

# HYPOTHESIS

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## CAN CYSTEINE DIRECT TYROSINE IN SIGNAL TRANSDUCTION FOR ENVIRONMENT-ORIENTED GENE CONTROL?

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### ABSTRACT

Signals are transduced from the cell surface to the nucleus through phosphorylation and dephosphorylation chain reactions of cellular proteins at tyrosine and serine/threonine. Recent evidence suggests that the signal generated through the protein modification at cysteine by oxidation/reduction crosstalks to the protein phosphorylation/dephosphorylation-linked one. I propose that the cysteine-oriented signal potentially directs the tyrosine-oriented one and this mechanism underlies the environment-oriented control of internal signaling for gene expression.

**Key Words:** Signal transduction, Oxidative stress, Protein tyrosine kinase, Reactive oxygen intermediate, Redox, Heavy metal

### INTRODUCTION

Living organisms are created and function through signal exchange between individual genes in the cells at distinct differentiation stages. Signal exchange can occur between any two different genes to bring about their expression, promotion or suppression, with proteins and other organic and inorganic molecules as the messengers. The signals are transduced from cell-surface receptors to the cell nucleus through chain reactions of these messenger molecules, similar to serial on/off switches. These apparently internal, closed reactions for signal transduction are likely to be affected by environmental stresses, which externally control cellular functions. Little is known about the environment-linked control of gene actions at the molecular level. I will discuss the potentially crucial role of oxidization/reduction (redox)-linked protein cysteine modification in environment-oriented gene control.

### INTERNAL SIGNAL TRANSDUCTION FOR GENE CONTROL

The ligand-mediated receptor crosslinkage, which depends on the complementarity of the structures of ligands and receptors, works as the first on-switch for the intracellular signal transduction.<sup>1)</sup> This initial event is followed by forced interaction of intracellular compartments of the receptors and molecules that intracellularly associate with the receptors. The receptors may work by themselves as protein tyrosine (or serine/threonine) kinases (PTKs) or

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phosphotyrosine (or phosphoserine/phosphothreonine) phosphatases (PTPases), or associate intracellularly with non-receptor type PTKs. Once these PTKs and PTPases are functionally mobilized or activated by receptor crosslinkage, they phosphorylate or dephosphorylate substrate proteins including other PTK molecules at tyrosine. This might cause conformational change in the substrate molecules, thereby regulating their functions. Protein phosphorylation and dephosphorylation at tyrosine also mediate association or dissociation of the two molecules that bear phosphorylated or dephosphorylated tyrosine and the specific acceptor domain for the phosphorylated tyrosine, named src homology-2 (SH2).<sup>2-8</sup> Activation of tyrosine kinases, which should occur after receptor crosslinkage, is frequently followed by activation of serine/threonine kinases such as protein kinase C (PKC)<sup>9,10</sup> and mitogen-activated protein kinases (MAPKs).<sup>11,12</sup> Phosphorylation and dephosphorylation of signal elements finally control the activities of transcription factors and cell cycle regulating elements.

### OXIDATIVE STRESS AND SIGNAL TRANSDUCTION

Interaction between cell surface receptors and their ligands, which initiates the intracellular signal delivery, is principally an internal event, except when the ligands are derived from the environment. Animals are, however subject to a number of environmental forces or stresses such as oxygen/oxidants, heat, ultraviolet rays, heavy metals, food and microorganisms (Fig. 1). Animal cells use oxygen by stepwisely reducing it to produce ATP, and a number of reactive oxygen intermediates (ROIs) are generated during this process. Many of the environmental stresses on the animals become oxidative stress because they promote generation of ROIs and related metabolites. For example, large amounts of ROIs are produced in phagocytes that have ingested invaders from the environment. This suggests that the internally regulated signal delivery may be modified by the oxidative stress or ROIs. The oxidative stress and ROIs are known to directly affect proteins, lipids and DNA for molecular damage.<sup>13</sup> Recent studies have, however, revealed that different types of oxidative stress, caused by oxidants, nitric oxide-generating agents, alkylating agents, ultraviolet rays and heavy metals, all share in promoting or inhibiting the tyrosine phosphorylation of cellular proteins<sup>14-22</sup> (Table 1). This suggests that the signal transduction elements are the target of the oxidative stress for modification.

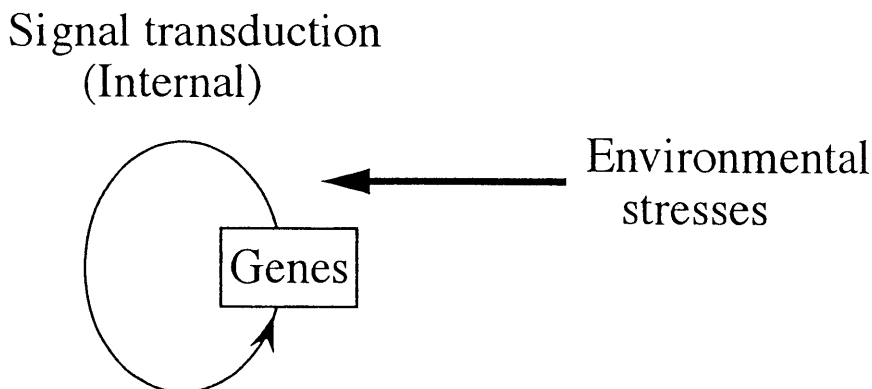


Fig. 1. Internally delivered signals that regulate expression and amplification of genes are subject to control by environmental stresses.

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Table 1. Recent reports on cysteine-oriented regulation of signal transduction.

Event	Agent	Suggested mechanism	Ref.
Promotion of protein tyrosine phosphorylation	Oxidant	Inactivation of PTPase	14, 15
	Heavy metal, NO	Modulation of PTPase activity	16, 17
	Heavy metal, MAA	Cell surface triggering	18
		Promotion of ligand-dependent signal	19
Inhibition of protein tyrosine phosphorylation	Oxidant	NS	20–22
Activation of receptor PTK (Ltk in ER)	Oxidant/IAA	Aggregation of Ltk by S-S bond	24
Activation of nonreceptor PTK (c-Src, Lck, c-Abl)	Ultraviolet ray	NS	49
	IR	NS	50
	Oxidant	NS	51, 52
	NO	Promotion of PTPase activity	17
	Heavy metal	Aggregation of cell surface receptors by S-Hg-S bond	25–27
Inactivation of nonreceptor PTK	Heavy metal	SH-modification of Src	54, 55
	Harbimycin, NEM	SH-modification of Src	45, 46
Inactivation of nonreceptor PTK	NEM	blocking interaction between CD4 and Lck	47
	Heavy metal/oxidant	SH-modification of PTPase	42–44
Inactivation of PKC	NO	SH-modification of PKC	48
Activation/inactivation of transcription factors (AP-1, NF- $\kappa$ B, steroid receptor)	Oxidant	Activation in vivo	60–62
		Inhibition in vitro	63
	Thioredoxin/Ref-1	Activation through reduction of S-S bond	67–69

PTK: protein tyrosine phosphatase	IAA: iodoacetamide
PTPase: phosphotyrosine phosphatase	NEM: N-ethylmaleimide
PKC: protein kinase C	IR: ionizing radiation
NO: nitrogen oxide-producing chemical	ER: endoplasmic reticulum
MAA: monoiodoacetic acid	NS: not specifically explained

### CYSTEINE-ORIENTED VS. TYROSINE-ORIENTED PROTEIN MODIFICATION

Among several reactive residues on proteins, thiol (SH)-groups of cysteines are the central target of the redox reaction.<sup>23)</sup> The cysteine-oriented redox reaction on protein molecules alters their status and structure through two mechanisms. First, it crosslinks two or more peptides with intermolecular S-S bonds. Second, it induces conformational changes in the molecules with intramolecular S-S bonds for adjacent SH-groups on the molecules. Both types of redox-linked protein modification are comparable to those of a tyrosine-mediated one, resulting from inter- and intra-molecular bonding between the phosphorylated tyrosine and its acceptor sequence on

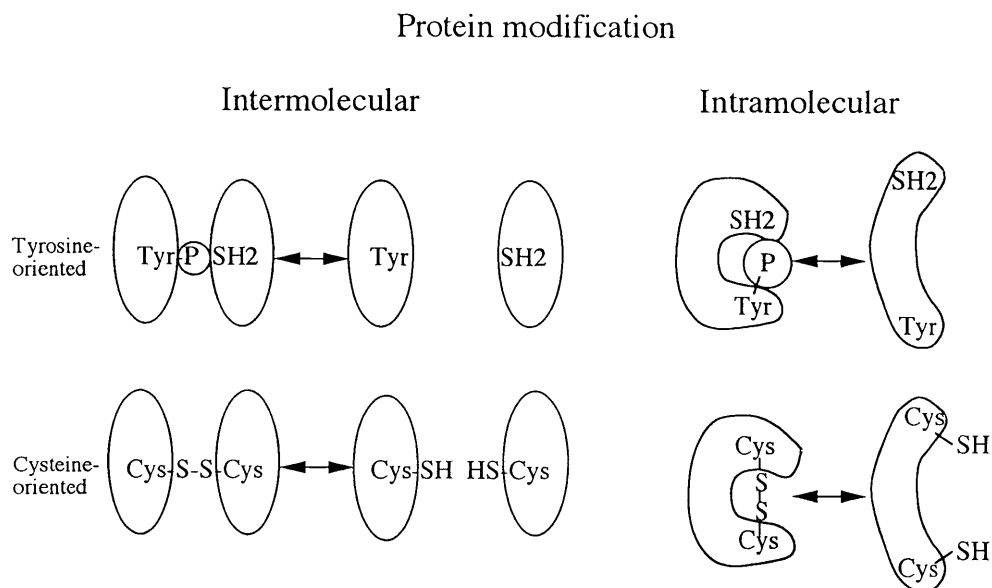


Fig. 2. Cysteine-oriented vs. tyrosine-oriented conformational changes of signal elements.

the SH2 domain<sup>4,8)</sup> (Fig. 2). The redox-linked cysteine-oriented protein modification therefore, probably fulfills the condition for working as an on/off switch of the signal transduction, just as the tyrosine-oriented modification does.

#### CYSTEINE-ORIENTED RECEPTOR CROSSLINKAGE AND ACTIVATION OF PTKs

Ben-Neriah and his colleagues<sup>24)</sup> reported that intracellularly inoculated oxidants and alkylating agents induced aggregation and activation of the receptor type PTKs in the endoplasmic reticulum. This was a result of the formation of intermolecular S-S bonds between the two PTK molecules, potentially catalyzed by protein disulfide isomerase. Independently, we provided evidence that the redox reaction might affect cell surface receptors for initiating signal transduction from the cell surface to the nucleus<sup>18,25-27)</sup> (Table 1). The study was carried out using  $\text{Hg}^{2+}$ , a well-known SH-reagent with an extraordinarily high association constant to cysteine SH-groups and able to replace the S-S bond with the functionally equivalent S-Hg-S bond.<sup>28-32)</sup> We showed that exposure of murine T lymphocytes to  $\text{Hg}^{2+}$  in vitro induces high grade phosphorylation of cellular proteins at tyrosine, and activation of Lck kinase, a nonreceptor PTK of the Src family. These events were accompanied by aggregation of a number of cell surface proteins including CD3 (a signal transducing element in the T cell receptor complex), CD4, CD45 (receptor type tyrosine phosphatase) and Thy-1 (glycosylphosphatidylinositol (GPI)-anchored cell membrane protein).<sup>25)</sup> Both tyrosine phosphorylation promotion of cellular proteins and aggregation of cell surface proteins seemed to be mediated by the reaction between SH-groups on cell surface proteins and  $\text{Hg}^{2+}$ , because reducing or SH-group-donating reagents neutralized the  $\text{Hg}^{2+}$  action. Evidence was further produced that the primary target of  $\text{Hg}^{2+}$  for promotion of protein tyrosine phosphorylation includes cell surface GPI-anchored proteins.<sup>27)</sup> GPI anchored proteins are

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known to associate with nonreceptor PTKs such as Lck and Src kinases across plasma membrane,<sup>33)</sup> and to transmit signals for cell activation.<sup>34–36)</sup> In earlier experiments, we co-cross-linked both T cell receptors and Thy-1 as a GPI-anchored protein with appropriate antibodies, and found that the two cell membrane proteins worked synergistically to deliver a high grade signal for extensive protein tyrosine phosphorylation and activation of Lck kinase.<sup>36,37)</sup> Based on these observations, we speculated that heterogeneous crosslinkages of multiple transmembrane and GPI-anchored proteins through S-Hg-S bonds are capable of generating extraordinarily high grade signals to activate PTKs.<sup>25)</sup> Correspondingly, Murakami et al.<sup>38)</sup> recently showed that crosslinkage of interleukin-6 receptors (IL-6R) by IL-6 as an inflammatory cytokine induces dimerization of gp130 (the signal transducing element of IL-6R) through an S-S bond, and this dimerization is associated with activation of nonreceptor PTKs. By analyzing the mechanism of activation of the *ret* proto-oncogene<sup>39,40)</sup> by multiple endocrine neoplasia 2A mutations, Asai et al.<sup>41)</sup> also showed that dimerization of the Ret kinase proteins through a *ret* mutation-linked S-S bond underlies their constitutive activation in the neoplasm. These reports support the view that dimerization or aggregation of cell surface receptor proteins through S-S bonds (or S-S bonds-replacing S-X-S bonds) is widely involved physiologically and pathologically in the initial mechanism of cell signaling.

PTPases AND PTKs AS THE TARGETS OF  
CYSTEINE-ORIENTED MODIFICATION

The S-S bond or S-Hg-S bond-mediated receptor crosslinkage may promote mutual interaction of intracellular elements that associate with the intracellular compartments of the cell membrane receptors for activation or regulation. The receptor crosslinkage, whether or not it is mediated by redox mechanism, might also promote production of ROIs in the cell, possibly through a protein phosphorylation-dependent signaling. These ROIs could in turn affect the cell surface receptors and intracellular signal elements secondarily for the SH-modification as a potential signal amplifying mechanism. Thus, intracellular signal elements could be the targets of both intracellularly introduced oxidants and the secondarily generated ROIs (Table 1). It was first suggested that this signal pathway operated on PTPases,<sup>15,42–44)</sup> known to be sensitive to SH-reagents *in vitro* for inactivation.<sup>43)</sup> Herbimycin A and some alkylating reagents have also been shown to directly affect Src and Lck kinases for inactivation.<sup>45,46)</sup> Furthermore, alkylating agents might inhibit the kinase activity by blocking the interaction between CD4 and Lck,<sup>47)</sup> and nitric oxide or nitric oxide-generating agents could inactivate PKC *in vitro*.<sup>48)</sup> These observations raised the possibility that nonreceptor PTKs and PKC are the direct target of the cysteine-oriented regulation. On the other hand, ultraviolet rays,<sup>49)</sup> ionic radiation,<sup>50)</sup> oxidants<sup>51,52)</sup> and anoxia,<sup>53)</sup> when live cells are subjected to them, have been shown to upregulate the c-Src, Lck and c-Abl kinases. The molecular mechanism of the kinase activation by oxidizing agents *in vivo*,<sup>49–53)</sup> and the contrast with the inactivation by SH-reagents *in vitro*,<sup>45,46)</sup> have not been well-explained. Recently, we demonstrated for the first time that the catalytic activity of Src kinase is upregulated by the SH-modification of the kinase protein with Hg<sup>2+</sup> in a cell-free system.<sup>54,55)</sup> Correspondingly, Veillette et al.<sup>56)</sup> reported that Lck kinase contains conserved cysteines crucial for enzymatic activity. Taken together, we speculate that the Src kinase bears the target structure for appropriate SH-modification to either upregulate or downregulate the kinase activity depending on the conditions.

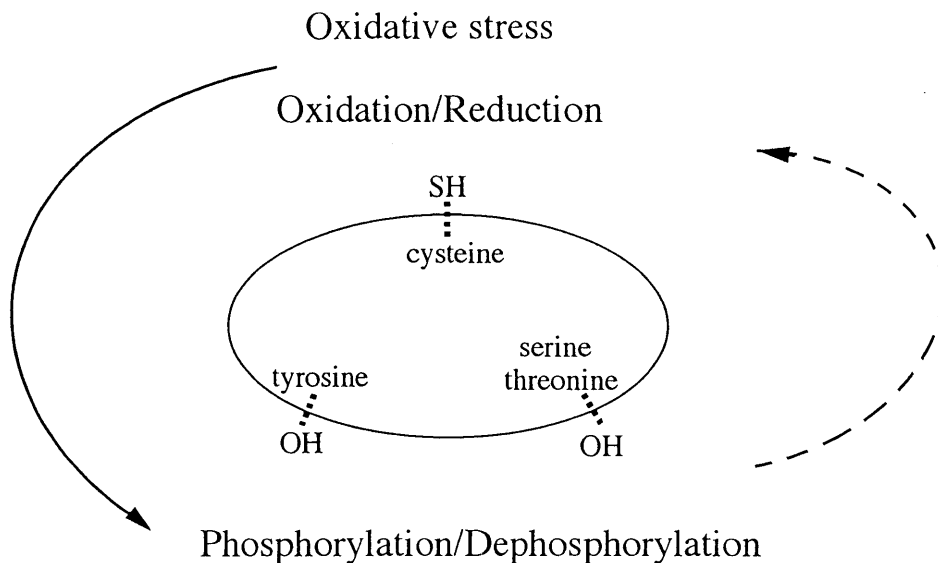


Fig. 3. Cysteine can direct tyrosine and serine/threonine on proteins in signal transduction.

#### RELATION BETWEEN CYSTEINE-ORIENTED AND TYROSINE-ORIENTED SIGNALS

For the  $\text{Hg}^{2+}$  model, we examined the relation of the redox-mediated cysteine-oriented regulation to the phosphorylation/dephosphorylation-dependent tyrosine-oriented control of Src kinase activity. Interestingly, activation of Src kinase by  $\text{Hg}^{2+}$  occurred independently of the regulation through phosphorylation/dephosphorylation of C-terminal Tyr-527 (the known regulatory site<sup>2-5</sup>) of c-Src kinase, selectively promoting Tyr-416 (the known autophosphorylation site) phosphorylation without definite change in the phosphorylation level of Y527.<sup>54,55,57</sup> We therefore proposed that the cysteine-oriented regulation could be upstream from the tyrosine-oriented one.<sup>54,55</sup> In other words, the cysteine may direct the tyrosine for cellular signal transduction (Fig. 3). This hypothetical principle underlies the action mechanism of the environmental stress controlling internal signal transduction.

#### THE SECOND WAVE OF THE CYSTEINE-ORIENTED SIGNAL

The signal initially triggered by the cysteine-oriented mechanism should further drive the tyrosine-oriented and the serine/threonine-oriented signal pathways. They include the phosphorylation and activation of MAPK<sup>11,12</sup> and stress-activated protein kinases (SAPK/JNK)<sup>58,59</sup> of the MAPK family, which phosphorylates and regulates transcription factors. Change in the activity of transcription factors is therefore expected following the oxidative stress-induced activation of PTKs. It has indeed been reported that transcription factors such as NF- $\kappa$ B could be activated in association with changes in the cellular redox status caused by oxidative stress.<sup>60-62</sup> The DNA binding activity of NF- $\kappa$ B was shown, however, to be inhibited by treatment with oxidizing and alkylating agents in vitro.<sup>63</sup> The stress-provoked signals induce production of stress proteins, such as superoxide dismutase<sup>64</sup> and thioredoxin,<sup>65,66</sup> which protect the cells from overly

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high oxidative stress. The stress proteins could also provoke the second wave of redox signal that regulates the action of transcription factors, such as AP-1,<sup>67)</sup> NF- $\kappa$ B<sup>68)</sup> and corticosteroid receptors.<sup>69)</sup> This potentially switches the inactivated form bearing the S-S bond on conserved cysteines to the activated form with reduced SH-groups.

In concert with the sophisticated phosphorylation/dephosphorylation-dependent regulatory mechanism, the first and second stages of redox-linked control could ultimately decide the levels of gene expression and expansion for physiological and pathological cell growth and death.<sup>25,70-72)</sup>

The view presented in this paper is mainly based on results of our recent study, obtained in collaboration with Drs. M. Pu, A.A. Akhand, M. Kato and K. Ohkusu in the Department of Immunology and Dr. M. Hamaguchi in the Laboratory of Molecular Pathology, Research Institute of Disease Mechanism and Control, Nagoya University School of Medicine.

## CONCLUDING REMARKS

In conclusion, I propose that the cysteine-oriented conformational change of signal elements works as a molecular on/off switch for signal transduction. This is an alternate to the known tyrosine-oriented or serine/threonine-oriented modification, and may even direct these for environmentally regulated internal signal transduction. In other words, the cysteines on proteins might be an open window to receive the "wind" of the environment, which promotes or regulates cellular functions.

## REFERENCES

- 1) Metzger, H.: Transmembrane signaling: the joy of aggregation. *J. Immunol.*, 149, 1477-1487 (1992).
- 2) Cooper, J.A. and King, C.S.: Dephosphorylation or antibody binding to the carboxy terminus stimulates pp60<sup>c-src</sup>. *Mol. Cell. Biol.*, 6, 4467-4477 (1986).
- 3) Cantley, L.C., Auger, K.R., Carpenter, C., Duckworth, B., Graziani, A., Kapeller, R. and Soltoff, S.: Oncogenes and signal transduction. *Cell*, 64, 281-302 (1991).
- 4) Koch, C.A., Anderson, D., Moran, M.F., Ellis, C. and Pawson, T.: SH2 and SH3 domains: elements that control interactions of cytoplasmic signaling proteins. *Science*, 252, 668-674 (1991).
- 5) Thomas, J.E., Soriano, P. and Brugge, J.S.: Phosphorylation of c-src on tyrosine 527 by another kinase. *Science*, 254, 568-571 (1991).
- 6) Weiss, A.: T cell antigen receptor signal transduction: a tale of tails and cytoplasmic protein-tyrosine kinases. *Cell*, 73, 209-212 (1993).
- 7) Cooper, J.A. and Howell, B.: The when and how of Src regulation. *Cell*, 73, 1051-1054 (1993).
- 8) Pawson, T.: Protein modules and signaling networks. *Nature*, 373, 573-580 (1995).
- 9) Nishizuka, Y.: The molecular heterogeneity of protein kinase C and its implications for cellular regulation. *Nature*, 334, 661-665, (1988).
- 10) Iwamoto, T., Hagiwara, M., Hidaka, H., Isomura, T., Kioussis, D. and Nakashima, I.: Accelerated proliferation and interleukin-2 production of thymocytes by stimulation of soluble anti-CD3 monoclonal antibody in transgenic mice carrying a rabbit protein kinase C. *J. Biol. Chem.*, 267, 18644-18648, 1992.
- 11) Kyriakis, J.M., App, H., Zhang, X.F., Banerjee, P., Brautigan, D.L. and Rapp, U.R.: Raf-1 activates MAP kinase-kinase. *Nature*, 358, 417-421 (1992).
- 12) Rozakis-Adcock, M., Fernley, R., Wade, J., Pawson, T. and Bowtell, D.: The SH2 and SH3 domains of mammalian Grb2 couple the EGF receptor to the Ras activator mSos 1. *Nature*, 363, 83-85 (1993).
- 13) Fridovich, I.: The biology of oxygen radicals. *Science*, 201, 875-879 (1978).
- 14) Heffetz, D. and Zick, Y.: H<sub>2</sub>O<sub>2</sub> potentiates phosphorylation of novel putative substrates for the insulin receptor kinase in intact Fao cells. *J. Biol. Chem.*, 264, 10126-10132 (1989).

- 15) Staal, F., Roederer, M., Anderson, M.T. and Herzenberg, L.A.: Redox regulation of NF- $\kappa$ B activation and therefore of HIV expression. In *Molecular basis of immune responses*, edited by Nariuchi, H., et al., pp.89–102 (1993), Academic press, Tokyo, Japan.
- 16) Lander, H.M., Levine, D.M. and Novogrodsky, A.: Stress stimuli-induced lymphocyte activation. *Cell. Immunol.*, 145, 146–155 (1992).
- 17) Lander, H.M., Sehajpal, P., Levine, D.M. and Novogrodsky, A.: Activation of human peripheral blood mononuclear cells by nitric oxide-generating compounds. *J. Immunol.*, 150, 1509–1516 (1993).
- 18) Rahman, S.M.J., Pu, M., Hamaguchi, M., Iwamoto, T., Isobe, K. and Nakashima, I.: Redox-linked ligand-independent cell surface triggering for extensive protein tyrosine phosphorylation. *FEBS Lett.*, 317, 35–38 (1993).
- 19) Ma, L., Pu, M., Yi, H., Akhand, A.A., Obata, N., Ohkusu, K., Kato, M., Iwamoto, T., Isobe, K., Hamaguchi, M. and Nakashima, I.: Multiphasic modulation of signal transduction into T lymphocytes by monoiodoacetic acid as a sulfhydryl reagent. *J. Cell. Biochem.*, 59, 33–41 (1995).
- 20) Rozsnyay, Z., Sarmay, G. and Gergely, J.: Phenylarsine oxide (PAO) blocks antigen receptor-induced calcium response and tyrosine phosphorylation of a distinct group of proteins. *FEBS Lett.*, 337, 197–205 (1993).
- 21) Anel, A., Mescher, M.F. and Kleinfeld, A.M.: Activated adhesion of CTL to MHC class I but not to fibronectin is inhibited by cis unsaturated fatty acids and phenylarsine oxide. *J. Immunol.*, 155, 1039-1-46 (1995).
- 22) Flescher, E., Ledbetter, J.A., Schieven, G.L., Vela-Roch, N., Fossum, D., Dang, H., Ogawa, N. and Talal, N.: Longitudinal exposure of human T lymphocytes to weak oxidative stress suppresses transmembrane and nuclear signal transduction. *J. Immunol.*, 153, 4880–4890 (1994).
- 23) Ziegler, D.M.: Role of reversible oxidation-reduction of enzyme thiols-disulfides in metabolic regulation. *Ann. Rev. Biochem.* 54, 305–329 (1985).
- 24) Bauskin, A.R., Alkalay, I. and Ben-Neriah, Y.: Redox regulation of a protein tyrosine kinase in the endoplasmic reticulum. *Cell*, 66, 685–696 (1991).
- 25) Nakashima, I., Pu, M., Nishizaki, N., Rosila, I., Ma, L., Katano, Y., Ohkusu, K., Rahman, S.M.J., Isobe, K., Hamaguchi, M. and Saga, K.: Redox mechanism as alternative to ligand binding for receptor activation delivering dysregulated cellular signals. *J. Immunol.*, 152, 1064–1071 (1994).
- 26) Katano, Y., Pu, M., Akhand, A.A., Hamaguchi, M., Koga, Y., Isobe, K., Fukuda, Y., Hayakawa, T. and Nakashima, I.: Evidence of redox-linked signaling for producing a giant signal complex. *J. Cell. Biochem.*, 57, 432–439 (1995).
- 27) Pu, M., Ma, L., Ohkusu, K., Isobe, K., Taguchi, R., Ikezawa, H., Hamaguchi, M. and Nakashima, I.: Direct evidence of involvement of glycosylphosphatidylinositol-anchored proteins in the heavy metal-mediated signal delivery into T lymphocytes. *FEBS Lett.*, 361, 295–298 (1995).
- 28) Stricks, W. and Kolthoff, I.M.: Reactions between mercuric mercury and cysteine and glutathione. Apparent dissociation constants, heats and entropies of formation of various forms of mercuric mercapto-cysteine and -glutathione. *J. Am. Chem. Soc.*, 75, 5673–5680 (1953).
- 29) Simpson, R.B.: Association constraints of methylmercury with sulfhydryl and other bases. *J. Am. Chem. Soc.*, 83, 4711–4717 (1961).
- 30) Steer, M.L., Tal, N. and Levitzki, A.: The role of sulfhydryl groups in the action and structure of mammalian  $\alpha$ -amilase. *Biochim. Biophys. Acta*, 334, 389–397 (1974).
- 31) Sperling, R., Burstein, Y. and Steinberg, I.Z.: Selective reduction and mercuration of cysteine IV-V in bovine pancreatic ribonuclease. *Biochemistry*, 8, 3810–3820 (1969).
- 32) Utschig, L.M., Bryson, J.W. and O'Halloran, T.V.: Mercury-199 NMR of the metal receptor site in MerR and its protein-DNA complex. *Science*, 268, 380–385 (1995).
- 33) Stefanova, I., Horeji, V., Ansoategui, I.J., Knapp, W. and Stockinger, H.: GPI-anchored cell surface molecules complex to protein tyrosine kinases. *Science*, 254, 1016–1021 (1993).
- 34) Kroczeck, R.A., Gunter, K.C., Germain, R.N. and Shevach, E.M.: Thy-1 functions as a signal transduction molecule in T lymphocytes and transfected B lymphocytes. *Nature*, 322, 181–184 (1986).
- 35) Rahman, S.M.J., Pu, M., Zhang, Y.-H., Hamaguchi, M., Iwamoto, T., Taguchi, R., Ikezawa, H., Isobe, K., Yoshida, T. and Nakashima, I.: Delivery of accessory signal for cell activation by exogenous phosphatidylinositol-specific phospholipase C. *FEBS Lett.*, 303, 193–196 (1992).
- 36) Nakashima, I., Zhang, Y.-H., Rahman, S.M.J., Yoshida, T., Isobe, K., Ding, L., Iwamoto, T., Hamaguchi, M., Ikezawa, H. and Taguchi, R.: Evidence of synergy between Thy-1 and CD3/TCR complex in signal deliver to murine thymocytes for cell death. *J. Immunol.*, 147, 1153–1162 (1991).



## OXIDATIVE STRESS AND CELL SIGNALING

- 37) Nakashima, I., Pu, M., Hamaguchi, M., Iwamoto, T., Rahman, S.M.J., Zhang, Y.-H., Kato, M., Ohkusu, K., Katano, Y., Yoshida, T., Koga, Y., Isobe, K. and Nagase, F.: Pathway of signal delivery to murine thymocytes triggered by co-crosslinking CD3 and Thy-1 for cellular DNA fragmentation and growth inhibition. *J. Immunol.*, 151, 3511–3520 (1993).
- 38) Murakami, M. et al.: IL-6-induced homodimerization of gp130 and associated activation of a tyrosine kinase. *Science*, 260, 1808–1810 (1993).
- 39) Takahashi, M., Asai, N., Iwashita, T., Isomura, T., Miyazaki, K. and Matsuyama, M.: Characterization of ret proto-oncogene products expressed in mouse L cells. *Oncogene*, 8, 2925–2929 (1983).
- 40) Iwamoto, T., Taniguchi, M., Asai, N., Ohkusu, K., Nakashima, I. and Takahashi, M.: cDNA cloning of mouse *ret* proto-oncogene and its sequence similarity to the cadherin superfamily. *Oncogene*, 8, 1087–1091 (1993).
- 41) Asai, N., Iwashita, T., Matsuyama, M. and Takahashi, M.: Mechanism of activation of the ret proto-oncogene by multiple endocrine neoplasia 2A mutations. *Mol. Cell. Biol.*, 15, 1613–1619 (1995).
- 42) Garcia-Morales, P., Minami, Y., Luong, E., Klausner, R.D., and Samelson, L.E.: Tyrosine phosphorylation in T cells is regulated by phosphatase activity: studies with phenylarsine oxide. *Proc. Nat. Acad. Sci. USA*, 87, 9255–9259 (1990).
- 43) Gerge, R.J. and Parker, C.W.: Preliminary characterization of phosphotyrosine phosphatase activities in human peripheral blood lymphocytes: identification of CD45 as a phosphotyrosine phosphatase. *J. Cell. Biochem.*, 42, 71–81 (1990).
- 44) Atkinson, T.P., Lee, C.-W., Rhee, S.G. and Hohman, R.J.: Orthovanadate induces translocation of phospholipase C-1 and -2 in permeabilized mast cells. *J. Immunol.*, 151, 1448–1455 (1993).
- 45) Uehara, Y., Fukazawa, H., Murakami, Y. and Mizuno, S.: Irreversible inhibition of v-src tyrosine kinase activity by herbimycin A and its abrogation by sulfhydryl compounds. *Biochem. Biophys. Res. Commun.*, 163, 803–809 (1989).
- 46) Fukazawa, H., Uehara, Y., Murakami, Y., Mizuno, S., Hamada, M. and Takeuchi, T.: Labeling of v-Src and BCR-ABL tyrosine kinases with [<sup>14</sup>C]herbimycin A and its use in the elucidation of the kinase inactivation mechanism. *FEBS Lett.*, 340, 155–158 (1994).
- 47) Kanner, S.B., Kavanagh, T.J., Grossmann, A., Hu, S.-L., Bolen, J.B., Rabinovitch, P.S. and Ledbetter, J.A.: Sulfhydryl oxidation down-regulates T-cell signaling and inhibits tyrosine phosphorylation of phospholipase C $\gamma$ 1. *Proc. Nat. Acad. Sci. USA*, 89, 300–304 (1992).
- 48) Gopalakrishna, R., Chen, Z.H. and Gundimeda, U.: Nitric oxide and nitric oxide-generating agents induces a reversible inactivation of protein kinase C activity and phorbol ester binding. *J. Biol. Chem.*, 268, 27180–27185 (1993).
- 49) Devary, V., Gottlieb, R.A., Smeal, T. and Karin, M.: The mammalian ultraviolet response is triggered by activation of Src tyrosine kinase. *Cell*, 71, 1081–1091 (1992).
- 50) Kharbanda, S., Ren, R., Pandey, P., Shafman, T.D., Feller, S.M., Weichselbaum, R.R. and Kufe, D.W.: Activation of the c-Abl tyrosine kinase in the stress response to DNA-damaging agents. *Nature*, 376, 785–788 (1995).
- 51) Nakamura, K., Hori, T., Sato, N., Sugie, K., Kawakami, T. and Yodoi, J.: Redox regulation of a src family protein tyrosine kinase p56<sup>lck</sup> in T cells. *Oncogene*, 8, 3133–3139 (1993).
- 52) Hardwick, J.S. and Sefton, B.M.: Activation of the Lck tyrosine protein kinase by hydrogen peroxide requires the phosphorylation of Tyr-394. *Proc. Nat. Acad. Sci. USA*, 92, 4527–4531 (1995).
- 53) Mukhopadhyay, D., Tsiokas, L., Zhou, X.-M., Foster, D., Brugge, J.S. and Sukhatme, V.P.: Hypoxic induction of human vascular endothelial growth factor expression through c-Src activation. *Nature*, 375, 577–581 (1995).
- 54) Pu, M., Hamaguchi, M., Sabe, S., Koike, T. and Nakashima, I.: Carboxyl-terminal tyrosine-independent redox-linked activation of Src kinases in vitro. submitted.
- 55) Nakashima, I., Pu, M., Hamaguchi, M., Akhand, A.A. and Kato, M.: Sulfhydryl-based regulation of Src family protein tyrosine kinases for cell growth and death. In *Proceedings of the Jakarta International Cancer Conference*, edited by Berkel, H., Faculty Medicine University Press, Jakarta, in press.
- 56) Veillette, A., Dumont, S. and Fournel, M.: Conserved cysteine residues are critical for the enzymatic function of the lymphocyte-specific tyrosine protein kinase p56<sup>lck</sup>. *J. Biol. Chem.*, 268, 17547–17553 (1993).
- 57) Pu, M., Akhand, A.A., Kato, M., Koike, T., Hamaguchi, M. and Nakashima, I.: Aggregation promotion and activation of Src kinase by mercuric chloride through a redox-linked mechanism. submitted.
- 58) Davis, R.J.: MAPKs: new JNK expands the group. *Trends Biochem. Sci.*, 19, 470–473 (1994).
- 59) Derijard, B., Raingeaud, J., Barret, T., Wu, I.H., Han, J., Ulvitch, R.J. and Davis, R.J.: Independent human MAP-kinase signal transduction pathways defined by MEK and MKK isoforms. *Science*, 267, 682–685 (1995).

- 60) Storz, G., Tartaglla, L.A. and Ames, B.N.: Transcriptional regulator of oxidative stress-inducible genes: Direct activation by oxidation. *Science*, 248, 189–194 (1990).
- 61) Staal, F.J.T., Roederer, M., Herzenberg, L.A. and Herzenberg, L.A.: Intracellular thiols regulate activation of nuclear factor kB and transcription of human immunodeficiency virus. *Proc. Nat. Acad. Sci. USA*, 87, 9943–9947 (1990).
- 62) Israel, N., Gougerot-Pocidallo, M.-A., Aillet, F. and Virelizier, J.-L.: Redox status of cells influences constitutive or induced NF-kB translocation and HIV monocytic cell lines. *J. Immunol.*, 149, 3386–3393 (1992).
- 63) Toledano, M.B. and Leonard, W.J.: Modulation of transcription factor NF-kB binding activity by oxidation-reduction in vitro. *Proc. Nat. Acad. Sci. USA*, 88, 4328–4332 (1991).
- 64) Fujii, J. and Taniguchi, N.: Phorbol ester induces manganese-superoxide dismutase in tumor necrosis factor-resistant cells. *J. Biol. Chem.*, 266, 23142–23146 (1991).
- 65) Holmgren, A.: Thioredoxin. *Annu. Rev. Biochem.*, 54, 237–271 (1985).
- 66) Hori, K., Katayama, M., Sato, N., Ishii, K., Waga, S. and Yodoi, J.: Neuroprotection by glial cells through adult T cell leukemia-derived factor/human thioredoxin (ADF/TRX). *Brain Res.*, 652, 304–310 (1994).
- 67) Abate, C., Patel, L., Rauskin, F.J.III and Curran, T.: Redox regulation of Fos and Jun DNA-binding activity in vitro. *Science*, 249, 1157–1161 (1990).
- 68) Hayashi, T., Ueno, Y. and Okamoto, T.: Oxido-reductive regulation of nuclear factor kB. Involvement of a cellular catalyst thioredoxin. *J. Biol. Chem.* 268, 11380–11388 (1993).
- 69) Grippo, J.F., Holmgren, A. and Pratt, W.B.: Proof that the endogeneous, heat-stable glucocorticoid receptor-activating factor is thioredoxin. *J. Biol. Chem.*, 260, 93–97 (1984).
- 70) Jiang, Y. and Moller, G.: In vitro effects of HgCl<sub>2</sub> on murine lymphocytes. I. Preferable activation of CD4<sup>+</sup> T cells in a responder strain. *J. Immunol.*, 154, 3138–3146 (1995).
- 71) Sato, N., Iwata, S., Nakamura, K., Hori, T., Mori, K. and Yodoi, J.: Thiol-mediated redox regulation of apoptosis. Possible roles of cellular thiols other than glutathione in T cell apoptosis. *J. Immunol.*, 154, 3194–3203 (1995).
- 72) Fisher, G.J., Datta, S.C., Talwar, H.S., Wang, Z.-Q., Varani, J., Kang, S. and Voorhees, J.J.: Molecular basis of sun-induced premature skin ageing and retinoid antagonism. *Nature*, 379, 335–339 (1996).