
C-47	0.083	0.088	0.108	0.055	0.306	0.177	0.054														
C-48	0.328	0.312	0.256	0.137	0.309	0.453	0.134	0.115													
C-49	0.235	0.451	0.042	0.124	0.077	0.091	0.543	0.146	0.200												
C-50	0.105	0.371	0.111	0.183	0.275	0.072	0.209	0.310	0.101	0.531											
C-51	0.230	0.199	0.046	0.061	0.268	0.110	0.071	0.208	0.212	0.148	0.298										
C-52	0.307	0.231	0.180	0.340	0.061		0.112		0.240	0.153	0.149	0.109									
C-53	0.360	0.181		0.089	0.629	0.275	0.203	0.162	0.250	0.109	0.101	0.185	0.062								
C-54	0.161	0.116	0.095		0.799	0.262		0.178	0.260		0.193	0.206	0.076	0.569							
C-55	0.184	0.239	0.091		0.294	0.019	0.048		0.181	0.051			0.201	0.222	0.317						
C-56	0.050	0.162	0.072	0.038	0.034	0.195		0.167	0.066				0.058		0.016	0.053					
C-57	0.019	0.059	0.044	0.139	0.044	0.061	0.049	0.026	0.043	0.045	0.163		0.068		0.140	0.053	0.099				
C-58	0.081		0.068	0.121	0.157	0.060		0.115	0.077		0.091	0.008	0.035		0.136	0.071	0.001	0.049			
C-59	0.203	0.349	0.039		0.089		0.515		0.080	0.351	0.115		0.116	0.194	0.087	0.118		0.059			
C-60	0.053	0.084	0.121	0.082		0.003	0.138	0.059	0.135	0.138			0.122	0.044	0.033	0.155		0.214	0.069	0.057	

590

591 Blanks indicate that the values are 0.000. Values in bold are $r \geq 0.50$

592 **Table 3** Comparison of genetic variability among clones identified in plantations and old
 593 planted trees and among individuals in natural populations of *Thujopsis dolabrata* var.
 594 *hondae*

	Region	Population ^a	<i>N</i>	<i>A_R</i>	<i>H_E</i>
Old planted trees			12	5.58	0.647
Plantations			14	4.90	0.616
Natural populations	Hokkaido & Aomori ^b	1–10	23.5	5.97	0.668
	Iwate & Yamagata ^b	11–14	24.0	5.94	0.649
	Niigata	15	31	5.67	0.633
	Ishikawa ^b	16, 17	24.0	5.39	0.586
	All ^b	1–17	24.1	5.88	0.652

595

596 *N*, number of samples; *A_R*, allelic richness based on 12 diploid individuals; *H_E*, gene
 597 diversity

598 ^a Numbers correspond to the population numbers in Table S1

599 ^b Average values over populations are shown

600 **Table 4** Probabilities of the first, second and third most likely regions from which *Thujopsis dolabrata* var. *hondae* clones originated, based on a
601 combination of assignment and exclusion tests

Clone	First		Second		Third	
	Population	Probability	Population	Probability	Population	Probability
C-1	Ishikawa	0.013	Iwate and Yamagata	0.000 *	Hokkaido and Aomori	0.000 *
C-6	Ishikawa	0.060	Niigata	0.005 *	Iwate and Yamagata	0.000 *
C-7	Hokkaido and Aomori	0.026	Ishikawa	0.005 *	Iwate and Yamagata	0.004 *
C-14	Iwate and Yamagata	0.001 *	Hokkaido and Aomori	0.001 *	Niigata	0.001 *
C-44	Ishikawa	0.111	Iwate and Yamagata	0.034	Hokkaido and Aomori	0.017
C-45	Ishikawa	0.164	Hokkaido and Aomori	0.008 *	Iwate and Yamagata	0.004 *
C-46	Ishikawa	0.002 *	Hokkaido and Aomori	0.000 *	Iwate and Yamagata	0.000 *
C-47	Hokkaido and Aomori	0.027	Ishikawa	0.012	Iwate and Yamagata	0.008 *
C-48	Ishikawa	0.078	Iwate and Yamagata	0.002 *	Hokkaido and Aomori	0.002 *
C-49	Ishikawa	0.008 *	Iwate and Yamagata	0.001 *	Niigata	0.000 *
C-50	Niigata	0.034	Ishikawa	0.015	Iwate and Yamagata	0.012
C-51	Ishikawa	0.177	Niigata	0.133	Iwate and Yamagata	0.130
C-52	Iwate and Yamagata	0.011	Ishikawa	0.004 *	Hokkaido and Aomori	0.003 *
C-53	Iwate and Yamagata	0.038	Ishikawa	0.010	Hokkaido and Aomori	0.009 *
C-54	Ishikawa	0.522	Iwate and Yamagata	0.478	Hokkaido and Aomori	0.411
C-55	Ishikawa	0.019	Hokkaido and Aomori	0.003 *	Iwate and Yamagata	0.001 *
C-56	Ishikawa	0.682	Niigata	0.300	Iwate and Yamagata	0.244
C-57	Ishikawa	0.215	Niigata	0.046	Hokkaido and Aomori	0.009 *
C-58	Niigata	0.134	Ishikawa	0.090	Iwate and Yamagata	0.068
C-59	Ishikawa	0.077	Niigata	0.002 *	Hokkaido and Aomori	0.000 *
C-60	Ishikawa	0.000 *	Hokkaido and Aomori	0.000 *	Iwate and Yamagata	0.000 *

602 Clones in boldface indicate those assigned to a single region, with criteria of probabilities greater than 0.01 for the first region and lower than 0.01
603 for the second and third regions
604 *, probability < 0.01

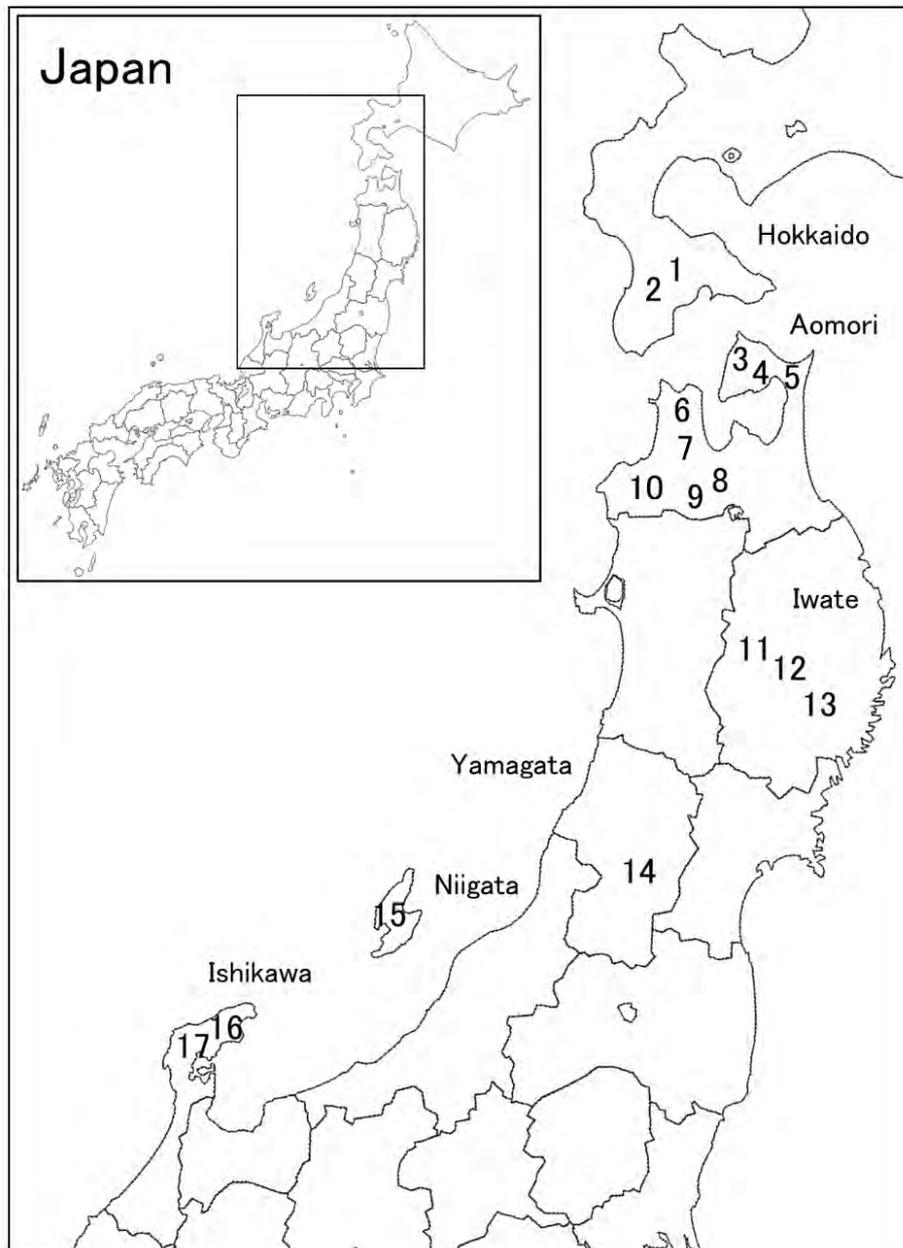


Fig. 1 Locations of 17 natural *Thujopsis dolabrata* var. *hondae* populations used in an assignment test as potential source populations for the clones identified from old planted trees and plantations. Numbers on the map correspond to the population numbers in Table S1

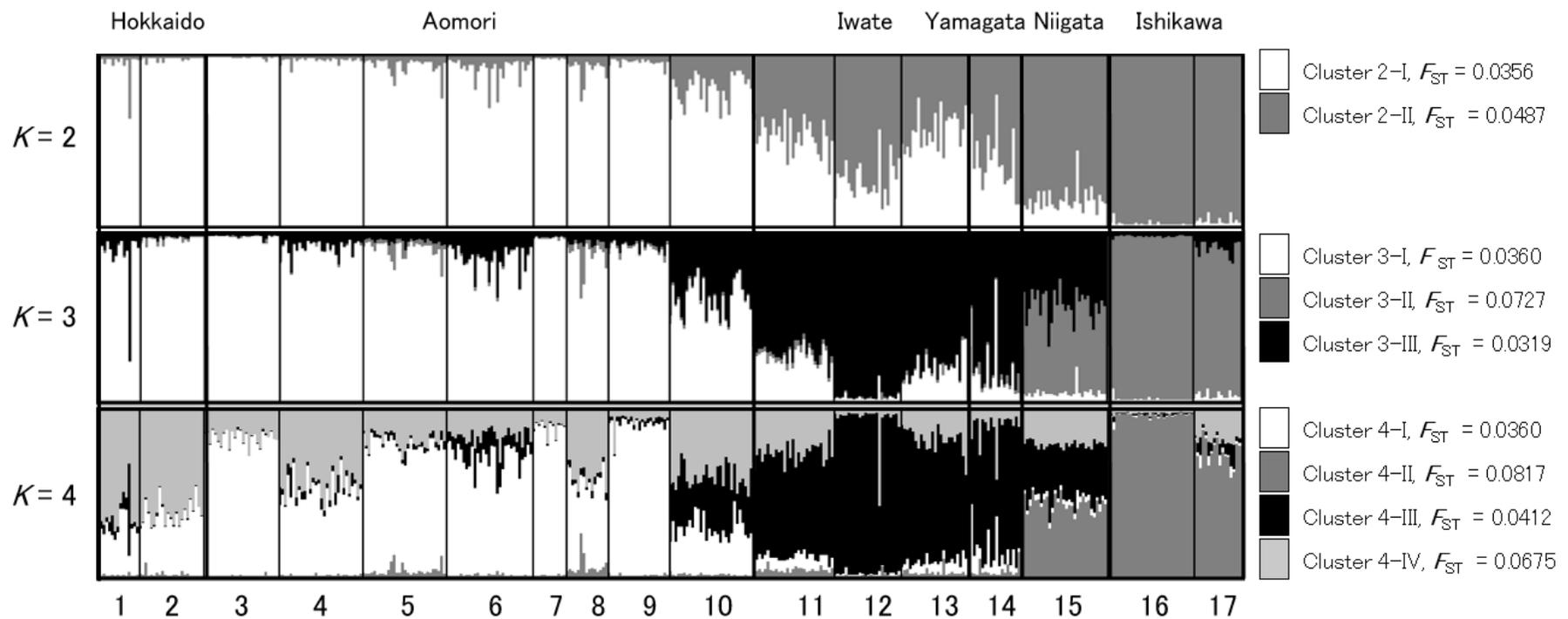


Fig. 2 Distribution of genetic clusters in each individual, at numbers of clusters (K) = 2, 3 and 4, for the 17 natural populations of *Thujopsis dolabrata* var. *hondae*, obtained by STRUCTURE analysis. F_{ST} values for the clusters indicate their genetic divergence from the ancestral population. Numbers below the chart correspond to the population numbers in Table S1.

Table S1 Locality, number of samples and level of genetic variability in old planted trees, plantations and natural populations of *Thujopsis dolabrata* var. *hondae*

Population no.	Locality	N	A	A_R	H_O	H_E	F_{IS}^a	r
Old planted trees	Ishikawa	12	5.58	5.58	0.588	0.647		0.102
Plantations	Ishikawa	14	5.11	4.90	0.568	0.616		0.183
Natural populations								
1	Minamidate, Assabu, Hokkaido	15	6.32	5.86	0.646	0.662	0.025	0.015
2	Todogawa, Esashi, Hokkaido	23	6.68	5.54	0.593	0.638	0.071	0.034
3	Okoppe, Aomori	27	7.53	5.97	0.673	0.674	0.002	0.028
4	Ohata, Aomori	30	8.00	6.02	0.628	0.676	0.071 *	0.026
5	Higashidori, Aomori	30	7.84	5.93	0.646	0.652	0.010	0.029
6	Nakazato, Nakadomari, Aomori	31	8.68	6.55	0.689	0.693	0.005	0.027
7	Ajigasawa, Aomori	12	5.84	5.84	0.746	0.671	-0.112	0.049
8	Mt. Cyobosan, Aomori	15	6.53	6.12	0.662	0.683	0.032	0.016
9	Owani, Aomori	22	6.84	5.76	0.624	0.652	0.042	0.029
10	Fukaura, Aomori	30	7.79	6.08	0.681	0.675	-0.009	0.028
11	Shizukuishi, Iwate	29	7.89	6.13	0.679	0.675	-0.006	0.028
12	Mt. Hayachine, Iwate	24	6.79	5.56	0.618	0.616	-0.004	0.027
13	Mt. Goyosan, Iwate	24	7.63	6.19	0.672	0.664	-0.012	0.025
14	Kaminoyama, Yamagata	19	6.68	5.87	0.637	0.641	0.006	0.033
15	Sado, Niigata	31	7.26	5.67	0.637	0.633	-0.005	0.033
16	Mt. Horyu, Ishikawa	31	6.42	5.03	0.514	0.567	0.093 *	0.050
17	Suzu, Ishikawa	17	6.47	5.74	0.573	0.605	0.054	0.060
Average		24.1	7.13	5.88	0.642	0.652		0.031

N , number of samples; A , number of alleles per locus; A_R , allelic richness based on 12 diploid individuals; H_O , observed heterozygosity; H_E , gene diversity; F_{IS} , inbreeding coefficient; r , coefficient of relatedness.

^aThe significance of departures from Hardy-Weinberg equilibrium was tested by randomization tests. Bonferroni correction was used to adjust P -values in multiple tests.

* $P < 0.05$.

Table S2 Genetic diversity at 19 microsatellite makers across 17 natural *Thujopsis dolabrata* var. *hondae* populations

Locus	TA	H_O	H_S	H_T	F_{IS}^a	F_{ST}^b	G'_{ST}
Tdest1	18	0.920	0.881	0.903	-0.040	0.025 **	0.213
Tdest3	10	0.428	0.452	0.456	0.070	0.011	0.013
Tdest11	14	0.809	0.831	0.849	0.029	0.016 **	0.131
Tdest14	18	0.864	0.884	0.896	0.006	0.013 **	0.118
Tdest17	11	0.654	0.678	0.780	0.040	0.136 **	0.424
Tdest21	28	0.890	0.903	0.926	0.020	0.025 **	0.272
Tdest24	16	0.695	0.725	0.754	0.037	0.037 **	0.144
Tdest29	9	0.461	0.473	0.498	0.043	0.048 **	0.096
Tdest35	33	0.863	0.910	0.939	0.048 *	0.031 **	0.364
Tdest37	4	0.417	0.419	0.418	-0.009	-0.002	-0.004
Tdest38	9	0.666	0.661	0.699	0.006	0.059 **	0.169
Tdest39	7	0.588	0.555	0.576	-0.045	0.033 **	0.086
Tdest42	11	0.557	0.563	0.585	0.004	0.036 **	0.090
Tdest43	14	0.765	0.752	0.783	-0.012	0.045 **	0.165
Tdest45	8	0.520	0.601	0.635	0.143 **	0.061 **	0.140
Tdest49	5	0.150	0.158	0.169	0.031	0.058 **	0.073
Tdest53	20	0.897	0.867	0.880	-0.031	0.017 **	0.119
Tdest56	6	0.661	0.650	0.686	-0.010	0.057 **	0.155
Tdest58	12	0.396	0.418	0.436	0.081	0.040 **	0.072
Average / overall	7.2	0.642	0.652	0.677	0.018 **	0.039 ***	0.114

TA, total number of alleles detected; H_O , observed heterozygosity; H_S , average gene diversity within populations; H_T , gene diversity in the total population; F_{IS} , inbreeding coefficient; F_{ST} , genetic differentiation measure; G'_{ST} , standardized genetic differentiation measure (Hedrick 2005).

^aThe significance of departures from Hardy-Weinberg equilibrium was tested by randomization tests.

^bThe significance of genetic differentiation among populations was tested by randomization tests.

Bonferroni correction was used to adjust P -values in multiple tests. * $P < 0.05$, ** $P < 0.01$,

*** $P < 0.001$.

Table S3 Results of analysis of molecular variance (AMOVA) of 17 natural populations of *Thujopsis dolabrata* var. *hondae* in four regions

Source of variation	d.f.	Sum of squares	Variance component	Percentage of variation (%)	Φ -Statistics	Probability
Among regions	3	456.4	1.38	8.7	$\Phi_{RT} = 0.087$	< 0.001
Among populations within regions	13	464.2	0.95	5.9	$\Phi_{PR} = 0.065$	< 0.001
Within populations	393	5356.5	13.63	85.4	$\Phi_{PT} = 0.146$	< 0.001

d.f., degree of freedom

Probabilities were based on permutation tests

Table S4 Results from a self-assignment test for 410 individuals from 17 natural populations of *Thujopsis dolabrata* var. *hondae* in four regions

Source region	<i>N</i>	%	Assigned region			
			Hokkaido and Aomori	Iwate and Yamagata	Niigata	Ishikawa
Hokkaido and Aomori	235	92.3	216	16	2	0
Iwate and Yamagata	96	85.3	9	81	4	1
Niigata	31	80.6	1	4	25	1
Ishikawa	50	80.0	0	1	9	40
Total	410	88.3	226	102	40	42

N, total number of samples in each region; %, percentage of samples that were assigned to the regions from which they were sampled

The numbers in boldface indicate the numbers of samples assigned to the originating regions

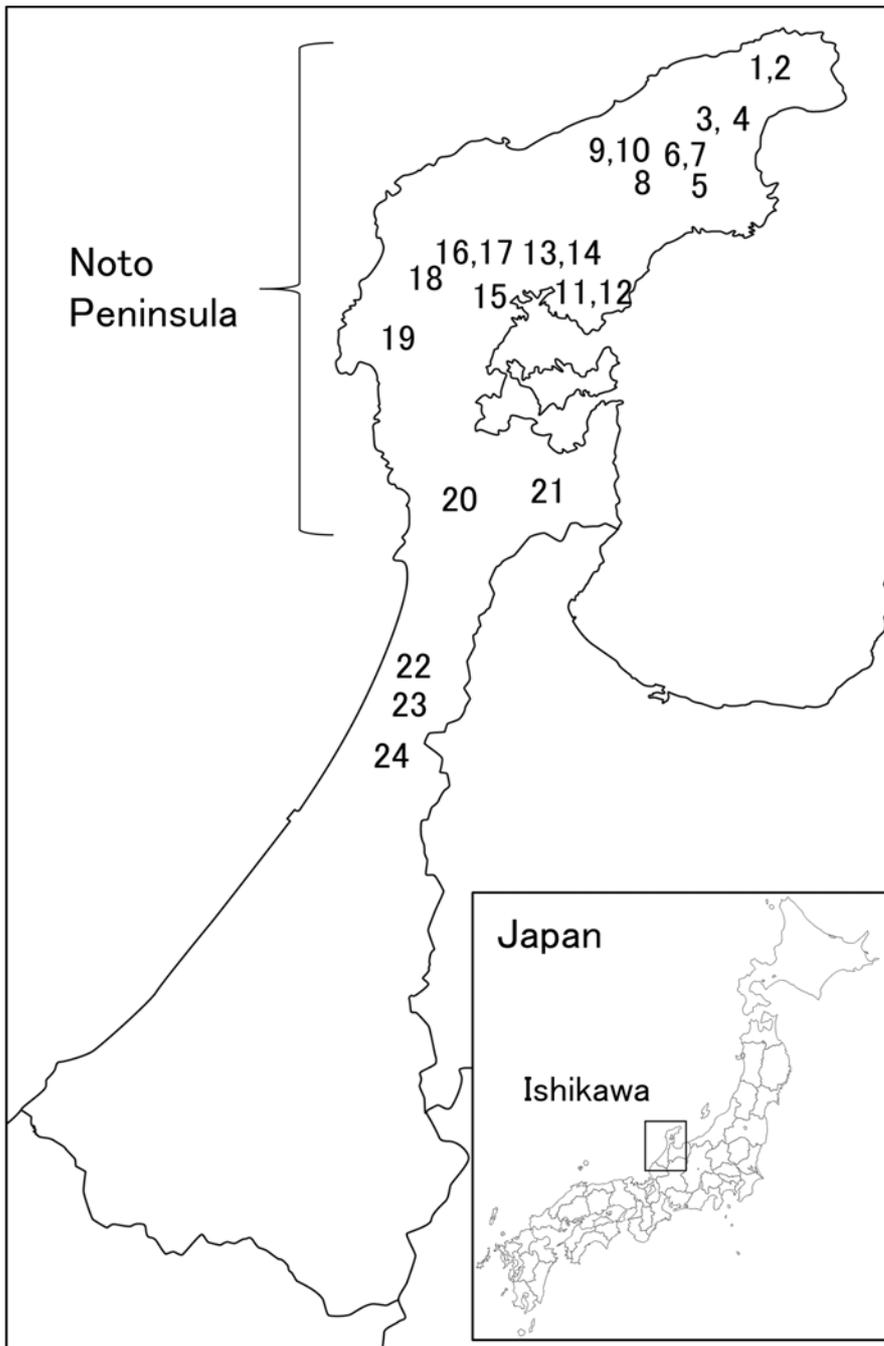


Fig. S1 Locations of the 24 old planted *Thujopsis dolabrata* var. *hondae* trees sampled at 12 shrines, four temples and one private garden in Ishikawa Prefecture, Japan. Numbers on the map correspond to the old tree numbers in Table 1

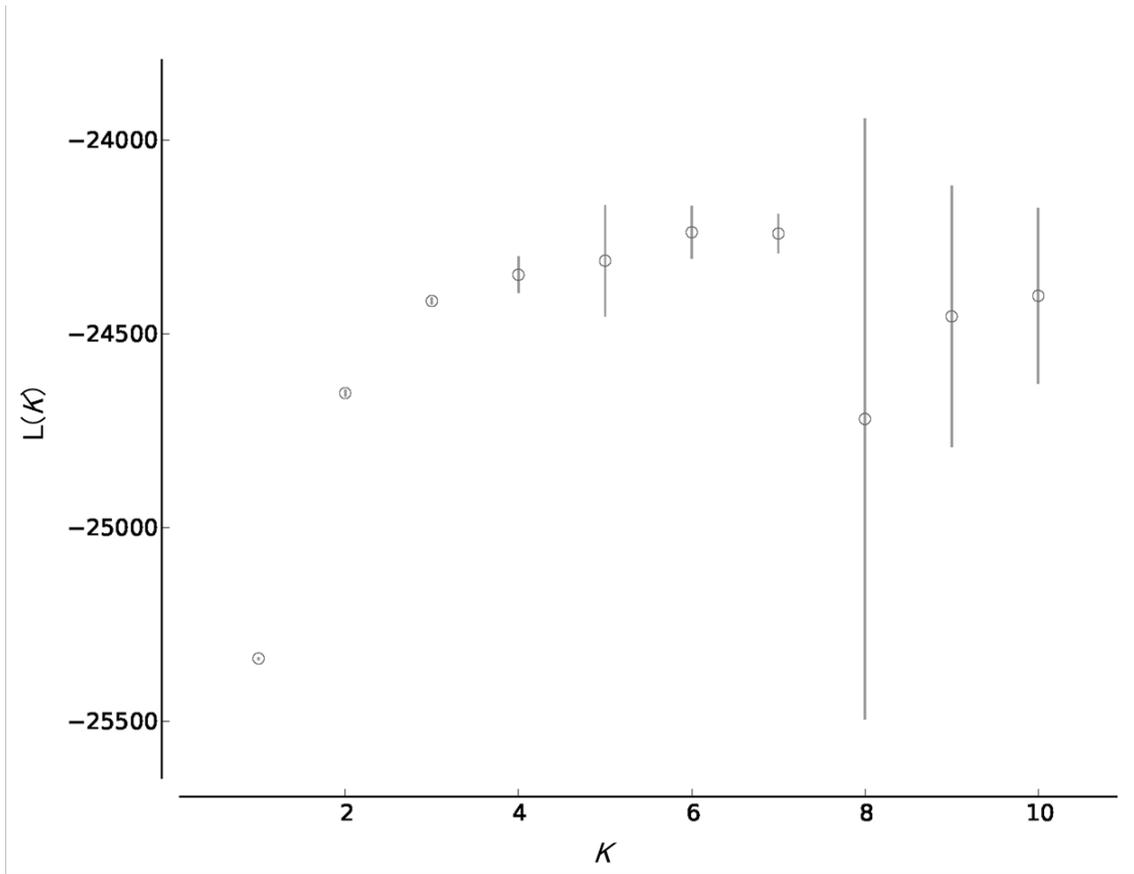


Fig. S2 Change in the mean \pm standard deviation of log probability of the data, $L(K)$, with increasing number of clusters (K), obtained using STRUCTURE analysis of the 17 natural populations of *Thujopsis dolabrata* var. *hondae*

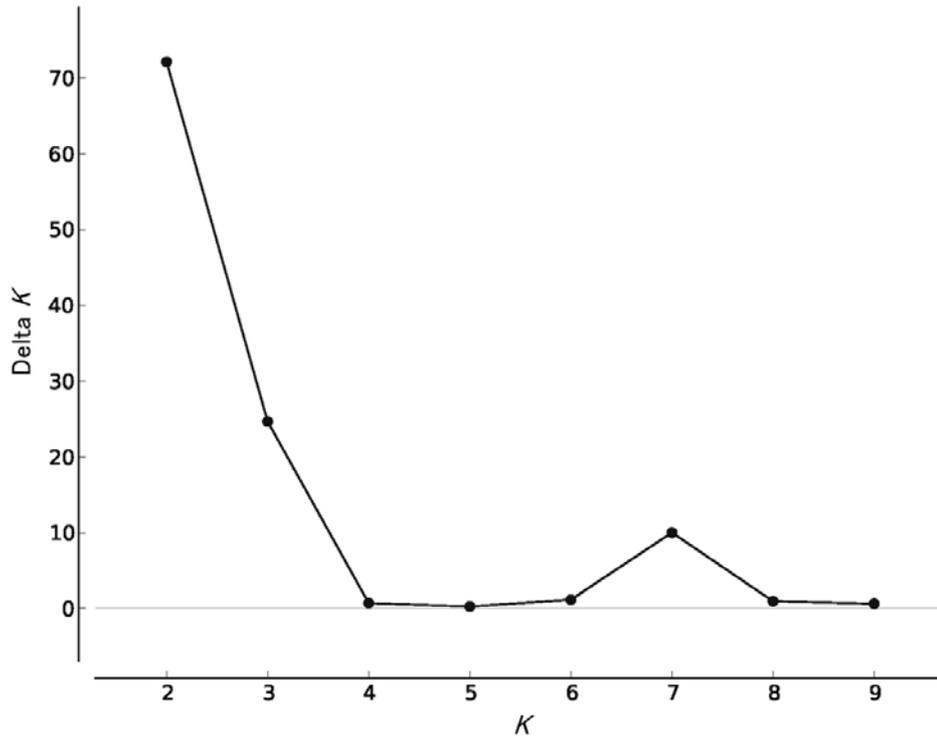


Fig. S3 Change in delta K with increasing number of clusters (K), obtained using STRUCTURE analysis of the 17 natural populations of *Thujopsis dolabrata* var. *hondae*